

(19) United States

(12) Patent Application Publication (10) Pub. No.: US 2004/0077574 A1

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Apr. 22, 2004 (43) Pub. Date:

(54) MODULATION OF BIOLOGICAL SIGNAL TRANSDUCTION BY RNA INTERFERENCE

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10/444,795 Appl. No.: (21)

(22)Filed: May 23, 2003

Related U.S. Application Data

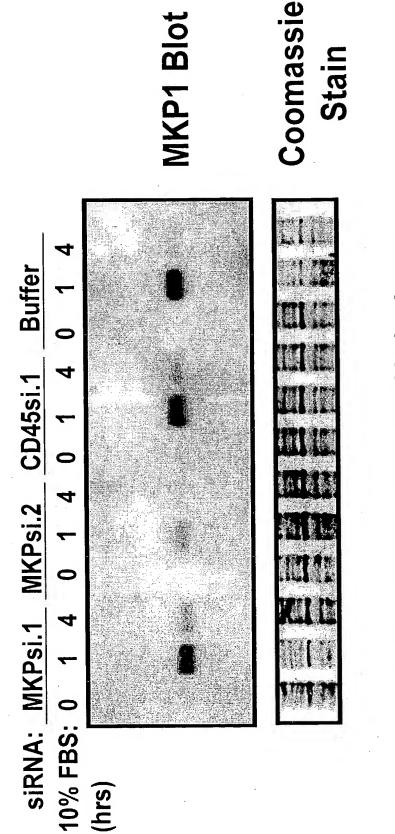
Provisional application No. 60/462,942, filed on Apr. 14, 2003. Provisional application No. 60/383,249, filed on May 23, 2002.

Publication Classification

Int. Cl.⁷ A61K 48/00; C07H 21/02; C12N 15/85

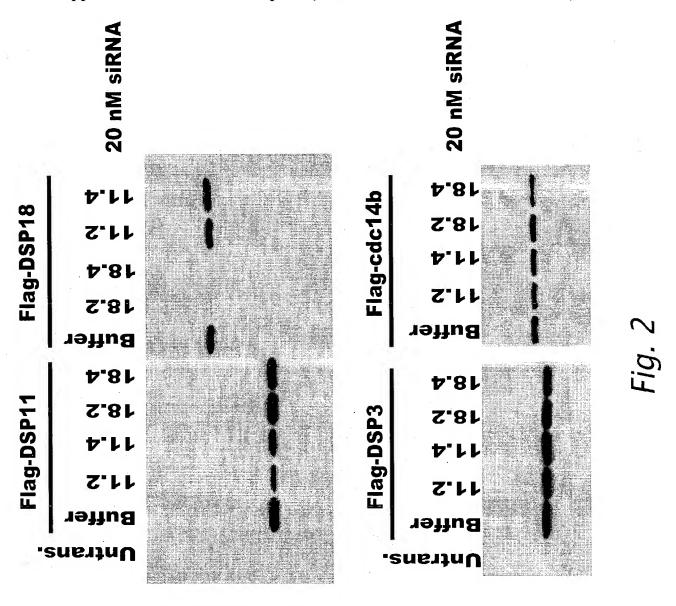
(57)**ABSTRACT**

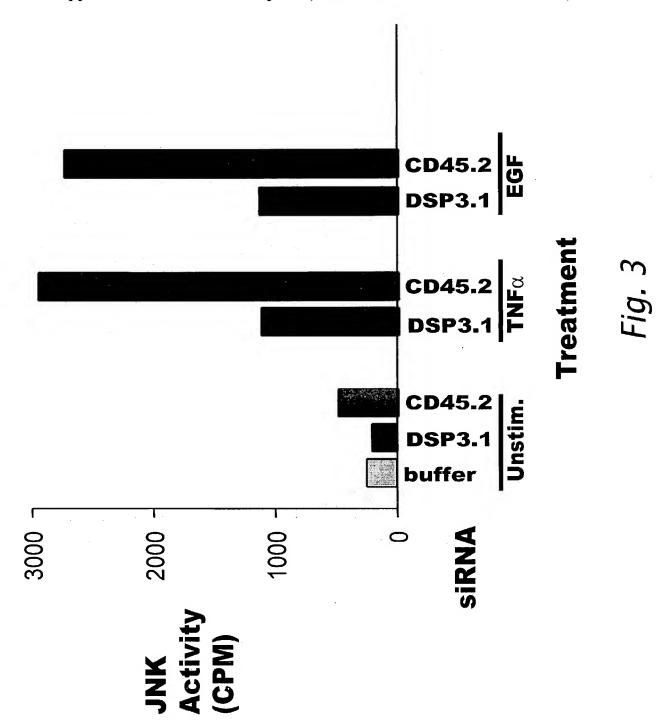
Compositions and methods relating to small interfering RNA (siRNA) polynucleotides are provided as pertains to modulation of biological signal transduction. Shown are siRNA polynucleotides that interfere with expression of members of the protein tyrosine phosphatase (PTP) class of enzymes that mediate signal transduction, and with certain MAP kinase kinases (MKK). In certain preferred embodiments siRNA modulate signal transduction pathways comprising SHP2, cdc14a/b, cdc25A/B/C, KAP, PTP- ϵ , PRL-3, CD45, dual specificity phosphatase-3 (DSP-3), MKK-4, and/or MKK-7. Modulation of PTP-mediated biological signal transduction has uses in diseases associated with defects in cell proliferation, cell differentiation and/or cell survival, such as metabolic disorders (including diabetes and obesity), cancer, autoimmune disease, infectious and inflammatory disorders and other conditions. The invention also provides siRNA polynucleotides that interfere with expression of chemotherapeutic target polypeptides, such as DHFR, thymidylate synthetase, and topoisomerase I.

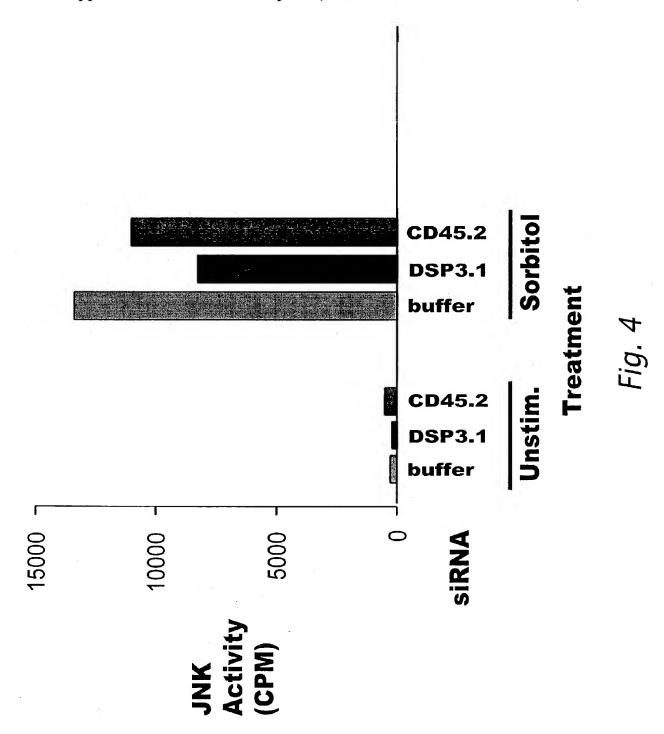


HeLa cells, transfected with siRNA duplexes 24 hr before stimulation with FBS.

Fig.







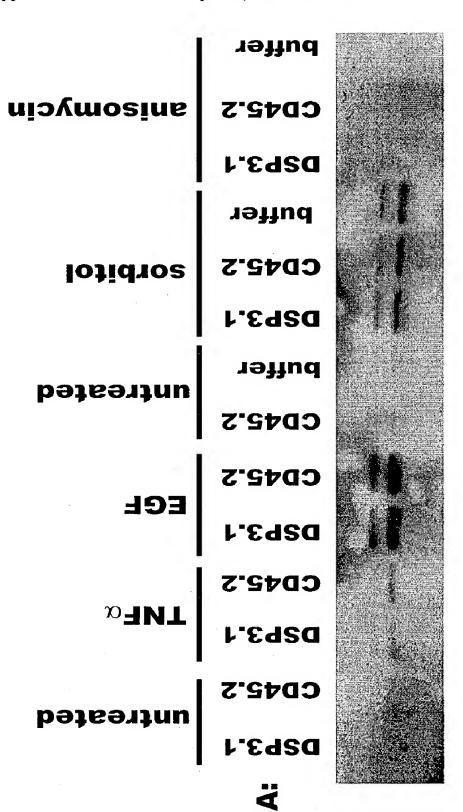


Fig. 5

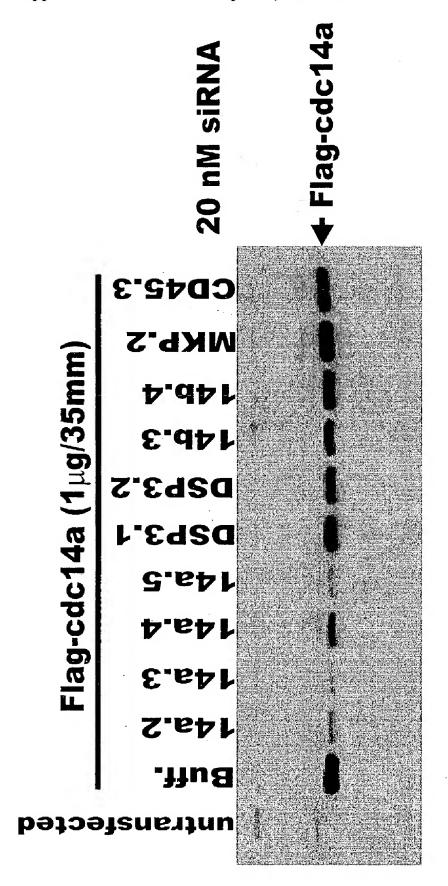
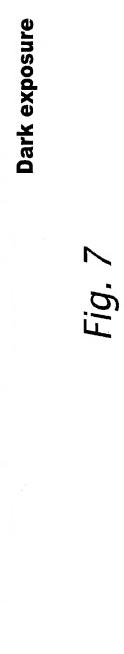
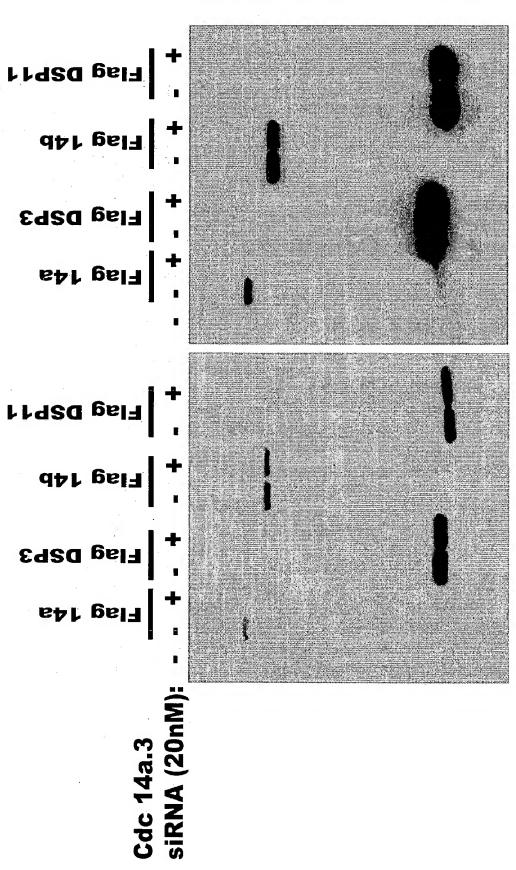
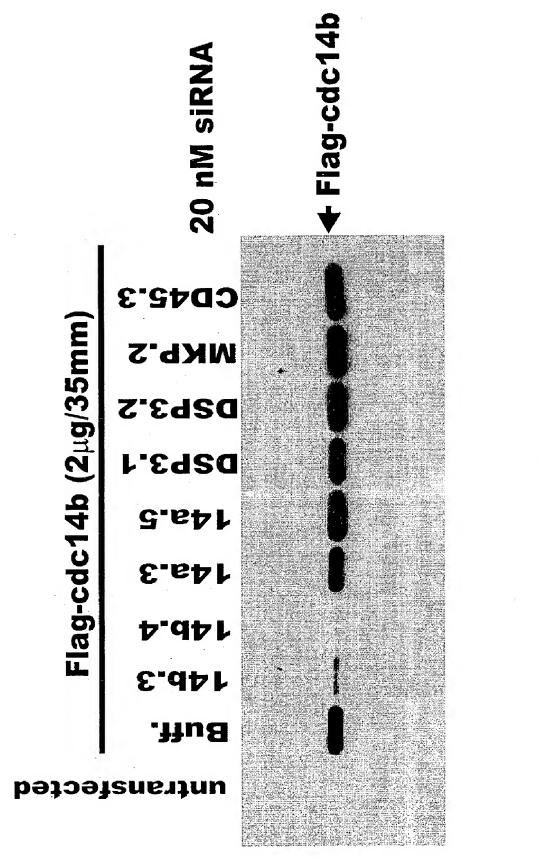
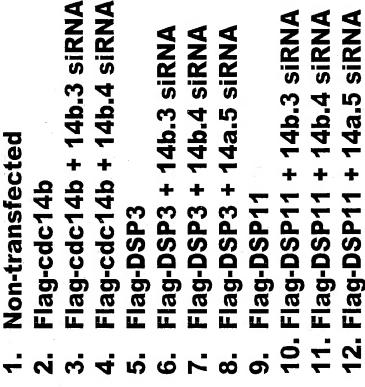


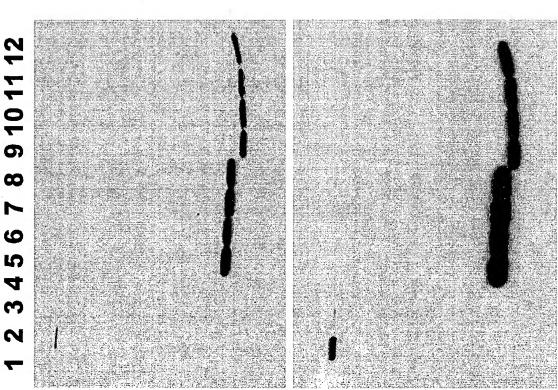
Fig. 6

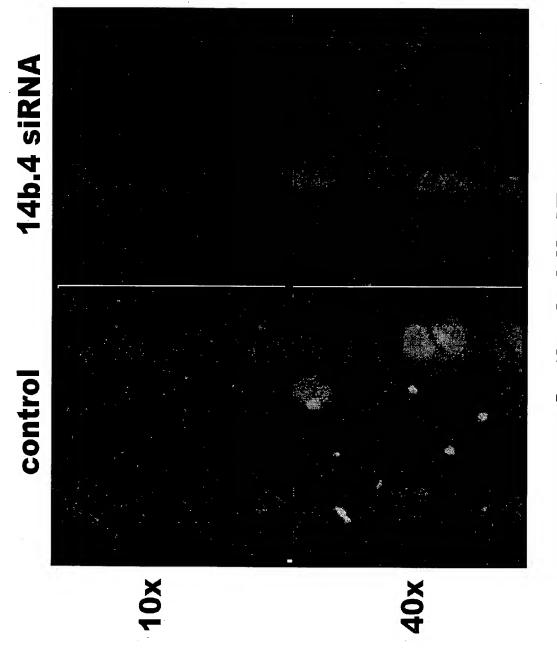






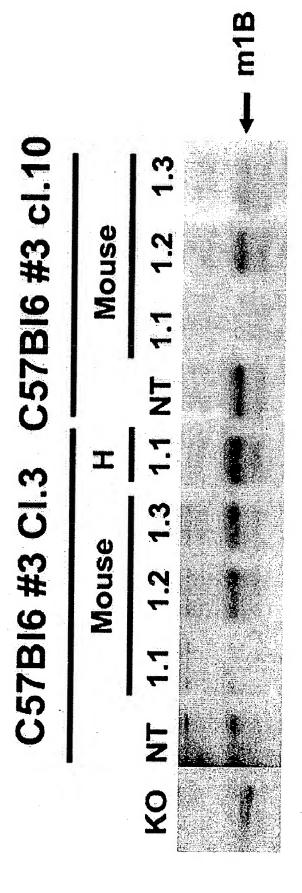






Anti cdc14b IF

Fig. 10



 Mouse fibroblasts were transfected with 200 nM RNAi oligonucleotides for a total of six days.

"NT" is non-transfected fibroblasts.

Fig. 12A

Prototypical DSP-18pr encoded by 708 base pairs

GGCCCCCGTTCCCCGCCAGGCTGCAGGCGTCGGGCCTGGGCCGTCAGGCCAGCTGTGACCGGATCGCTTC CCGGGCGGCGAGCTGGGGGTGCACCCGGACCGCCCCCCGGGATCATGGCAATGGCATGACCAAGGTAC TTCCTGGACTCTACCTCGGAAACTTCATTGATGCCAAAGACCTGGATCAGCTGGGCCGAAATAAGATCACA TGATACCCCTGAGGTACCCATCAAAAAGCACTTCAAAGAATGTATCAACTTCATCCACTGCCGCCCTTA ATGGGGGGAACTGCCTTGTGCACTGCTTTGCAGGCATCTCTCGCAGCACCACGATTGTGACAGCGTATGTG ATGACTGTGACGGGGCTAGGCTGGCGGGACGTGCTTGAAGCCATCAAGGCCACCAGGCCCATCGCCAACCC CAACCCAGGCTTTAGGCAGCAGCTTGAAGAGTTTGGCTGGGCCAGTTCCCAGAAGCTTCGCCGGCAGCTGG TGCCGGCAGGCTCCGCGGCCTCCTCCGCCGGGCCGCACTCAGCAGCCTCCGAGGGAACCGTGCA TCTCTTGCCTCCCCGGTGTCTGTCCCGCAAGGGCGGCAAGTGAGGATGCAG

Fig. 12B

Prototypical DSP-18pr polypeptide sequence 235 amino acids

MGNGMTKVLPGLYLGNFIDAKDLDQLGRNKITHIISIHESPQPLLQDITYLRIPVADTPEVPIKKHFKECI NFIHCCRLNGGNCLVHCFAGISRSTTIVTAYVMTVTGLGWRDVLEAIKATRPIANPNPGFRQQLEEFGWAS SQKLRRQLEERFGESPFRDEEELRALLPLCKRCRQGSATSASSAGPHSAASEGTVQRLVPRTPREAHRPLP LLARVKQTFSCLPRCLSRKGGK*

Fig. 13A

DSP-18a cDNA

GGCCCCCGTTCCCCGCCAGGCTGCAGGCGTCGGGCCTGGGCCGTCAGGGCAGCTGTGACCGGATCGCTTC CCGGGCGGCGAGCTGGGGTGCACCCGGACCGCCCCCGGGATCATGGCAATGGCATGACCAAGGTAC TTCCTGGACTCTACCTCGGAAACTTCATTGATGCCAAAGACCTGGATCAGCTGGGCCGAAATAAGATCACA TGATACCCCTGAGGTACCCATCAAAAAGCACTTCAAAGAATGTATCAACTTCATCCACTGCTGCCGCCTTA ATGGGGGGAACTGCCTTGTGCACTGCTTTGCAGGCATCTCTCGCAGCACCACGATTGTGACAGCGTATGTG ATGACTGTGACGGGGCTAGGCTGGCGGGACGTGCTTGAAGCCATCAAGGCCACCAGGCCCATCGCCAACCC CAACCCAGGCTTTAGGCAGCAGCTTGAAGAGTTTGGCTGGGCCAGTTCCCAGAAGGGTGCCAGACATAGGA CCTCAAAAACCTCTGGTGCCCAATGCCCTCCGATGACTTCAGCAACCTGGATGGTCACCGGACCCAAAGTA CGGACGTGCAGGTGCAGCTTCGGCCTGGGAGCTCGTCCTGCACTCTAAGTGCCTCAACCGAGCGCCCAGAT GGGTCCTCAACCCCTGGCAACCCCGATGGCATCACTCACCTTCAATGCAGCTGCCTCCATCCTAAGCGAGC CACACTAAGCCCATAGACTTGGGGCCTCCCCCGGCACATCACCCCAGGTCTGCCGGACGGCAGAGGTGGATC GCGGCCTTCCACTCCTGTCACGGGGCCCCGGAACTCGAGAGTAGGCCACACCGCCCCCAGCTGGGCAT GGGGCTTCGGCAGGAAACTGAACTTGATCTTGAGGCCCCAGAAAGGCAGCAACTGGAGCAGAAGCAAGACT CATTAAAACGTTTGCTTAAAGTTTTTTACCAATAATTAGATCATCAGGGTTGTTTAGTGTGGGATCAAGCCA TTCTTTATTCTGGGGGCTGGGAAGGATCCCAAAACAGGGAACTTGGCCGAACCCTGGGCTTTGGATGCTAA CCACTGAAGTACCAGCACCTGTAGGATGCTGTCTTTGAAGAAACTGAGGCGGACCTCCAAATGCAGCCCTA AGGCAGAGGTCAACGTGGAAGACCAGCCCTTCTCCAAGCCCCACTGGTCTTTGCAAGCTGTACGTTGTAGG CAATCTGAGAACTGGAAAGGGGGACTACAACCAGAAAGTTGGTTACCCTGCCATGGGAATAAAGTAGCTGT TTTCCACCCCAAAAAAAAAAAAAAAAAAAAAA

Fig. 13B

DSP-18a polypeptide (181 amino acids)

MGNGMTKVLPGLYLGNFIDAKDLDQLGRNKITHIISIHESPQPLLQDITYLRIPVADTPEVPIKKHFKECI NFIHCCRLNGGNCLVHCFAGISRSTTIVTAYVMTVTGLGWRDVLEAIKATRPIANPNPGFRQQLEEFGWAS SQKGARHRTSKTSGAQCPPMTSATWMVTGPKVPDLSVLR*

Fig. 14A

DSP-18b cDNA

GGCCCCCGTTCCCCGCCAGGCTGCAGGCGTCGGGCCTGGGCCGTCAGGGCAGCTGTGACCGGATCGCTTC CCGGGCGGCGAGCTGGGGTGCACCCGGACCGCCCCCCGGGATCATGGCAATGGCATGACCAAGGTAC TTCCTGGACTCTACCTCGGAAACTTCATTGATGCCAAAGACCTGGATCAGCTGGGCCGAAATAAGATCACA TGATACCCCTGAGGTACCCATCAAAAAGCACTTCAAAGAATGTATCAACTTCATCCACTGCCGCCTTA ATGGGGGGAACTGCCTTGTGCACTGCTTTGCAGGCATCTCTCGCAGCACCACGATTGTGACAGCGTATGTG ATGACTGTGACGGGGCTAGGCTGCGGGGACGTGCTTGAAGCCATCAAGGCCACCAGGCCCATCGCCAACCC CAACCCAGGCTTTAGGCAGCAGCTTGAAGAGTTTGGCTGGGCCAGTTCCCAGAAGGGTGCCAGACATAGGA CTTCTCCGCAGCGCTGGTGCGCGAAGCCACCGGGCGCACAGCCCAGCGCTGTCGTCTGAGTCCGCGGGC GGCCGCCGAGCGCCTGCTGGGGCCGCCACCTCACGTTGCAGCAGGATGGTCACCGGACCCAAAGTACCAGA GTGCAGGTGCAGCTTCGGCCTGGGAGCTCGTCCTGCACTCTAAGTGCCTCAACCGAGCGCCCAGATGGGTC CTCAACCCCTGGCAACCCCGATGGCATCACTCACCTTCAATGCAGCTGCCTCCATCCTAAGCGAGCCGCTT TAAGCCCATAGACTTGGGGCCTCCCCGGCGCACATCACCCAGGTCTGCCGGACGCAGAGGTGGATCGCG GCCTTCCACTCCTCTGTCACGGGGCCCCGGAACTCGAGAGTAGGCCACACCGCCCCCAGCTGGGCATGGG GCTTCGGCAGGAAACTGAACTTGATCTTGAGGCCCCAGAAAGGCAGCAACTGGAGCAGAAGCAAGACTTCA TAAAACGTTTGCTTAAAGTTTTTTACCAATAATTAGATCATCAGGGTTGTTTAGTGTGGGATCAAGCCATAA TTTATTCTGGGGGCTGGGAAGGATCCCAAAACAGGGAACTTGGCCGAACCCTGGGCTTTGGATGCTAACCA CTGAAGTACCAGCACCTGTAGGATGCTGTCTTTGAAGAAACTGAGGCGGACCTCCAAATGCAGCCCTAAGG CAGAGGTCAACGTGGAAGACCAGCCCTTCTCCAAGCCCCACTGGTCTTTGCAAGCTGTACGTTGTAGGCAA TCTGAGAACTGGAAAGGGGGACTACAACCAGAAAGTTGGTTACCCTGCCATGGGAATAAAGTAGCTGTTTT CCACCCCAAAAAAAAAAAAAAAAAAAAAA

Fig. 14B

DSP-18b polypeptide (298 amino acids)

MGNGMTKVLPGLYLGNFIDAKDLDOLGRNKITHIISIHESPOPLLODITYLRIPVADTPEVPIKKHFKEC1 NFIHCCRLNGGNCLVHCFAGISRSTTIVTAYVMTVTGLGWRDVLEAIKATRPIANPNPGFRQQLEEFGWAS SQKGARHRTSKTSGAQCPPMTSATCLLAARVALLSAALVREATGRTAQRCRLSPRAAAERLLGPPPHVAAG WSPDPKYQICLCFGEEDPGPT0HPKE0LIMADV0V0LRPGSSSCTLSASTERPDGSSTPGNPDG1THL0CS CLHPKRAASSSCTR*

DSP-18c cDNA

Fig. 15A

GGCCCCCGTTCCCCGCCAGGCTGCAGGCGTCGGGCCTGGGCCGTCAGGGCAGCTGTGACCGGAT CGCTTCCCGGGCGGCGAGCTGGGGGTGCACCCGGACCGCCCCCGGGATCATGGCAATGGCA TGACCAAGGTACTTCCTGGACTCTACCTCGGAAACTTCATTGATGCCAAAGACCTGGATCAGCTG GGCCGAAATAAGATCACACACACATCATCTCTATCCATGAGTCACCCCAGCCTCTGCTGCAGGATAT CACCTACCTTCGCATCCCGGTCGCTGATACCCCTGAGGTACCCATCAAAAAGCACTTCAAAGAAT GTATCAACTTCATCCACTGCTGCCGCCTTAATGGGGGGAACTGCCTTGTGCACTGCTTTGCAGGC ATCTCTCGCAGCACCACGATTGTGACAGCGTATGTGATGACTGTGACGGGGCTAGGCTGGCGGGA CGTGCTTGAAGCCATCAAGGCCACCAGGCCCATCGCCAACCCCAACCCAGGCTTTAGGCAGCAGC TTGAAGAGTTTGGCTGGGCCAGTTCCCAGAAGGGTGCCAGACATAGGACCTCAAAAACCTCTGGT GCCCAATGCCCTCCGATGACTTCAGCAACCTGGATGGTCACCGGACCCAAAGTACCAGATCTGTC GTGCAGGTGCAGCTTCGGCCTGGGAGCTCGTCCTGCACTCTAAGTGCCTCAACCGAGCGCCCAGA AAGCGAGCCGCTTCCTCTTCTTGTACCCGCTGAAGGCAAGCCCCCAACAGGGGGGCTCCCTACTC CCACCCAACCCTGCCCACACTAAGCCCATAGACTTGGGGCCTCCCCCGGCACATCACCCAGGTCT GCCGGACGCAGAGGTGGATCGCGGCCTTCCACTCCTCTGTCACGGGGCCCCGGAACTCGAGAGT AGGCCTCACCGCCCCCAGCTGGGCATGGGGCTTCGGCAGGAAACTGAACTTGATCTTGAGGCCA GCAGAAAGGCAGCAACTGGAGCAGAAGCAAGACTTCATCTCTTGCTGACAGCCCAATTTGTCAAT -AGCGCTTTCCTCAGAGCCAGCCTTAACCTGCTGTTGAGTCCATTAAAACGTTTGCTTAAAGTTTT TACCAATAAAAAAAAAAAAAAAAAAAAAAA

Fig. 15B

DSP-18d cDNA

GGCCCCCGTTCCCCGCCAGGCTGCAGGCGTCGGGCCTGGGCCGTCAGGGCAGCTGTGACCGGATCGCTTC CCGGGCGGCGAGCTGGGGGTGCACCCGGACCGCCCCCGGGATCATGGCAATGGCATGACCAAGGTAC TTCCTGGACTCTACCTCGGAAACTTCATTGATGCCAAAGACCTGGATCAGCTGGGCCGAAATAAGATCACA TGATACCCCTGAGGTACCCATCAAAAAGCACTTCAAAGAATGTATCAACTTCATCCACTGCCGCCTTA ATGGGGGGAACTGCCTTGTGCACTGCTTTGCAGGCATCTCTCGCAGCACCACGATTGTGACAGCGTATGTG ATGACTGTGACGGGGCTAGGCTGGCGGGACGTGCTTGAAGCCATCAAGGCCACCAGGCCCATCGCCAACCC CAACCCAGGCTTTAGGCAGCAGCTTGAAGAGTTTGGCTGGGCCAGTTCCCAGAAGGGTGCCAGACATAGGA CCTCAAAAACCTCTGGTGCCCAATGCCCTCCGATGACTTCAGCAACCTGGATGGTCACCGGACCCAAAGTA CGGACCTAGTCTCTTTTATTCTGGGGGCTGGGAAGGATCCCAAAACAGGGAACTTGGCCGAACCCTG GGCTTTGGATGCTAACCACTGAAGTACCAGCACCTGTAGGATGCTGTCTTTGAAGAAACTGAGGCGGACCT CCAAATGCAGCCCTAAGGCAGAGGTCAACGTGGAAGACCAGCCCTTCTCCAAGCCCCACTGGTCTTTGCAA GCTGTACGTTGTAGGCAATCTGAGAACTGGAAAGGGGGGACTACAACCAGAAAGTTGGTTACCCTGCCATGG

Fig. 16A

DSP-18e cDNA

GGCCCCCGTTCCCCGCCAGGCTGCAGGCGTCGGGCCTGGGCCGTCAGGGCAGCTGTGACCGGATCGCTTC CCGGGCGGCGAGCTGGGGGTGCACCCGGGACCGCCCCCGGGATCATGGCCAATGGCATGACCAAGGTAC TTCCTGGACTCTACCTCGGAAACTTCATTGATGCCAAAGACCTGGATCAGCTGGGCCGAAATAAGATCACA TGATACCCCTGAGGTACCCATCAAAAAGCACTTCAAAGAATGTATCAACTTCATCCACTGCTGCCGCCTTA ATGGGGGGAACTGCCTTGTGCACTGCTTTGCAGGCATCTCTCGCAGCACCACGATTGTGACAGCGTATGTG ATGACTGTGACGGGGCTAGGCTGGCGGGACGTGCTTGAAGCCATCAAGGCCACCAGGCCCATCGCCAACCC CAACCCÁGGCTTTAGGCÁGCAGCTTAAGAGTTTGGCTGGGCCAGTTCCCAGAAGGATGGTCACCGGÁCCCA CATGGCGGACCTAGTCTCTTTTATTCTGGGGGCTGGGAAGGATCCCAAAACAGGGAACTTGGCCGAA CCCTGGGCTTTGGATGCTAACCACTGAAGTACCAGCACCTGTAGGATGCTGTCTTTGAAGAAACTGAGGCG GACCTCCAAATGCAGCCCTAAGGCAGAGGTCAACGTGGAAGACCAGCCCTTCTCCAAGCCCCACTGGTCTT TGCAAGCTGTACGTTGTAGGCAATCTGAGAACTGGAAAGGGGGGACTACAACCAGAAAGTTGGTTACCCTGC

Fig. 16B

DSP-18e polypeptide (159 amino acids)

MGNGMTKVLPGLYLGNFIDAKDLDQLGRNKITHIISIHESPQPLLQDITYLRIPVADTPEVPIKKHFKECI NFIHCCRLNGGNCLVHCFAGISRSTTIVTAYVMTVTGLGWRDVLEAIKATRPIANPNPGFROOLKSLAGPV PRRMVTGPKVPDLSVLR*

Fig. 17A

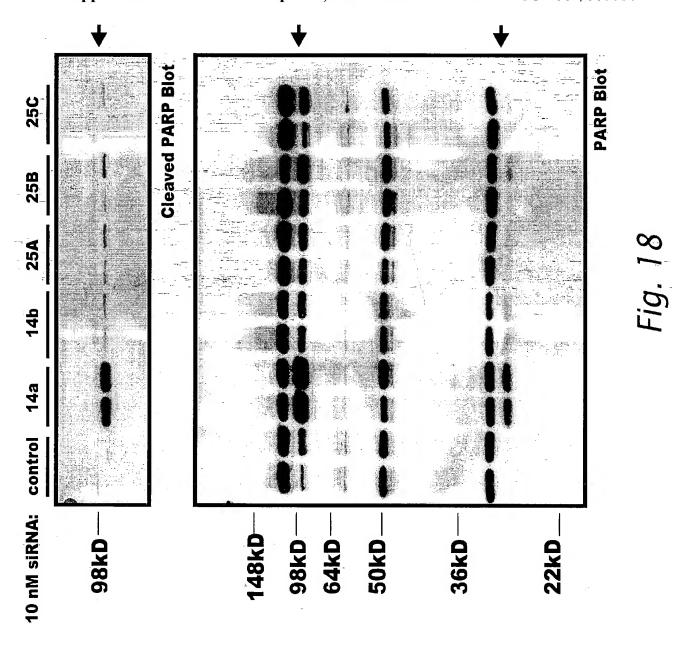
DSP-18f cDNA

GGCCCCCGTTCCCCGCCAGGCTGCAGGCGTCGGGCCTGGGCCGTCAGGGCAGCTGTGACCGGATCGCTTC CCGGGCGGCGAGCTGGGGGTGCACCCGGACCGCCCCCGGGATCATGGCAATGGCATGACCA**AGG**TAC TTCCTGGACTCTACCTCGGAAACTTCATTGATGCCAAAGACCTGGATCAGCTGGGCCGAAATAAGATCACA TGATACCCCTGAGGTACCCATCAAAAAGCACTTCAAAGAATGTATCAACTTCATCCACTGCTGCCGCCTTA ATGGGGGAACTGCCTTGTGCACTGCTTTGCAGGCATCTCTCGCAGCACCACGATTGTGACAGCGTATGTG ATGACTGTGACGGGGCTAGGCTGGCGGGACGTGCTTGAAGCCATCAAGGCCACCAGGCCCATCGCCAACCC CAACCCAGGCTTTAGGCAGCAGCTTGAAGAGTTTGGCTGGGCCAGTTCCCAGAAGGGCTTTTACCAACCTC ATAAGCTGTTGTGAGAACCAATTGAGACACTGCAGGAAAGTGTTTAGCCAGGCCCAGCACTGATGAGCAGT CGGATGGTCACCGGACCCAAAGTACCAGATCTGTCTGTGCTTCGGTGAGGAGGACCCGGGCCCCACACAGC ACCCCAAGGAGCAGCTCATCATGGCGGACCTAGTCTCTTCTTTATTCTGGGGGGCTGGGAAGGATCCCAA AACAGGGAACTTGGCCGAACCCTGGGCTTTGGATGCTAACCACTGAAGTACCAGCACCTGTAGGATGCTGT CTITGAAGAAACTGAGGCGGACCTCCAAATGCAGCCCTAAGGCAGAGGTCAACGTGGAAGACCAGCCCTTC TCCAAGCCCCACTGGTCTTTGCAAGCTGTACGTTGTAGGCAATCTGAGAACTGGAAAGGGGGACTACAACC

Fig. 17B

DSP-18f polypeptide (154 amino acids)

MGNGMTKVLPGEYLGNFIDAKDLDQLGRNKITHIISIHESPQPLLQDITYLRIPVADTPEVPIKKHFKECI NFIHCCRLNGGNCLVHCFAGISRSTTIVTAYVMTVTGLGWRDVLEAIKATRPIANPNPGFRQQLEEFGWAS SOKGFYOPHKLL*





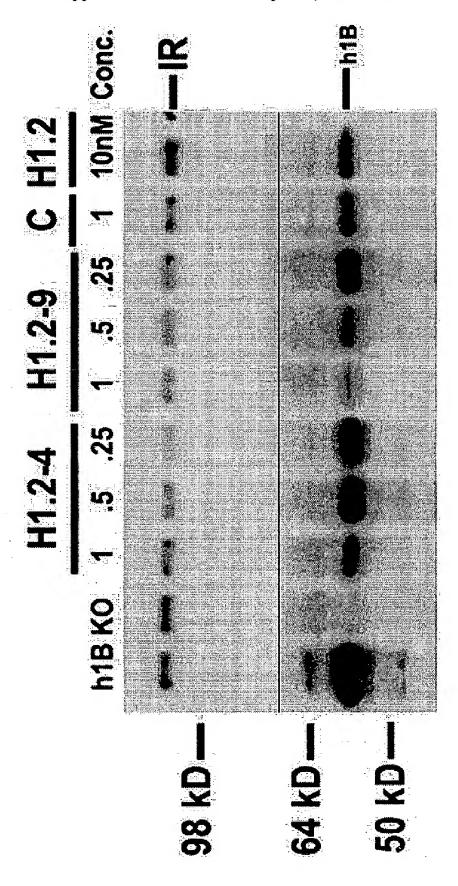


Fig. 20A

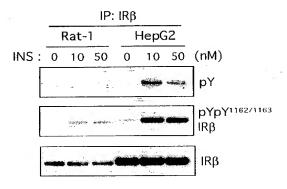


Fig. 20B

HepG2 Control +siRNA INS: 0 1 2 5 10 20 0 1 2 5 10 20 (min) p-AKT AKT TC45 PTP1B

Fig. 20C

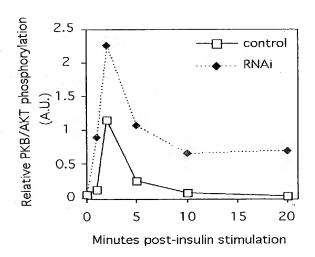


Fig. 21A

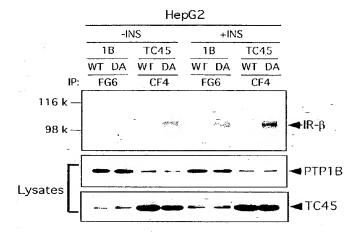
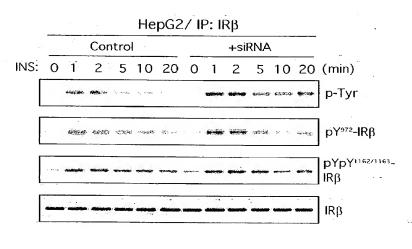


Fig. 21B

Fig. 21C



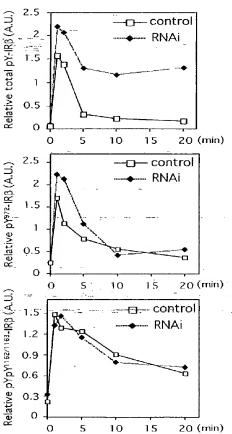
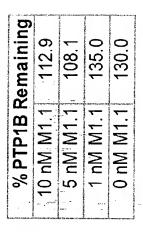
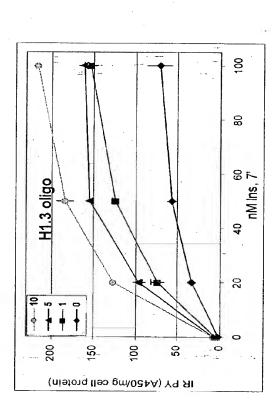
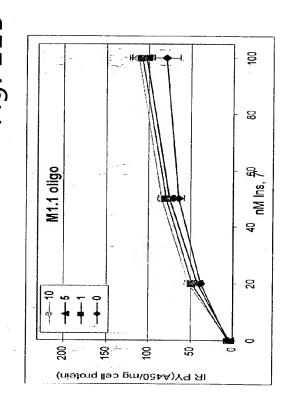
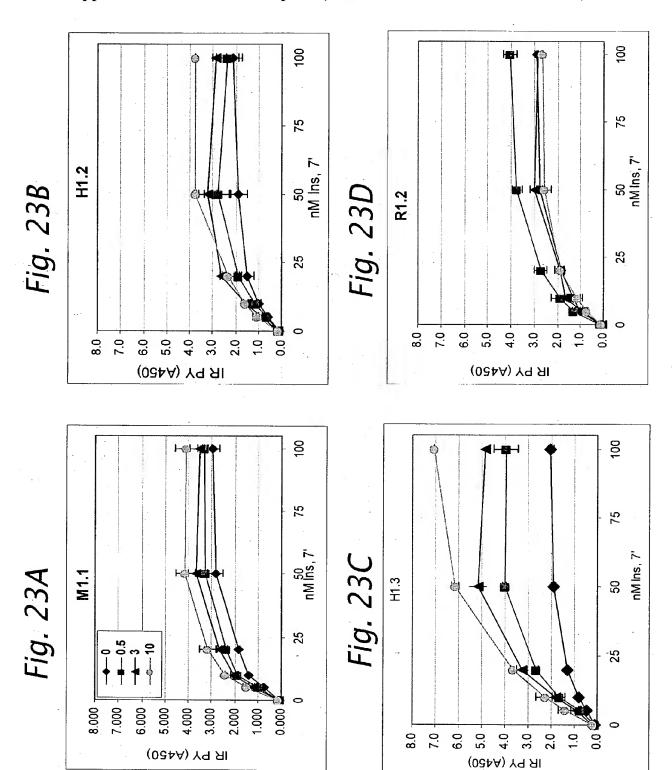


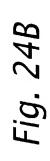
Fig. 22A

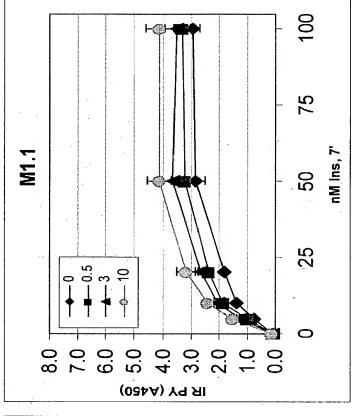












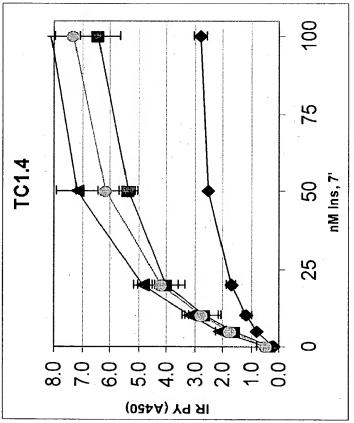


Fig. 24A

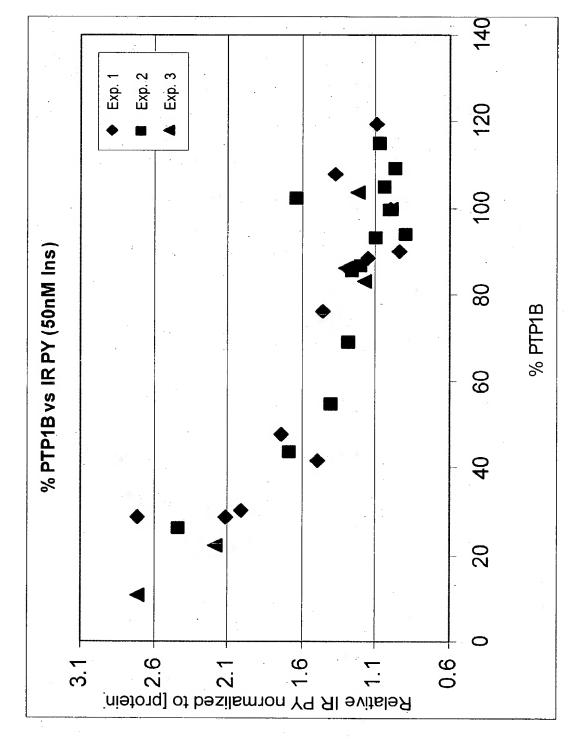
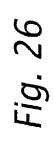
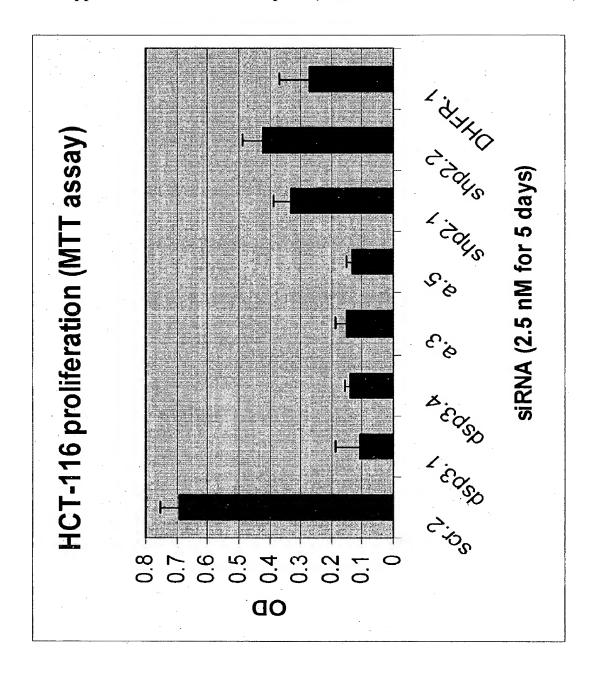
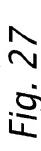
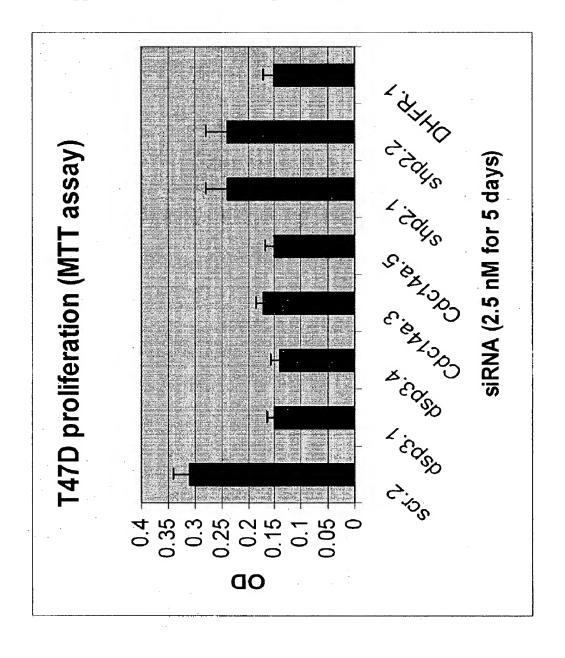


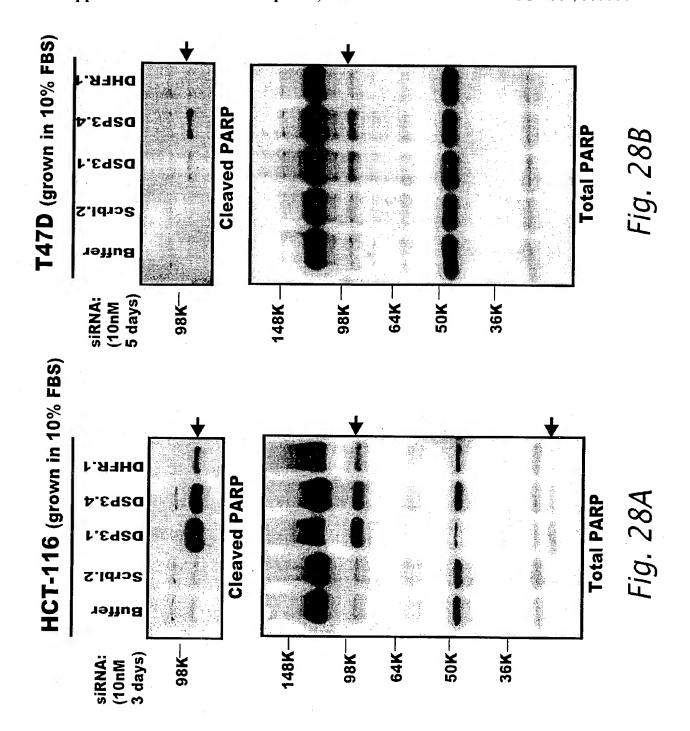
Fig. 25











DSP-13 Encoding Polynucleotide

cctgggaaga agttatctat ctctcgagtg acattcaaga tataccgtac ccctcggttc 60 tgtaagteet etaagttgga ggeatteeat tetgageegg eeceatgace etgageaegt 120 tggcccgcaa gaggaaggcg cccctcgctt gcacctgcag cctcggtggc cccgacatga 180 tteettaett eteegeeaac geggteatet egeagaacge eateaaceag eteateageg 240 agagetttet aaetgteaaa ggtgetgeee tttttetace aeggggaaat ggeteateea 300 caccaagaat cagccacaga cggaacaagc atgcaggcga tctccaacag catctccaag 360 caatgtteat tttaeteege eeagaagaca acateagget ggetgtaaga etggaaagta 420 cttaccagaa tcgaacacgc tatatggtag tggtttcaac taatggtaga caagacactg 480 aagaaagcat cgtcctagga atggatttct cctctaatga cagtagcact tgtaccatgg 540 gcttagtttt gcctctctgg agcgacacgc taattcattt ggatggtgat ggtgggttca 600 gtgtatcgac ggataacaga gttcacatat tcaaacctgt atctgtgcag gcaatgtggt 660 ctgcactaca gagcttacac aaggcttgtg aagtcgccag agcgcataac tactacccag 720 geagectatt teteaettgg gtgagttatt atgagageca tateaaetea gateaateet 780 cagteaatga atggaatgea atgeaagatg taeagteeca eeggeeegae teteeagete 840 tetteacega catacetaet gaacgtgaac gaacagaaag getaattaaa accaaattaa 900 gggagatcat gatgcagaag gatttggaga atattacatc caaagagata agaacagagt 960 1020 tagtgatect tggteaaatg gatageeeta eacagatatt tgageatgtg tteetggget 1080 cagaatggaa tgcctccaac ttagaggact tacagaaccg aggggtacgg tatatcttga 1140 atgtcactcg agagatagat aacttcttcc caggagtctt tgagtatcat aacattcggg 1200 tatatgatga agaggcaacg gatctcetgg cgtactggaa tgacacttac aaattcatct 1260 ctaaagcaaa gaaacatgga tctaaatgcc ttgtgcactg caaaatgggg gtgagtcgct 1320 cagecteeae egtgattgee tatgeaatga aggaatatgg etggaatetg gaeegageet 1380 atgactatgt gaaagaaaga cgaacggtaa ccaagcccaa cccaagcttc atgagacaac 1440 tggaagagta tcaggggatc ttgctggcaa gettectagg cttgattcat ggagggaggg 1500 acaagccetg gggagagaaa agcacagaat ttgagtcagt agatctggtt tecatteetg 1560 gttcacccte ttgetgeaac cetgagaagt tacttcacat ttctcatcct tacctgaccc 1620 catctataaa atgaaaatca agagatccat ctcacagggt tattgtgaat aaaaatgtgt 1680 ttgaatgttt ataaaaaaaa aaaaaaaaaa a 1711

Met Thr Leu Ser Thr Leu Ala Arg Lys Arg Lys Ala Pro Leu Ala Cys Thr Cys Ser Leu Gly Gly Pro Asp Met Ile Pro Tyr Phe Ser Ala Asn Ala Val Ile Ser Gln Asn Ala Ile Asn Gln Leu Ile Ser Glu Ser Phe Leu Thr Val Lys Gly Ala Ala Leu Phe Leu Pro Arg Gly Asn Gly Ser Ser Thr Pro Arg Ile Ser His Arg Arg Asn Lys His Ala Gly Asp Leu Gln Gln His Leu Gln Ala Met Phe Ile Leu Leu Arg Pro Glu Asp Asn Ile Arg Leu Ala Val Arg Leu Glu Ser Thr Tyr Gln Asn Arg Thr Arg Tyr Met Val Val Ser Thr Asn Gly Arg Gln Asp Thr Glu Glu Ser Ile Val Leu Gly Met Asp Phe Ser Ser Asn Asp Ser Ser Thr Cys Thr Met Gly Leu Val Leu Pro Leu Trp Ser Asp Thr Leu Ile His Leu Asp Gly Asp Gly Gly Phe Ser Val Ser Thr Asp Asn Arg Val His Ile Phe Lys Pro Val Ser Val Gln Ala Met Trp Ser Ala Leu Gln Ser Leu His Lys Ala Cys Glu Val Ala Arg Ala His Asn Tyr Tyr Pro Gly Ser Leu Phe Leu Thr Trp Val Ser Tyr Tyr Glu Ser His Ile Asn Ser Asp Gln Ser Ser Val Asn Glu Trp Asn Ala Met Gln Asp Val Gln Ser His Arg Pro Asp Ser Pro Ala Leu Phe Thr Asp lle Pro Thr Glu Arg Glu Arg Thr Glu Arg Leu Ile Lys Thr Lys Leu Arg Glu Ile Met Met Gln Lys Asp Leu Glu Asn Ile Thr Ser Lys Glu Ile Arg Thr Glu Leu Glu Met Gln Met Val Cys Asn Leu Arg Glu Phe Lys Glu Phe Ile Asp Asn Glu Met Ile Val Ile Leu Gly Gln Met Asp Ser Pro Thr Gln Ile Phe Glu His Val Phe Leu Gly Ser Glu Trp Asn Ala Ser Asn Leu Glu Asp Leu Gln Asn Arg Gly Val Arg Tyr Ile Leu Asn Val Thr Arg Glu Ile Asp Asn Phe Pro Gly Val Phe Glu Tyr His Asn Ile Arg Val Tyr Asp Glu Glu Ala Thr Asp Leu Leu Ala Tyr Trp Asn Asp Thr Tyr Lys Phe Ile Ser Lys Ala Lys Lys His Gly Ser Lys Cys Leu Val His Cys Lys Met Gly Val Ser Arg Ser Ala Ser Thr Val Ile Ala Tyr Ala Met Lys Glu Tyr Gly Trp Asn Leu Asp Arg Ala Tyr Asp Tyr Val Lys Glu Arg Arg Thr Val Thr Lys Pro Asn Pro Ser Phe Met Arg Gln Leu Glu Glu Tyr Gln Gly Ile Leu Leu Ala Ser Phe Leu Gly Leu Ile His Gly Gly Arg Asp Lys Pro Trp Gly Glu Lys Ser Thr Glu Phe Glu Ser Val Asp Leu Val Ser Ile Pro Gly Ser Pro Ser Cys Cys Asn Pro Glu Lys Leu Leu His Ile Ser His Pro Tyr Leu Thr Pro Ser Ile Lys

DSP-14 Encoding Polynucleotide

ggccagtggg ggtggctggg cgtgcggctg ctacatgccc cacggaccag aacctcccga 60 cgcggccagg ccccggcaca cccagctgca gaaaggagag aaaatccctt ggctctaaaa 120 tracatetgg agaagtgaag acaagcetea agaatgeeta eteatetgee aagaggetgt 180 cgccgaagat ggaggaggaa ggggaggagg aggactactg cacccetgga gcctttgagc 240 tggagegget ettetggaag ggeagteece agtacaceca egteaacgag gtetggeeca 300 agetetacat tggcgatgag gegaeggege tggaeegeta taggetgeag aaggeggggt 360 teacgeacgt getgaacgeg geceaeggee getggaacgt ggacactggg eecgactact 420 accgcgacat ggacatccag taccacggcg tggaggccga cgacctgccc accttcgacc 480 teagtgtett ettetaceeg geggeageet teategaeag agegetaage gaegaeeaea 540 gtaagateet ggtteaetge gteatgggee geageeggte ageeaecetg gteetggeet 600 acctgatgat ccacaaggac atgaccetgg tggacgccat ccagcaagtg gccaagaacc 660 getgegteet eeegaacegg ggetttttga ageageteeg ggagetggae aageagetegg 720 tgcagcagag gcgacggtcc cagcgccagg acggtgagga ggaggatggc agggagctgt 780 aggecegact cacagggeca geagaggeae ttggggacag aggggagagg cagaacatag 840 ccctggccta ggactccaga gaagggatgg tgaaaccgaa getegaetet tecaaaccat 900 cttgttcaac ttccccatgt gtgctgggga cagggaggac ccagagctgc ccccgggcag 960 agetgagege teageetete ageaaaatgg gagggaeggg eteeeegget etgggteaea 1020 gaggagcatg ccacgctgca ccaagtetee tgetttggtt ttgttttttt ggtgagaagg 1080 aagaggaaa aagatttta aaatgtgtag gcagtatgtt gtgattaaac gtttggcttt 1140 1165 gtccaaaaaa aaaaaaaaaa aaaaa

Fig. 30A

DSP-14 Polypeptide Sequence

Met Thr Ser Gly Glu Val Lys Thr Ser Leu Lys Asn Ala Tyr Ser Ser Ala Lys Arg Leu Ser Pro Lys Met Glu Glu Glu Glu Glu Glu Glu Glu Asp Tyr Cys Thr Pro Gly Ala Phe Glu Leu Glu Arg Leu Phe Trp Lys Gly Ser Pro Gln Tyr Thr His Val Asn Glu Val Trp Pro Lys Leu Tyr Ile Gly Asp Glu Ala Thr Ala Leu Asp Arg Tyr Arg Leu Gln Lys Ala Gly Phe Thr His Val Leu Asn Ala Ala His Gly Arg Trp Asn Val Asp Thr Gly Pro Asp Tyr Tyr Arg Asp Met Asp Ile Gln Tyr His Gly Val Glu Ala Asp Asp Leu Pro Thr Phe Asp Leu Ser Val Phe Phe Tyr Pro Ala Ala Ala Phe Ile Asp Arg Ala Leu Ser Asp Asp His Ser Lys Ile Leu Val His Cys Val Met Gly Arg Ser Arg Ser Ala Thr Leu Val Leu Ala Tyr Leu Met Ile His Lys Asp Met Thr Leu Val Asp Ala Ile Gln Gln Val Ala Lys Asn Arg Cys Val Leu Pro Asn Arg Gly Phe Leu Lys Gln Leu Arg Glu Leu Asp Lys Gln Leu Val Gln Gln Arg Arg Arg Ser Gln Arg Gln Asp Gly Glu Glu Glu Asp Gly Arg Glu Leu

Fig. 30B

MODULATION OF BIOLOGICAL SIGNAL TRANSDUCTION BY RNA INTERFERENCE

CROSS-REFERENCE TO RELATED APPLICATION

[0001] This application claims the benefit of U.S. Provisional Patent Application No. 60/383,249 filed May 23, 2002, and U.S. Provisional Patent Application No. 60/462, 942 filed Apr. 14, 2003, which are incorporated herein by reference in their entirety.

BACKGROUND OF THE INVENTION

[0002] 1. Technical Field

The present invention relates generally to compositions and methods useful for treating conditions associated with defects in cell proliferation, cell differentiation, and cell survival. The invention is more particularly related to double-stranded RNA polynucleotides that interfere with expression of protein tyrosine phosphatases, and polypeptide variants thereof. The invention is also particularly related to double-stranded RNA polynucleotides that interfere with expression of MAP kinases and MAP kinase kinases and chemotherapeutic target polypeptides, and polypeptide variants thereof. The present invention is also related to the use of such RNA polynucleotides to alter activation of signal transduction pathway components or to alter cellular metabolic processes that lead to proliferative responses, cell differentiation and development, and cell survival.

[0004] 2. Description of the Related Art

Reversible protein tyrosine phosphorylation, coordinated by the action of protein tyrosine kinases (PTKs) that phosphorylate certain tyrosine residues in polypeptides, and protein tyrosine phosphatases (PTPs) that dephosphorylate certain phosphotyrosine residues, is a key mechanism in regulating many cellular activities. It is becoming apparent that the diversity and complexity of the PTPs and PTKs are comparable, and that PTPs are equally important in delivering both positive and negative signals for proper function of cellular machinery. Regulated tyrosine phosphorylation contributes to specific pathways for biological signal transduction, including those associated with cell division, cell survival, apoptosis, proliferation and differentiation. Defects and/or malfunctions in these pathways may underlie certain disease conditions for which effective means for intervention remain elusive, including for example, malignancy, autoimmune disorders, diabetes, obesity and infection.

[0006] The protein tyrosine phosphatase (PTP) family of enzymes consists of more than 100 structurally diverse proteins in vertebrates, including almost 40 human PTPs that have in common the conserved 250 amino acid PTP catalytic domain, but which display considerable variation in their non-catalytic segments (Charbonneau and Tonks, 1992 Annu. Rev. Cell Biol. 8:463-493; Tonks, 1993 Semin. Cell Biol. 4:373-453; Andersen et al., Mol. Cell Biol. 21:7117-36 (2001)). This structural diversity presumably reflects the diversity of physiological roles of individual PTP family members, which in certain cases have been demonstrated to have specific functions in growth, development and differentiation (Desai et al., 1996 Cell 84:599-609; Kishihara et al., 1993 Cell 74:143-156; Perkins et al., 1992

Cell 70:225-236; Pingel and Thomas, 1989 Cell 58:1055-1065; Schultz et al., 1993 Cell 73:1445-1454). The PTP family includes receptor-like and non-transmembrane enzymes that exhibit exquisite substrate specificity in vivo and that are involved in regulating a wide variety of cellular signaling pathways (Andersen et al., Mol. Cell. Biol. 21:7117 (2001); Tonks and Neel, Curr. Opin. Cell Biol. 13:182 (2001)). PTPs thus participate in a variety of physiologic functions, providing a number of opportunities for therapeutic intervention in physiologic processes through alteration (i.e., a statistically significant increase or decrease) or modulation (e.g., up-regulation or down-regulation) of PTP activity.

[0007] Although recent studies have also generated considerable information regarding the structure, expression and regulation of PTPs, the nature of many tyrosine phosphorylated substrates through which the PTPs exert their effects remains to be determined. Studies with a limited number of synthetic phosphopeptide substrates have demonstrated some differences in the substrate selectivities of different PTPs (Cho et al., 1993 Protein Sci. 2: 977-984; Dechert et al., 1995 Eur. J. Biochem. 231:673-681). Analyses of PTP-mediated dephosphorylation of PTP substrates suggest that catalytic activity may be favored by the presence of certain amino acid residues at specific positions in the substrate polypeptide relative to the phosphorylated tyrosine residue (Salmeen et al., 2000 Molecular Cell 6:1401; Myers et al., 2001 J. Biol. Chem. 276:47771; Myers et al., 1997 Proc. Natl. Acad. Sci. USA 94:9052; Ruzzene et al., 1993 Eur. J. Biochem. 211:289295; Zhang et al., 1994 Biochemistry 33:2285-2290). Thus, although the physiological relevance of the substrates used in these studies is unclear, PTPs display a certain level of substrate selectivity in vitro.

[0008] The PTP family of enzymes contains a common evolutionarily conserved segment of approximately 250 amino acids known as the PTP catalytic domain. Within this conserved domain is a unique signature sequence motif, CX₅R (SEQ ID NO: ___), that is invariant among all PTPs. In a majority of PTPs, an 11 amino acid conserved ([IIV]HCXAGXXR[S/T)G sequence (SEO NO:)) containing the signature sequence motif is found. The cysteine residue in this motif is invariant in members of the family and is essential for catalysis of the phosphotyrosine dephosphorylation reaction. It functions as a nucleophile to attack the phosphate moiety present on a phosphotyrosine residue of the incoming substrate. If the cysteine residue is altered by site-directed mutagenesis to serine (e.g., in cysteine-to-serine or "CS" mutants) or alanine (e.g., cysteine-to-alanine or "CA" mutants), the resulting PTP is catalytically deficient but retains the ability to complex with, or bind, its substrate, at least in vitro.

[0009] The CS mutant of one PTP, PTP1B (PTP-1B), is an example of such a PTP. Catalytically deficient mutants of such enzymes that are capable of forming stable complexes with phosphotyrosyl polypeptide substrates may be derived by mutating a wildtype protein tyrosine phosphatase catalytic domain invariant aspartate residue and replacing it with an amino acid that does not cause significant alteration of the Km of the enzyme but that results in a reduction in Kcat, as disclosed, for example, in U.S. Pat. Nos. 5,912,138 and 5,951,979, in U.S. application Ser. No. 09/323,426 and in PCT/US97/13016 and PCT/JUS00/14211. For instance,

mutation of Asp 181 in PTP1B to alanine to create the aspartate-to-alanine (D to A or DA) mutant PTP1B-D181A results in a PTP1B "substrate trapping" mutant enzyme that forms a stable complex with its phosphotyrosyl polypeptide substrate (e.g., Flint et al., 1997 *Proc. Natl. Acad. Sci.* 94:1680). Substrates of other PTPs can be identified using a similar substrate trapping approach, for example substrates of the PTP family members PTP-PEST (Garton et al., 1996 *J. Mol. Cell. Biol.* 16:6408), TCPTP (Tiganis et al., 1998 *Mol. Cell Biol.* 18:1622), PTP-HSCF (Spencer et al., 1997 *J. Cell Biol.* 138:845), and PTP-H1 (Zhang et al., 1999 *J. Biol. Chem.* 274:17806).

[0010] Mitogen-activated protein kinases (MAP-kinases) are components of conserved cellular signal transduction pathways that have a variety of conserved members and that that are integral to the cell's response to stimuli such as growth factors, hormones, cytokines, and environmental stresses. MAP-kinases are activated by phosphorylation by MAP-kinase kinases at a dual phosphorylation motif that has the sequence Thr-X-Tyr, in which phosphorylation at the tyrosine and threonine residues is required for activity. Activated MAP-kinases phosphorylate several transduction targets, including effector protein kinases and transcription factors. Inactivation of MAP-kinases is mediated by dephosphorylation at the Thr-X-Tyr site by dual-specificity phosphatases referred to as MAP-kinase phosphatases. In higher eukaryotes, the physiological role of MAP-kinase signaling has been correlated with cellular events such as proliferation, oncogenesis, development, and differentiation. Accordingly, the ability to regulate signal transduction via these pathways could lead to the development of treatments and preventive therapies for human diseases associated with MAP-kinase signaling, such as cancer.

[0011] Dual-specificity protein tyrosine phosphatases (dual-specificity phosphatases) dephosphorylate both phosphotyrosine and phosphothreonine/serine residues (Walton et al., Ann. Rev. Biochem. 62:101-120, 1993). More than 50 dual-specificity phosphatases that dephosphorylate and inactivate a MAP-kinase have been identified (Shen et al., Proc. Natl. Acad. Sci. USA 98:13613-18 (2001)), including MKP-1 (WO 97/00315; Keyse and Emslie, Nature 59:644-647 (1992)); MKP-2 (WO97/00315); MKP-4, MKP-5, MKP-7, Hb5 (WO 97/06245); PAC1 (Ward et al., Nature 367:651-654 (1994)); HVH2 (Guan and Butch, J. Biol. Chem. 270:7197-7203 (1995)); and PYST1 (Groom et al., EMBO J. 15:3621-3632 (1996)). These dual-specificity phosphatases differ in expression, tissue and subcellular distribution, and specificity for MAP-kinase family members. Expression of certain dual-specificity phosphatases is induced by stress or mitogens, but others appear to be expressed constitutively in specific cell types. The regulation of dual-specificity phosphatase expression and activity is critical for control of MAP-kinase mediated cellular functions, including cell proliferation, cell differentiation and cell survival. For example, dual-specificity phosphatases may function as negative regulators of cell proliferation. It is likely that there are many such dual-specificity phosphatases, with varying specificity with regard to cell type or activation.

[0012] In contrast to the role of most dual-specificity phosphatases to inactivate MAP-kinases, one enzyme, herein referred to as dual-specificity phosphatase 3 (DSP-3), has been reported to have the capability to function as a

selective activator of the JNK MAP-kinase signaling pathway (Shen et al., supra; WO 01/21812). DSP-3 appears also to affect the activity of other kinases involved in the JNK pathway (Shen et al., supra; WO 01/21812). For example, overexpression of DSP-3 leads to activation of MKK4, a MAP-kinase kinase that functions upstream of JNK (Shen et al., supra; Lawler et al., Curr. Biol. 8:1387-90 (1998); Yang et al., Proc. Natl. Acad. Sci. USA 94: 3004-3009 (1997)).

[0013] Activation of JNK is believed to be involved in several physiological processes, including embryonic morphogenesis, cell survival, and apoptosis. A number of JNK signaling pathway substrates have been identified, including c-Jun, ATF2, ELK-1 and others. JNK signaling has also been associated with various disease conditions, such as tumor development, ischemia and reperfusion injury, diabetes, hyperglycemia-induced apoptosis, cardiac hypertrophy, inflammation, and neurodegenerative disorders.

[0014] One non-transmembrane PTP, PTP1B, recognizes several tyrosine-phosphorylated proteins as substrates, many of which are involved in human disease. For example, therapeutic inhibition of PTP1B in the insulin signaling pathway may serve to augment insulin action, thereby ameliorating the state of insulin resistance common in Type II diabetes patients. PTP1B acts as a negative regulator of signaling that is initiated by several growth factor/hormone receptor PTKs, including p210 Bcr-Abl (LaMontagne et al., Mol. Cell Biol. 18:2965-75 (1998); LaMontagne et al., Proc. Natl. Acad. Sci. USA 95:14094-99 (1998)), receptor tyrosine kinases, such as EGF receptor, PDGF receptor, and insulin receptor (IR) (Tonks et al., Curr. Opin. Cell Biol. 13:182-95 (2001)), and JAK family members such as Jak2 and others (Myers et al., J. Biol. Chem. 276:47771-74 (2001)), as well as signaling events induced by cytokines (Tonks and Neel, 2001). Activity of PTP1B is regulated by modifications of several amino acid residues, such as phosphorylation of Ser residues (Brautigan and Pinault, 1993; Dadke et al., 2001; Flint et al., 1993), and oxidation of the active Cys residue in its catalytic motif (Lee et al., 1998; Meng et al., 2002) which is evolutionary conserved among protein tyrosine phosphatases and dual phosphatase family members (Andersen et al., 2001).

[0015] Disruption of the murine PTP1B gene homolog in a knock-out mouse model results in PTP1B^{-/-} mice exhibiting enhanced insulin sensitivity, decreased levels of circulating insulin and glucose, and resistance to weight gain even on a high-fat diet, relative to control animals having at least one functional PTP1B gene (Elchebly et al., Science 283:1544 (1999)). Insulin receptor hyperphosphorylation has also been detected in certain tissues of PTP1B deficient mice, consistent with a PTP1B contribution to the physiologic regulation of insulin and glucose metabolism (Id.). PTP-1B-deficient mice exhibit decreased adiposity (reduced fat cell mass but not fat cell number), increased basal metabolic rate and energy expenditure, and enhanced insulin-stimulated glucose utilization (Klaman et al., 2000 Mol. Cell. Biol. 20:5479). Additionally, altered PTP activity has been correlated with impaired glucose metabolism in other biological systems (e.g., McGuire et al., Diabetes 40:939 (1991); Myerovitch et al., J. Clin. Invest. 84:976 (1989); Sredy et al., Metabolism 44:1074 (1995)), including PTP involvement in biological signal transduction via the insulin receptor (see, e.g., WO 99/46268 and references cited therein).

[0016] An integration of crystallographic, kinetic, and PTP1B-peptide binding assays illustrated the interaction of PTP1B and insulin receptor (IR) (Salmeen et al., Mol. Cell 6:1401-12 (2000)). The insulin receptor (IR) comprises two extracellular α subunits and two transmembrane β subunits. Activation of the receptor results in autophosphorylation of tyrosine residues in both β subunits, each of which contains a protein kinase domain. Extensive interactions that form between PTP1B and insulin receptor kinase (IRK) encompass tandem pTyr residues at 1162 and 1163 of IRK, such that pTyr-1162 is located in the active site of PTP1B (id.). The Asp/Glu-pTyr-pTyr-Arg/Lys motif has been implicated for optimal recognition by PTP1B for IRK. This motif is also present in other receptor PTKs, including Trk, FGFR, and Axl. In addition, this motif is found in the JAK family of PTKs, members of which transmit signals from cytokine receptors, including a classic cytokine receptor that is recognized by the satiety hormone leptin (Touw et al., Mol. Cell. Endocrinol. 160:1-9 (2000)).

[0017] Changes in the expression levels of PTP1B have been observed in several human diseases, particularly in diseases associated with disruption of the normal patterns of tyrosine phosphorylation. For example, the expression of PTP1B is induced specifically by the p210 Bcr-Abl oncoprotein, a PTK that is directly responsible for the initial manifestations of chronic myelogenous leukemia (CML) (LaMontagne et al., Mol. Cell. Biol. 18:2965-75 (1998); LaMontagne et al., Proc. Natl. Acad. Sci. USA 95:14094-99 (1998)). Expression of PTPB1 in response to this oncoprotein is regulated, in part, by transcription factors Sp1, Sp3, and Egr-1 (Fukada et al., J. Biol. Chem. 276:25512-19 (2001)). These transcription factors have been shown to bind to a p210 Bcr-Abl responsive sequence (PRS) in the human PTP1B promoter, located between 49 to -37 base pairs from the transcription start site, but do not appear to mediate certain additional, independent PTP1B transcriptional events, for which neither transcription factor(s) nor transcription factor recognition element(s) have been defined (id.).

[0018] Diabetes mellitus is a common, degenerative disease affecting 5-10% of the human population in developed countries, and in many countries, it may be one of the five leading causes of death. Approximately 2% of the world's population has diabetes, the overwhelming majority of cases (>97%) being type 2 diabetes and the remainder being type 1. In type 1 diabetes, which is frequently diagnosed in children or young adults, insulin production by pancreatic islet beta cells is destroyed. Type 2 diabetes, or "late onset" or "adult onset" diabetes, is a complex metabolic disorder in which cells and tissues cannot effectively use available insulin; in some cases insulin production is also inadequate. At the cellular level, the degenerative phenotype that may be characteristic of late onset diabetes mellitus includes, for example, impaired insulin secretion and decreased insulin sensitivity, i.e., an impaired response to insulin.

[0019] Studies have shown that diabetes mellitus may be preceded by or is associated with certain related disorders. For example, an estimated forty million individuals in the U.S. suffer from late onset impaired glucose tolerance (IGT). IGT patients fail to respond to glucose with increased insulin secretion. Each year a small percentage (5-10%) of IGT individuals progress to insulin deficient non-insulin dependent diabetes (NIDDM). Some of these individuals further

progress to insulin dependent diabetes mellitus (IDDM). NIDDM and IDDM are associated with decreased release of insulin by pancreatic beta cells and/or a decreased response to insulin by cells and tissues that normally exhibit insulin sensitivity. Other symptoms of diabetes mellitus and conditions that precede or are associated with diabetes mellitus include obesity, vascular pathologies, and various neuropathies, including blindness and deafness.

[0020] Type 1 diabetes is treated with lifelong insulin therapy, which is often associated with undesirable side effects such as weight gain and an increased risk of hypoglycemia. Current therapies for type 2 diabetes (NIDDM) include altered diet, exercise therapy, and pharmacological intervention with injected insulin or oral agents that are designed to lower blood glucose levels. Examples of such presently available oral agents include sulfonylureas, biguanides, thiazolidinediones, repaglinide, and acarbose, each of which alters insulin and/or glucose levels. None of the current pharmacological therapies, however, controls the disease over its full course, nor do any of the current therapies correct all of the physiological abnormalities in type 2 NIDDM, such as impaired insulin secretion, insulin resistance, and excessive hepatic glucose output. In addition, treatment failures are common with these agents, such that multi-drug therapy is frequently necessary.

[0021] In certain metabolic diseases or disorders, one or more biochemical processes, which may be either anabolic or catabolic (e.g., build-up or breakdown of substances, respectively), are altered (e.g., increased or decreased in a statistically significant manner) or modulated (e.g., up- or down-regulated to a statistically significant degree) relative to the levels at which they occur in a disease-free or normal subject such as an appropriate control individual. The alteration may result from an increase or decrease in a substrate, enzyme, cofactor, or any other component in any biochemical reaction involved in a particular process. Altered (i.e., increased or decreased in a statistically significant manner relative to a normal state) PTP activity can underlie certain disorders and suggests a PTP role in certain metabolic diseases.

[0022] RNA interference (RNAi) is a polynucleotide sequence-specific, post-transcriptional gene silencing mechanism effected by double-stranded RNA that results in degradation of a specific messenger RNA (mRNA), thereby reducing the expression of a desired target polypeptide encoded by the mRNA (see, e.g., WO 99/32619; WO 01/75164; U.S. Pat. No. 6,506,559; Fire et al., Nature 391:806-11 (1998); Sharp, Genes Dev. 13:139-41 (1999); Elbashir et al. *Nature* 411:494-98 (2001); Harborth et al., *J*. Cell Sci. 114:4557-65 (2001)). RNAi is mediated by doublestranded polynucleotides as also described hereinbelow, for example, double-stranded RNA (dsRNA), having sequences that correspond to exonic sequences encoding portions of the polypeptides for which expression is compromised. RNAi reportedly is not effected by double-stranded RNA polynucleotides that share sequence identity with intronic or promoter sequences (Elbashir et al., 2001). RNAi pathways have been best characterized in Drosophila and Caenorhabditis elegans, but "small interfering RNA" (siRNA) polynucleotides that interfere with expression of specific polypeptides in higher eukaryotes such as mammals (including humans) have also been considered (e.g., Tuschl, 2001 Chembiochem. 2:239-245; Sharp, 2001 Genes Dev. 15:485;

Bernstein et al., 2001 RNA 7:1509; Zamore, 2002 Science 296:1265; Plasterk, 2002 Science 296:1263; Zamore 2001 Nat. Struct. Biol. 8:746; Matzke et al., 2001 Science 293:1080; Scadden et al., 2001 EMBO Rep. 2:1107).

[0023] According to a current non-limiting model, the RNAi pathway is initiated by ATP-dependent, processive cleavage of long dsRNA into double-stranded fragments of about 18-27 (e.g., 19, 20, 21, 22, 23, 24, 25, 26, etc.) nucleotide base pairs in length, called small interfering RNAs (siRNAs) (see review by Hutvagner et al., Curr. Opin. Gen. Dev. 12:225-32 (2002); Elbashir et al., 2001; Nykänen et al., Cell 107:309-21 (2001); Zamore et al., Cell 101:25-33 (2000); Bass, Cell 101:235-38 (2000)). In Drosophila, an enzyme known as "Dicer" cleaves the longer doublestranded RNA into siRNAs; Dicer belongs to the RNase III family of dsRNA-specific endonucleases (WO 01/68836; Bernstein et al., Nature 409:363-66 (2001)). Further according to this non-limiting model, the siRNA duplexes are incorporated into a protein complex, followed by ATPdependent unwinding of the siRNA, which then generates an active RNA-induced silencing complex (RISC) (WO 01/68836). The complex recognizes and cleaves a target RNA that is complementary to the guide strand of the siRNA, thus interfering with expression of a specific protein (Hutvagner et al., supra).

[0024] In C. elegans and Drosophila, RNAi may be mediated by long double-stranded RNA polynucleotides (WO 99/32619; WO 01/75164; Fire et al., 1998; Clemens et al., Proc. Natl. Acad. Sci. USA 97:6499-6503 (2000); Kisielow et al., Biochem. J. 363:1-5 (2002); see also WO 01/92513 (RNAi-mediated silencing in yeast)). In mammalian cells, however, transfection with long dsRNA polynucleotides (i.e., greater than 30 base pairs) leads to activation of a non-specific sequence response that globally blocks the initiation of protein synthesis and causes mRNA degradation (Bass, Nature 411:428-29 (2001)). Transfection of human and other mammalian cells with double-stranded RNAs of about 18-27 nucleotide base pairs in length interferes in a sequence-specific manner with expression of particular polypeptides encoded by messenger RNAs (mRNA) containing corresponding nucleotide sequences (WO 01/75164; Elbashir et al., 2001; Elbashir et al., Genes Dev. 15:188-200 (2001)); Harborth et al., J. Cell Sci. 114:4557-65 (2001); Carthew et al., Curr. Opin. Cell Biol. 13:244-48 (2001); Mailand et al., Nature Cell Biol. Advance Online Publication (Mar. 18, 2002); Mailand et al. 2002 Nature Cell Biol. 4:317).

[0025] siRNA polynucleotides may offer certain advantages over other polynucleotides known to the art for use in sequence-specific alteration or modulation of gene expression to yield altered levels of an encoded polypeptide product. These advantages include lower effective siRNA polynucleotide concentrations, enhanced siRNA polynucleotide stability, and shorter siRNA polynucleotide oligonucleotide lengths relative to such other polynucleotides (e.g., antisense, ribozyme or triplex polynucleotides). By way of a brief background, "antisense" polynucleotides bind in a sequence-specific manner to target nucleic acids, such as mRNA or DNA, to prevent transcription of DNA or translation of the mRNA (see, e.g., U.S. Pat. No. 5,168,053; U.S. Pat. No. 5,190,931; U.S. Pat. No. 5,135,917; U.S. Pat. No. 5,087,617; see also, e.g., Clusel et al., 1993 Nuc. Acids Res. 21:3405-11, describing "dumbbell" antisense oligonucle-

otides). "Ribozyme" polynucleotides can be targeted to any RNA transcript and are capable of catalytically cleaving such transcripts, thus impairing translation of mRNA (see, e.g., U.S. Pat. No. 5,272,262; U.S. Pat. No. 5,144,019; and U.S. Pat. Nos. 5,168,053, 5,180,818, 5,116,742 and 5,093, 246; U.S. 2002/193579). "Triplex" DNA molecules refers to single DNA strands that bind duplex DNA to form a colinear triplex molecule, thereby preventing transcription (see, e.g., U.S. Pat. No. 5,176,996, describing methods for making synthetic oligonucleotides that bind to target sites on duplex DNA). Such triple-stranded structures are unstable and form only transiently under physiological conditions. Because single-stranded polynucleotides do not readily diffuse into cells and are therefore susceptible to nuclease digestion, development of single-stranded DNA for antisense or triplex technologies often requires chemically modified nucleotides to improve stability and absorption by cells. siRNAs, by contrast, are readily taken up by intact cells, are effective at interfering with the expression of specific polypeptides at concentrations that are several orders of magnitude lower than those required for either antisense or ribozyme polynucleotides, and do not require the use of chemically modified nucleotides.

[0026] Importantly, despite a number of attempts to devise selection criteria for identifying oligonucleotide sequences that will be effective in siRNA based on features of the desired target mRNA sequence (e.g., percent GC content, position from the translation start codon, or sequence similarities based on an in silico sequence database search for homologues of the proposed siRNA) it is presently not possible to predict with any degree of confidence which of myriad possible candidate siRNA sequences that can be generated as nucleotide sequences that correspond to a desired target mRNA (e.g., dsRNA of about 18-27 nucleotide base pairs) will in fact exhibit siRNA activity (i.e., interference with expression of the polypeptide encoded by the mRNA). Instead, individual specific candidate siRNA polynucleotide or oligonucleotide sequences must be generated and tested to determine whether interference with expression of a desired polypeptide target can be effected. Accordingly, no routine method exists in the art for designing a siRNA polynucleotide that is, with certainty, capable of specifically altering the expression of a given PTP polypeptide, and thus for the overwhelming majority of PTPs no effective siRNA polynucleotide sequences are presently known.

[0027] Currently, therefore, desirable goals for the rapeutic regulation of biological signal transduction include modulation of PTP (e.g., PTP-1B, DSP-3, SHP-2, KAP, PRL-3, cdc14 or cdc25 or other PTP)-mediated cellular events include, inter alia, inhibition or potentiation of interactions among PTP-binding molecules, substrates and binding partners, or of other agents that regulate PTP activities. Accordingly, a need exists in the art for an improved ability to intervene in the regulation of phosphotyrosine signaling, including regulating PTPs by altering PTP catalytic activity, PTP binding to PTP substrate molecules, and/or PTP-encoding gene expression. An increased ability to so regulate PTPs may facilitate the development of methods for modulating the activity of proteins involved in phosphotyrosine signaling pathways and for treating conditions associated with such pathways. The present invention fulfills these needs and further provides other related advantages.

SUMMARY OF THE INVENTION

[0028] Briefly stated, the present invention provides siRNA compositions and methods for modulating biological signal transduction. In one aspect the present invention provides isolated small interfering RNA (siRNA) polynucleotide, comprising at least one nucleotide sequence selected from the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, and the complementary polynucleotide thereto. The small interfering RNA polynucleotide is capable of interfering with expression of a polypeptide, which polypeptide comprises an amino acid sequence as set forth in a sequence SEQ ID NO: 779, SEQ ID NO 789, SEQ ID NO 791, SEQ ID NO 797, SEQ ID NO 799, SEQ ID NO 801, SEQ ID NO 803, SEQ ID NO 805, SEQ ID NO 807, SEQ ID NO 809, SEQ ID NO 811, or SEQ ID NO 813.

[0029] In certain embodiments, the nucleotide sequence of the siRNA polynucleotide differs by one, two, three or four nucleotides at any of positions 1-19 of a sequence selected from the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493. In other embodiments, the nucleotide sequence of the siRNA polynucleotide differs by at least two, three or four nucleotides at any of positions 1-19 of a sequence selected from the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493. In particular embodiments the invention provides an isolated siRNA polynucleotide comprising a nucleotide sequence selected from SEQ ID NOS: 4, or the complement thereof; from SEQ ID NOS: 100, 105, or the complement thereof; from SEQ ID NOS: 120, 125, or 130; or the complement thereof, from SEQ ID NOS: 140, 145, or 150, or the complement thereof; from SEQ ID NOS: 440 or 445, or the complement thereof; from SEQ ID NOS: 455 or 460; from SEQ ID NO: 465, or the complement thereof; from SEQ ID NOS: 470 or 475, or the complement thereof; from SEQ ID NOS: 480, 485, or 490, or the complement thereof.

[0030] In certain embodiments the invention provides the above siRNA polynucleotides that comprise at least one synthetic nucleotide analogue of a naturally occurring nucleotide. In certain other embodiments, the siRNA polynucleotide is linked to a detectable label, wherein the detectable label is a reporter molecule. In particular embodiments, the reporter molecule is a dye, a radionuclide, a luminescent group, a fluorescent group, or biotin. In other particular embodiments, the fluorescent group is fluorescein isothiocyanate and in other particular embodiments, the detectable label is a magnetic particle.

[0031] The invention also provides a pharmaceutical composition comprising an siRNA polynucleotide selectted from the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, and a physiologically acceptable carrier. In particular embodiments, the the carrier comprises a liposome.

[0032] The invention also provides a recombinant nucleic acid construct comprising a polynucleotide that is capable of directing transcription of a small interfering RNA (siRNA), the polynucleotide comprising: (i) a first promoter; (ii) a second promoter; and (iii) at least one DNA polynucleotide segment comprising at least one nucleotide sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, or a complement thereto, wherein each DNA polynucleotide segment and its complement are operably linked to at least one of the first and second promoters, and wherein the promoters are oriented to direct transcription of the DNA polynucleotide segment and its reverse complement. In certain embodiments, the recombinant nucleic acid construct comprises at least one enhancer that is selected from a first enhancer operably linked to the first promoter and a second enhancer operably linked to the second promoter. In certain other embodiments, the recombinant nucleic acid construct comprises at least one transcriptional terminator that is selected from (i) a first transcriptional terminator that is positioned in the construct to terminate transcription directed by the first promoter and (ii) a second transcriptional terminator that is positioned in the construct to terminate transcription directed by the second promoter. The invention also provides that the siRNA transcribed from the recombinant nucleic acid construct is capable of interfering with expression of a polypeptide, wherein the polypeptide comprises an amino acid sequence as set forth in a sequence selected from SEQ ID NO: 779, SEQ ID NO 789, SEQ ID NO 791, SEQ ID NO 797, SEQ ID NO 799, SEQ ID NO 801, SEQ ID NO 803, SEQ ID NO 805, SEQ ID NO 807, SEQ ID NO 809, SEQ ID NO 811, or SEQ ID NO 813.

[0033] The present invention also provides a recombinant nucleic acid construct comprising a polynucleotide that is capable of directing transcription of a small interfering RNA (siRNA), the polynucleotide comprising at least one promoter and a DNA polynucleotide segment, wherein the DNA polynucleotide segment is operably linked to the promoter, and wherein the DNA polynucleotide segment comprises (i) at least one DNA polynucleotide that comprises at least one nucleotide sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, or a complement thereto; (ii) a spacer sequence comprising at least 4 nucleotides operably linked to the DNA polynucleotide of (i); and (iii) the reverse complement of the DNA polynucleotide of (i) operably linked to the spacer sequence. In certain embodiments, the siRNA polynucleotide transcribed from the recombinant nucleic acid construct comprises an overhang of at least one and no more than four nucleotides, the overhang being located immediately 3' to (iii). In certain particular embodiments, the spacer sequence comprises at least 9 nucleotides. In certain other specific embodiments the spacer sequence comprises two uridine nucleotides that are contiguous with (iii). In one embodiment, the recombinant nucleic acid construct comprises at least one transcriptional terminator that is operably linked to the DNA polynucleotide segment. The invention also provides a host cell that is transformed or transfected with such a recombinant nucleic acid construct as disclosed herein.

[0034] In one embodiment, the invention provides a pharmaceutical composition comprising an siRNA polynucleotide and a physiologically acceptable carrier, wherein the siRNA polynucleotide is selected from (i) an RNA polynucleotide that comprises at least one nucleotide sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493; (ii) an RNA polynucleotide that comprises at least one nucleotide sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, and the complementary polynucleotide thereto; (iii) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by one, two or three nucleotides at any of positions 1-19 of a sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, or (iv) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by two, three or four nucleotides at any of positions 1-19 of a sequence selected from the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493. In certain particular embodiments, the physiologically acceptable carrier comprises a liposome.

[0035] The present invention also provides a method for interfering with expression of a polypeptide, or variant thereof, comprising contacting a subject that comprises at least one cell which is capable of expressing the polypeptide with a siRNA polynucleotide for a time and under conditions sufficient to interfere with expression of the polypeptide, wherein: (a) the polypeptide comprises an amino acid sequence as set forth in a sequence selected from SEQ ID NO: 779, SEQ ID NO 789, SEQ ID NO 791, SEQ ID NO 797, SEQ ID NO 799, SEQ ID NO 801, SEQ ID NO 803, SEO ID NO 805, SEO ID NO 807, SEO ID NO 809, SEO ID NO 811, or SEQ ID NO 813, (b) the siRNA polynucleotide is selected from (i) an RNA polynucleotide which comprises at least one nucleotide sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, (ii) an RNA polynucleotide that comprises at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, and the complementary polynucleotide thereto, (iii) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by one, two or three nucleotides at any of positions 1-19 of a sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, or (iv) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by two, three or four nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493.

[0036] In another embodiment, the invention provides a method for interfering with expression of a polypeptide that comprises an amino acid sequence as set forth in a sequence selected from SEQ ID NO: 779, SEQ ID NO 789, SEQ ID NO 791, SEQ ID NO 791, SEQ ID NO 801, SEQ ID NO 803, SEQ ID NO 805, SEQ ID NO 807, SEQ ID NO 809, SEQ ID NO 811, or SEQ ID NO 813, or a variant of said polypeptide, said method comprising contacting, under conditions and for a time sufficient to interfere with expression of the polypeptide, (i) a subject that comprises at least one cell that is capable of expressing the polypeptide, and (ii) a recombinant nucleic acid construct according to the present invention as described herein.

[0037] In another embodiment, the invention provides a method for identifying a component of a signal transduction pathway comprising: (A) contacting a siRNA polynucleotide and a first biological sample comprising at least one cell that is capable of expressing a target polypeptide, or a variant of said polypeptide, under conditions and for a time sufficient for target polypeptide expression when the siRNA polynucleotide is not present, wherein (i) the target polypeptide comprises an amino acid sequence as set forth in a sequence selected from SEQ ID NO: 779, SEQ ID NO 789, SEQ ID NO 791, SEQ ID NO 797, SEQ ID NO 799, SEQ ID NO 801, SEQ ID NO 803, SEQ ID NO 805, SEQ ID NO 807, SEQ ID NO 809, SEQ ID NO 811, SEQ ID NO 813, SEQ ID NO 823, SEQ ID NO 825, or SEQ ID NO:827; (2) the siRNA polynucleotide is selected from (i) an RNA polynucleotide which comprises at least one nucleotide sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, (ii) an RNA polynucleotide that comprises at least one nucleotide sequence selected from SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, and the complementary polynucleotide thereto; (iii) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by one, two or three nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493, (iv) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by two, three or four nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493; and (B) comparing a level of phosphorylation of at least one protein that is capable of being phosphorylated in the cell with a level of phosphorylation of the protein in a control sample that has not been contacted with the siRNA polynucleotide, wherein an altered level of phosphorylation of the protein in the presence of the siRNA polynucleotide relative to the level of phosphorylation of the protein in an absence of the siRNA polynucleotide indicates that the protein is a component of a signal transduction pathway. The invention also provides a small interfering RNA (siRNA) polynucleotide, comprising an RNA polynucleotide which comprises at least one nucleotide sequence selected from SEQ ID NOS:4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, or 490-493. Certain further embodiments relate to isolated siRNA polynucleotides that comprise nucleotide sequences having the above recited SEQ ID NOS, including compositions and methods for producing and therapeutically using such siRNA.

[0038] These and other embodiments of the present invention will become apparent upon reference to the following detailed description and attached drawings. All references disclosed herein are hereby incorporated by reference in their entireties as if each was incorporated individually. Also incorporated by reference are co-pending application Ser. No. _____ and Ser. No. _____ (attorney docket numbers 200125.441 and 200125.448, respectively), which have been filed concurrently.

BRIEF DESCRIPTION OF THE DRAWINGS

[0039] FIG. 1 presents an immunoblot analysis of the expression of MKP-1 polypeptide in HeLa cells co-transfected with sequence-specific siRNA polynucleotides (MKPsi.1 (MKP.1, SEQ ID NO:_____), lanes 1-3; MKPsi.2 (MKP.2, SEQ ID NO:_____), lanes 4-6) and a non-specific sequence siRNA (CD45si.1, lanes 7-9). The immunoblot of HeLa cell extracts was probed with an anti-MKP-1 antibody (upper). A second SDS-PAGE gel in which the HeLa cell extracts were separated was stained with Coomassie Blue (lower).

[0040] FIG. 2 shows an immunoblot analysis of 292-HEK cell lysates from cells co-transfected with FLAG®-DSP-11, FLAG®-DSP-18, FLAG®-DSP-3, and FLAG®-cdc14b expression vectors and siRNAs specific for DSP-11 or DSP-18. The presence of each polypeptide was detected using an anti-FLAG® antibody (Sigma-Aldrich, St. Louis, Mo.). The upper immunoblot shows the level of expression of FLAG®-DSP-11 in untransfected 293-HEK cells (lane 1); 293-HEK cells transfected with FLAG®-DSP-11 vector DNA only (buffer) (lane 2), siRNA DSP11.2 (lane 3), siRNA DSP11.4 (lane 4), siRNA DSP18.2 (lane 5), and siRNA DSP18.2 (lane 6); and the level of expression of 293-HEK cells transfected with FLAG®-DSP-18 vector DNA only (buffer) (lane 7); 293-HEK cells co-transfected with siRNA DSP11.2 (lane 8), siRNA DSP11.4 (lane 9), siRNA DSP18.2 (lane 10), and siRNA DSP18.2 (lane 11). The lower immunoblot shows the level of FLAG®-DSP-3 in untransfected 293-HEK cells (lane 1); 293-HEK cells transfected with FLAG®-DSP-3 vector DNA only (buffer) (lane 2); 293-HEK cells co-transfected with siRNA DSP11.2 (lane 3), siRNA DSP11.4 (lane 4), siRNA DSP18.2 (lane 5), and siRNA DSP18.2 (lane 6); and the level of expression of FLAG®-cdc14b in 293-HEK cells transfected with FLAG®-cdc14b vector DNA only (buffer) (lane 7); 293-HEK cells co-transfected with siRNA DSP11.2 (lane 8), siRNA DSP11.4 (lane 9), siRNA DSP18.2 (lane 10), and siRNA DSP18.2 (lane 11).

[0041] FIG. 3 shows the effect on JNK activation by sequence-specific siRNA interference of DSP-3 polypeptide expression. HeLa cells were co-transfected with a DSP-3 recombinant expression vector and DSP3.1 siRNA (SEQ ID NO:1) or 60 pmoles (100 nM final) CD45.2 (SEQ ID NO:_____). After transfection, cells were stimulated with either tumor necrosis factor-alpha (TNF- α) or epidermal growth factor (EGF) or were unstimulated (Unstim.).

[0042] FIG. 4 shows the effect on JNK activation by sequence-specific siRNA interference of DSP-3 polypeptide expression. HeLa cells were co-transfected with a DSP-3 recombinant expression vector and DSP3.1 siRNA (SEQ ID NO:_____) or 60 pmoles (100 nM final) CD45.2 (SEQ ID NO:_____). After transfection, cells were stimulated with sorbitol.

[0043] FIG. 5 presents an immunoblot analysis of ERK phosphorylation in HeLa cells co-transfected with a DSP-3 recombinant expression vector and DSP-3 specific siRNA DSP3.1, non-specific CD45.2 siRNA, or siRNA annealing buffer and then stimulated with TNF-α, EGF, sorbitol, and anisomycin. Lane 1: unstimulated cells transfected with DSP3.1 siRNA; lane 2: unstimulated cells transfected with CD45.2 siRNA; lane 3: cells transfected with DSP3.1 siRNA and stimulated with TNF- α ; lane 4: cells transfected with CD45.2 siRNA and stimulated with TNF-α; lane 5: cells transfected with DSP3.1 siRNA and stimulated with EGF; lane 6: cells transfected with CD45.2 siRNA and stimulated with EGF; lane 7: unstimulated cells transfected with CD45.2 siRNA; lane 8: unstimulated cells transfected with siRNA annealing buffer; lane 9: cells transfected with DSP3.1 siRNA and stimulated with sorbitol; lane 10: cells transfected with CD45.2 siRNA and stimulated with sorbitol; lane 11; cells transfected with siRNA annealing buffer and stimulated with sorbitol; lane 12: cells transfected with DSP3.1 siRNA and stimulated with anisomycin; lane 13: cells transfected with CD45.2 siRNA and stimulated with anisomycin; lane 14: cells transfected with siRNA annealing buffer and stimulated with anisomycin.

[0044] FIG. 6 shows an immunoblot analysis of FLAG®-tagged cdc14a expression in 293-HEK cells co-transfected with cdc14a.2 (lane 3); cdc14a.3 (lane 4); cdc14a.4 (land 5); cdc14a.5 (lane 6); DSP3.1 (lane 7); DSP3.2 (lane 8); cdc14b.3 (lane 9); cdc14b.4 (lane 10); MKP.2 (lane 11); CD45.3 (lane 12); no siRNA (lane 2). Untransfected cells were prepared as a control (lane 1). Expression was detected using an anti-FLAG® antibody (Sigma-Aldrich).

[0045] FIG. 7 presents an immunoblot of expression of FLAG®-tagged dual specificity phosphatases in 293-HEK cells that were co-transfected with cdc14a.3 siRNA (denoted by +). Lanes 2 and 3: expression of FLAG®-tagged cdc14a; lanes 4 and 5: expression of FLAG®-tagged DSP-3; lanes 6 and 7: expression of FLAG®-tagged cdc14b; lanes 8 and 9: FLAG®-tagged DSP-11. The immunoblot to the right is an over-exposure of the immunoblot on the left to detect low concentrations of expressed polypeptides.

[0046] FIG. 8 shows an immunoblot analysis of FLAG®-tagged cdc14b expression in 293-HEK cells co-transfected with cdc14b.3 (lane 3); cdc14b.4 (lane 4); cdc14a.3 (land 5); cdc14a.5 (lane 6); DSP3.1 (lane 7); DSP3.2 (lane 8); MKP.2 (lane 9); CD45.3 (lane 10); no siRNA (lane 2). Untransfected cells were prepared as a control (lane 1). Expression was detected using an anti-FLAG® antibody (Sigma-Aldrich).

[0047] FIG. 9 presents an immunoblot of expression of FLAG®-tagged dual specificity phosphatases in 293-HEK cells co-transfected with either cdc14a or cdc14b specific siRNAs. Expression of the phosphatases was detected with an anti-FLAG® antibody. 293-HEK cells were transfected as follows: no expression vector or siRNA (lane 1); FLAG®-tagged cdc14b only (lane 2); FLAG®-tagged cdc14b and cdc14b.3 siRNA (lane 3); FLAG®-tagged cdc14b and cdc14b.4 (lane 5); FLAG®-tagged DSP-3 only (lane 5); FLAG®-tagged DSP-3 and cdc14b.3 siRNA (lane 6); FLAG®-tagged DSP3 and cdc14b.4 siRNA (lane 7); FLAG®-tagged DSP-3 and cdc14a.5 siRNA (lane 8); FLAG®-tagged DSP-11 only (lane 9); FLAG®-tagged DSP-11 and cdc14b.3 siRNA (land 10); FLAG®-tagged DSP-11 and cdc14b.4 siRNA (lane 11); and FLAG®-tagged DSP-11 and cdc14a.5 siRNA.

[0048] FIG. 10 depicts the expression of cdc14b polypeptide in HeLa cells co-transfected with cdc14b.4 siRNA detected by immunocytochemistry (top right, 10x magnification; bottom right, 40x magnification) and in the absence of a specific siRNA (top left, 10x magnification; bottom right, 40x magnification).

[0049] FIG. 11 depicts an immunoblot of the effect on endogenous expression of murine PTP1B by siRNAs specific for the murine PTP1B or the human PTP1B polynucleotide sequences. Expression was detected using a murine anti-PTP1B monoclonal antibody. Data are presented for two different clones of C57B16 #3 murine cells. Both clones were transfected with mPTP1B1.1 siRNA (lanes 3 and 8); MPTP1B1.2 (lanes 4 and 9); mPTP1B1.3 (lanes 5 and 10). One clone, C57B16 #3 clone 3, was transfected with hPTP1B1.1 (lane 6). Lane 2: untransfected C57B16 #3, clone 3; lane 7: untransfected C57B16 #3, clone 10.

[0050] FIG. 12 presents an extended consensus cDNA sequence encoding prototypical DSP-18 (DSP-18pr) (FIG. 12A) [SEQ ID NO:_____] and the deduced DSP-18pr amino acid sequence (FIG. 12B) [SEQ ID NO:_____]. In FIG. 12A, initiating methionine (ATG) and stop (TGA) codons and intron/exon splice junctions are depicted in bold type with the splice donor sequences in bold without underscore, and the splice acceptor sequences in bold with underscore. In FIG. 12B, initiating methionine and the phosphatase active site are depicted in bold type.

[0051] FIG. 13 presents nucleotide and amino acid sequences for a DSP-18 isoform, DSP-18a. FIG. 13A presents a cDNA sequence for DSP-18a [SEQ ID NO:____], with the start (ATG) and stop (TGA) codons and intron/exon splice junctions indicated in bold; intron/exon splice junctions are depicted in bold type with the splice donor sequences in bold without underscore and the splice acceptor sequences in bold with underscore. FIG. 13B presents the amino acid sequence of the DSP-18a polypeptide [SEQ ID NO:_____] encoded by SEQ ID NO:_____, with the phosphatase active site depicted in bold type.

[0052] FIG. 14 presents nucleotide and amino acid sequences for a DSP-18 isoform, DSP-18b. FIG. 14A presents a cDNA sequence for DSP-18b [SEQ ID NO:____], with the start (ATG) and stop (TGA) codons and intron/exon splice junctions indicated in bold; intron/exon splice junctions are depicted in bold type with the splice donor sequences in bold without underscore and the splice acceptor sequences in bold with underscore. FIG.

14B presents the amino acid sequence of the DSP-18b polypeptide [SEQ ID NO:____] encoded by SEQ ID NO:____, with the phosphatase active site depicted in bold type.

[0053] FIG. 15 presents nucleotide sequences for DSP-18 isoforms, DSP-18c and DSP-18d. FIG. 15A presents a cDNA sequence for DSP-18c [SEQ ID NO:_____ with the start (ATG) and stop (TGA) codons and intron/exon splice junctions indicated in bold. FIG. 15B presents a cDNA sequence for DSP-18d [SEQ ID NO:____], with the start (ATG) and stop (TGA) codons and intron/exon splice junctions indicated in bold. DSP-18c [SEQ ID NO:____] encoded by SEQ ID NO:____, and DSP-18d [SEQ ID NO:____] encoded by SEQ ID NO:____, both share the 181 amino acid sequence encoded by the open reading frame of DSP-18a (see FIG. 15).

[0054] FIG. 16 presents nucleotide and amino acid sequences for DSP-18 isoforms, DSP-18e and DSP-18f. FIG. 16A presents a cDNA sequence for DSP-18e [SEQ ID NO:____], with the start (ATG) and stop (TGA) codons and intron/exon splice junctions indicated in bold. FIG. 16 presents the amino acid sequence of DSP-18e polypeptide [SEQ ID NO:____] encoded by SEQ ID NO:____, with the phosphatase active site sequence in boldface type.

[0055] FIG. 17A presents nucleotide and amino acid sequences for DSP-18f. FIG. 17A presents a cDNA sequence for DSP-18f [SEQ ID NO: _____], with the start (ATG) and stop (TGA) codons and intron/exon splice junctions indicated in bold. FIG. 17B presents the amino acid sequence of DSP-18f polypeptide [SEQ ID NO: _____] encoded by SEQ ID NO: _____, with the phosphatase active site sequence in boldface type.

[0056] FIG. 18 represents an immunoblot of cleavage of poly(ADP-ribose) polymerase (PARP) in HeLa cells transfected with cell division cycle protein sequence specific siRNA polynucleotides (10 nM). The upper immunoblot was probed with an antibody that specifically binds to cleaved PARP, and the lower immunoblot was probed with an anti-PARP antibody. The siRNA polynucleotides transfected into the HeLa cells were as follows: lanes 1 and 2, no siRNA; lanes 3 and 4, cdc14a.5; lanes 5 and 6, cdc14b.4; lanes 7 and 8 Cdc25A.2; lanes 9 and 10, Cdc25B.4; and lanes 11 and 12, Cdc25C.1.

[0057] FIG. 19 depicts an immunoblot analysis of the expression of human PTP-1 B co-transfected into 1BKO+HIR murine fibroblasts with human PTP-1B siRNA hairpin vectors. Expression was detected with an anti-human PTP1B antibody (h1B) (lower portion of immunoblot). As a protein expression control, cell lysates were probed with an anti-human insulin receptor (IR) antibody (upper portion of immunoblot).

[0058] FIG. 20 illustrates insulin-induced activation of PKB/Akt in HepG2 cells following ablation of TC45 by RNA interference. FIG. 20A represents an immunoblot of serum-deprived Rat-1 and HEPG2 cells that were exposed to varying concentrations of insulin (INS) as shown. The insulin receptor (IR) was immunoprecipitated from cell lysates with an anti-IR- β antibody followed by immunoblotting with an anti-phosphotyrosine antibody (pY) (top panel); an anti-pYpY^{1162/1163}-IR- β antibody (middle panel); and an anti-IR β antibody. FIG. 20B represents an immu-

noblot of HepG2 cell lysates prepared from cells that were untransfected (control) or transfected with TCPTP1 siRNA (SEQ ID NO:_____) (+siRNA). The lysates were immunoblotted with an anti-phospho-PKB/Akt antibody (p-AKT) (first immunoblot); anti-PKB/Akt antibody (AKT) (second immunoblot); anti-TC45 (TC45) antibody (third immunoblot); and an anti-PTP1B antibody (PTP1B). FIG. 20C represents a densitometric analysis of the gel image to illustrate the ratio of phosphorylated PKB/Akt to total PKB/Akt.

[0059] FIG. 21 provides an immunoblot indicating that tyrosine phosphorylated IR-β is a substrate of TC45. HepG2 cells overexpressing wild-type (WT) or substrate trapping mutant (DA) forms of PTP1B (1B) and TC45 were either not treated with insulin (-INS) or stimulated with insulin for 5 minutes (+INS), lysed, separated by SDS-PAGE, and immunoprecipitated with anti-PTP1B antibody (FG6) or anti-TC45 antibody (CF4). The immunoprecipitates were immunoblotted with an anti-IR-β antibody (top panel, FIG. 21A); anti-PTP1B antibody FG6 (middle panel, FIG. 21A); and anti-TCPTP antibody CF4 (bottom panel, FIG. 21A). FIG. 21B depicts immunoblots of HepG2 cells that were serumstarved and untransfected (control) or transfected with TC45 siRNA (100 nM) and then stimulated with 10 nM insulin (INS) for the indicated times. The insulin receptor was immunoprecipitated from cell lysates with an anti-IR-β antibody, which was then immunoblotted with the following antibodies: anti-phosphotyrosine (p-Tyr) (first immunoblot); anti-p Y^{972} -IR- β (second immunoblot); anti-p $Y^{1162/1163}$ -IR-β (third immunoblot); and anti-IR-β (fourth immunoblot). FIG. 21C presents densitometric analyses of the gel image to show the ratio of phosphorylated IR-β to total IR-β for total phosphotyrosine (top panel); phosphorylation of Tyr 972 (middle panel); and phosphorylation of the activation loop tyrosines 1162 and 1163 (lower panel).

[0060] FIG. 22 presents the results of an ELISA in which the level of insulin receptor (IR) phosphorylated tyrosine was measured in 293-HEK HIR cells transfected with 0, 0.5, 3, or 10 nM hPTP1B1.3 (H1.3, SEQ ID NO:______) (FIG. 22A) or mPTP1B1.1b (M1.1, SEQ ID NO:______) (FIG. 22B) siRNAs. The level of expression of human PTP1B in the cells was compared by immunoblot (see tables to right of each figure).

[0061] FIG. 23 depicts the results of an ELISA in which the level of insulin receptor (IR) phosphorylated tyrosine was measured in 293-HEK HIR cells transfected with 0, 0.5, 3, or 10 nM siRNAs. The siRNA polynucleotides transfected into the cells included hPTP1B1.2 (H1.2, SEQ ID NO:_____); hPTP1B1.3 (H1.3, SEQ ID NO:_____); mPTP1B1.1b (M1.1, SEQ ID NO:_____); and rPTP1B1.2 (R1.2, SEQ ID NO:_____). Seventy-two hours after transfection, cells were exposed to insulin for 7 minutes at the designated concentrations. Cell lysates were prepared and coated onto 96-well plates and probed with an anti-pY-IR-β antibody.

[0062] FIG. 24 depicts the results of an ELISA in which the level of insulin receptor (IR) phosphorylated tyrosine was measured in 293-HEK HIR cells transfected with 0, 0.5, 3, or 10 nM hTCPTP1.4 siRNA (TC1.4, SEQ ID NO:_____) (FIG. 24A) and mPTP1B1.1b siRNA (M1.1, SEQ ID NO:_____) (FIG. 24B). Seventy-two hours after transfection, cells were exposed to insulin for 7 minutes at

the designated concentrations. Cell lysates were prepared and coated onto 96-well plates and probed with an anti-pY-IR- β antibody.

[0063] FIG. 25 represents ELISA data from three separate experiments that represent the level of insulin receptor phosphorylation in cells transfected with hPTP1B1.3 and stimulated with 50 nM insulin (Ins). Each data point represents the average optical density measured in duplicate wells.

[0064] FIG. 26 illustrates an MTT assay comparing proliferation of HCT-116 cells transfected with siRNAs specific for DSP-3 (dsp3.1 (SEQ ID NO:_____) and dsp3.4 (SEQ ID NO:_____)); cdc14a (a.3 (SEQ ID NO:_____) and a.5 (SEQ ID NO:_____)); SHP-2 (shp2.1 (SEQ ID NO:_____); and DHFR (DHFR.1 (SEQ ID NO:_____), and DHFR (DHFR.1 (SEQ ID NO:_____). As a control, HCT-116 cells were transfected with nonspecific siRNA (scr.2 (SEQ ID NO:_____)). Each bar represents the average optical density for six wells.

[0065] FIG. 27 illustrates an MTT assay comparing proliferation of T47D cells transfected with siRNAs specific for DSP-3 (dsp3.1 (SEQ ID NO:_____) and dsp3.4 (SEQ ID NO:_____); cdc14a (Cdc14a.3 (SEQ ID NO:_____) and Cdc14a.5 (SEQ ID NO:_____); SHP-2 (shp2.1 (SEQ ID NO:_____); and DHFR (DHFR.1 (SEQ ID NO:_____)). As a control, T47D cells were transfected with nonspecific siRNA (scr.2 (SEQ ID NO:_____)).

[0066] FIG. 28 represents an immunoblot of cleavage of PARP in HCT-116 cells (FIG. 28A) and T47D (FIG. 28B) transfected with buffer only (lane 1); (scrb1.2 (SEQ ID NO: _____) (lane 2); DSP3.1 (SEQ ID NO: _____) (lane 3); DSP3.4 (SEQ ID NO: _____) (lane 4); and DHFR.1 (lane 5).

[0067] FIG. 29 presents nucleotide and amino acid sequences for DSP-13. FIG. 29A presents a cDNA sequence for DSP-13 [SEQ ID NO:_____], with the start (ATG) and stop (TGA) codons indicated in bold and underlined. FIG. 29B presents the amino acid sequence of the DSP-13 polypeptide [SEQ ID NO:_____] encoded by SEQ ID NO:_____.

[0068] FIG. 30 presents nucleotide and amino acid sequences for DSP-14. FIG. 30A presents a cDNA sequence for DSP-14 [SEQ ID NO:_____], with the start (ATG) and stop (TGA) codons indicated in bold and underlined. FIG. 30B presents the amino acid sequence of the DSP-14 polypeptide [SEQ ID NO:_____] encoded by SEQ ID NO:_____.

DETAILED DESCRIPTION OF THE INVENTION

[0069] The present invention is directed in part to the unexpected discovery of short RNA polynucleotide sequences that are capable of specifically modulating expression of a desired polypeptide, such as a DSP-3, SHP-2, KAP, PRL-3, cdc14 or cdc25 polypeptide, or a variant of any such polypeptide. Without wishing to be bound by theory, the RNA polynucleotides of the present invention specifically reduce expression of a desired target polypeptide through recruitment of small interfering RNA (siRNA) mechanisms. In particular, and as described in

greater detail herein, according to the present invention there are provided compositions and methods that relate to the surprising identification of certain specific RNAi oligonucleotide sequences of 19, 20, 21, 22, 23, 24, 25, 26 or 27 nucleotides that can be derived from corresponding polynucleotide sequences encoding the desired DSP-3, SHP-2, KAP, PRL-3, cdc14, cdc25, or other specified target polypeptide. These sequences cannot be predicted through any algorithm, sequence alignment routine, or other systematic paradigm, but must instead be obtained through generation and functional testing for RNAi activity of actual candidate oligonucleotides, such as those disclosed for the first time herein.

[0070] In preferred embodiments of the invention, the siRNA polynucleotide interferes with expression of a DSP-3, SHP-2, KAP, PRL-3, cdc14, cdc25, or other herein specified target polypeptide or a variant thereof, and comprises a RNA oligonucleotide or RNA polynucleotide uniquely corresponding in its nucleotide base sequence to the sequence of a portion of a target polynucleotide encoding the target polypeptide, for instance, a target mRNA sequence or an exonic sequence encoding such mRNA. Hence, according to non-limiting theory, the siRNA polynucleotides of the present invention direct sequence-specific degradation of mRNA encoding a desired DSP-3, SHP-2, KAP, PRL-3, cdc14 or cdc25 target polypeptide, the expression of which is consequently compromised. As also described herein, certain embodiments of the invention relate to siRNA polynucleotides that specifically interfere with expression of PTPs that are dual specificity phosphatases, including DSP-3, DSP-11, DSP-13, DSP-14, and DSP-18; certain other embodiments relate to RNAi interference with expression of the MAP kinase kinase (MKK) target polypeptide MKK4; certain other embodiments relate to RNAi interference with expression of target polypeptides that interact with chemotherapeutic agents, for example, the target polypeptides dihydrofolate reductase (DHFR), thymidylate synthetase, and topoisomerase I. The invention relates in preferred embodiments to siRNA polynucleotides that interfere with expression of specific polypeptides in mammals, which in certain particularly preferred embodiments are humans and in certain other particularly preferred embodiments are non-human mammals.

[0071] Exemplary sequences for the target polypeptides described herein include, for instance, DSP-3 (WO 00/60092; SEQ ID NO:24 encoded by SEQ ID NO:23); cdc14A (e.g., GenBank Accession Nos. AF122013, AF064102, AF064103; Li et al., 1997 J. Biol. Chem. 272:29403; U.S. Pat. No. 6,331,614; e.g., SEQ ID NO:34 encoded by SEQ ID NO:33) or cdc14B (e.g., GenBank Accession Nos. AF064104, AF064105, AF023158; Li et al., 1997 J. Biol. Chem. 272:29403; e.g., SEQ ID NO:36 encoded by SEQ ID NO:35); cdc25A ((e.g., GenBank Accession Nos. NM_001789, AF527417, NM_133571); cdc25B (e.g., GenBank Accession Nos. NM_133572, NM_023117, NM_021872; NM_021872; M81934); and cdc25C (e.g., GenBank Accession Nos. NM_001790, NM_022809); PTPε (e.g., Genbank Accession Nos. NM_006504 (SEQ ID NOS: and NM 130435(SEO ID NOS:)); KAP (e.g., Genbank Accession No. L27711; Hannon et al., Proc. Natl. Acad. Sci. USA 91:1731-35 (1994); Demetrick et al., Cytogenet. Cell Genet. 69:190-92 (1995)); PRL-3 (e.g., Zhao et al., Genomics 35:172-81 (1996); Genbank Accession Nos.

(NM_003479 (SEQ ID NOS: NM_080392 (SEQ ID NOS: NM 080391 (SEQ ID NOS: NM_032611 (SEQ ID NOS: and NM_007079 (SEQ ID NOS: SHP-2 (GenBank Accession Nos. D13540 (SEQ ID NOS: _); L03535 (SEQ ID NOS:); L07527 (SEQ ID NOS:); X70766 (SEQ ID NOS:); L08807 (SEQ ID NO:); 78088); S39383 (SEQ ID NO: (SEQ ID NOS:); D84372 (SEQ ID NOS:); U09307); CD45 (e.g., (Charbonneau (SEO ID NOS: et al., Proc. Natl. Acad. Sci. USA 85:7182-86 (1988); Genbank Accession Nos. NM_080922 (SEQ ID NOS:), NM_080921 (SEQ ID NOS: NM_002838 (SEQ ID NOS: NM_080923) (SEQ ID NOS:); GenBank Ace. No. XM_16748; e.g., SEQ ID NO: 32 encoded by SEQ); DSP-11 (WO ID NO:31); SEQ ID NOS: 01/05983, SEQ ID NO:26 encoded by SEQ ID NO:25); DSP-18 (U.S. application Ser. No. 10/151,320, SEQ ID NO:28 encoded by SEQ ID NO:27); DSP-13 (U.S. application Ser. No. 09/775,925; SEQ ID NO: encoded by _); DSP-14 (U.S. application Ser. No. SEQ ID NO: 09/847,519; SEQ ID NO:_ encoded by SEQ ID); WO 01/46394); MKP-1 (WO 97/00315; Keyse et al., 1992 Nature 59:644; SEQ ID NO:30 encoded by SEQ ID NO:29). According to the contemplated invention, the siRNA polynucleotide expressly does not consist of a CDC14a.5 polynucleotide having a sequence set forth in SEQ ID NO:10 (Mailand et al., 2002 Nature Cell Biol. 4:317).

[0072] In certain embodiments of the invention, an siRNA polynucleotide interferes with expression of a component of a signaling transduction pathway, for example, components of the JNK signaling transduction pathway such as MKK4 (e.g., GenBank Accession Nos. L36870 (SEQ ID NO: and), NM_009157, and NM_009157; SEO ID NO: encoded by SEQ ID NO:)) and MKK7 (e.g., GenBank Accession Nos. AF013588 (SEQ ID encoded by SEQ ID NO: AF026216, and to related compositions and methods. (See also Shen et al., Proc. Natl. Acad. Sci. USA 98:13613-18 (2001)). In certain other embodiments of the invention, the siRNA polynucleotide interferes with expression of a cellular polypeptide or enzyme that is associated with a cellular malfunction or defect (e.g., in a cancer or malignancy, an enzyme that is overexpressed or constitutively expressed and is associated with cell survival, proliferation, apoptosis, cell division, and differentiation). For example, the siRNA polynucleotide may comprise a sequence specific for dihydrofolate reductase (DHFR) (e.g., GenBank Accession No: NM_000791; SEQ ID NO: _ ____ encoded by SEQ ID)); thymidylate synthetase e.g., GenBank Accession No: NM_001071 (SEQ ID NO:); topoisomerase I (e.g., GenBank SEQ ID NO: Accession No: J03250; SEQ ID NO: encoded by SEQ ID NO:_ _)); IkappaB kinase (IKK) alpha (e.g., GenBank Accession No. AF080157; SEQ ID NO: encoded by SEO ID NO:); GenBank Accession No. AF009225; GenBank Accession No. AF012890); IKKbeta e.g., GenBank Accession No. AF080158; SEQ ID NO: _); GenBank Accesencoded by SEQ ID NO: sion No. AF031416; GenBank Accession No. AF029684);

or IKKgamma e.g., GenBank Accession No. AF074382; SEQ ID NO: _____ encoded by SEQ ID NO: _____); GenBank Accession No. AF091453).

[0073] In another preferred embodiment, the siRNA polynucleotides provided interfere with expression of DSP-3, SHP-2, CD45, PTP ϵ , KAP, cdc14a, cdc14b, cdc25A, cdc25B, cdc25C, and PRL-3. According to non-limiting theory, the siRNA polynucleotides of the present invention direct sequence-specific degradation of mRNA encoding a PTP such as SHP2, PTP ϵ , or a dual specificity phosphatase (e.g., DSP-3, KAP, cdc14a, cdc14b, cdc25A, cdc25B, cdc25C, CD45, or PRL-3) by a mechanism known as RNA interference (RNAi). The invention is not intended, however, to be so limited, and certain embodiments relate to RNA interference of other PTPs and dual specificity phosphatases (e.g., DSP-11, DSP-13, DSP-14, and DSP-18), and to interference with expression of other polypeptides and components of signal transduction pathways including mitogen activated protein (MAP) kinases, which include a MAP kinase kinase (e.g., MAPKKK or MEKK) that activates a MAP/ERK kinase (e.g., MAPKK or MEK), which then stimulates a phosphorylation-dependent increase in the activity of the MAP kinase. Upon activation, a MAP kinase can phosphorylate a variety of intracellular targets including transcription factors, transcriptional adaptor proteins, membrane and cytoplasmic substrates, and other protein kinases. In certain preferred embodiments, a siRNA polynucleotide interferes with expression of a MAP kinase kinase that is a component of the JNK signal transduction pathway, for example, MKK4 or MKK7. In other preferred embodiments, a siRNA polynucleotide interferes with expression of a cellular polypeptide or enzyme that is associated with a cellular malfunction or defect in cancer or malignancy, and which may be overexpressed or constitutively expressed in the tumor cell.

[0074] In addition, other preferred polypeptides include polypeptides that are targets of chemotherapeutic agents or drugs. Examples of chemotherapeutic target polypeptides include enzymes in the folate metabolic pathway, for example, thymidylate synthetase, which is a target of fluoropyrmidines. Another enzyme in this pathway is dihydrofolate reductase (DHFR), which is targeted by antifolate agents, such as methotrexate. DNA processing enzymes, including topoisomerase I and topoisomerase II, are also targets of chemotherapeutic agents. Other examples of chemotherapeutic target polypeptides include microtubule polypeptides, which are chemotherapeutic targets of taxanes and vinca alkaloids. According to non-limiting theory, these chemotherapeutic target polypeptides may become resistant to a drug or agent, that is, resistance may be manifested by overexpression or constitutive expression of the chemotherapeutic target polypeptide in a target cell. The overexpression of such a target polypeptide may be reduced by introducing a specific siRNA polynucleotide into the cell. In certain embodiments of the invention, a siRNA polynucleotide interferes with expression of such chemotherapeutic target polypeptides. For example, siRNA polynucleotides of the present invention that interfere with expression of a chemotherapeutic target polypeptide comprise sequences specific for dihydrofolate reductase (DHFR), thymidylate synthetase, topoisomerase I, and IKKgamma.

[0075] SiRNA Polynucleotides

[0076] As used herein, the term "siRNA" means either: (i) a double stranded RNA oligonucleotide, or polynucleotide, that is 18 base pairs, 19 base pairs, 20 base pairs, 21 base pairs, 22 base pairs, 23 base pairs, 24 base pairs, 25 base pairs, 26 base pairs, 27 base pairs, 28 base pairs, 29 base pairs or 30 base pairs in length and that is capable of interfering with expression and activity of a PTP-1B polypeptide, or a variant of the PTP-1B polypeptide, wherein a single strand of the siRNA comprises a portion of a RNA polynucleotide sequence that encodes the PTP-1B polypeptide, its variant, or a complementary sequence thereto; (ii) a single stranded oligonucleotide, or polynucleotide of 18 nucleotides, 19 nucleotides, 20 nucleotides, 21 nucleotides, 22 nucleotides, 23 nucleotides, 24 nucleotides, 25 nucleotides, 26 nucleotides, 27 nucleotides, 28 nucleotides, 29 nucleotides or 30 nucleotides in length and that is either capable of interfering with expression and/or activity of a target polypeptide such as DSP-3, SHP-2, KAP, PRL-3, cdc14 or cdc25, or a variant of the target polypeptide, or that anneals to a complementary sequence to result in a dsRNA that is capable of interfering with target polypeptide expression, wherein such single stranded oligonucleotide comprises a portion of a RNA polynucleotide sequence that encodes the target polypeptide, its variant, or a complementary sequence thereto; or (iii) an oligonucleotide, or polynucleotide, of either (i) or (ii) above wherein such oligonucleotide, or polynucleotide, has one, two, three or four nucleic acid alterations or substitutions therein.

[0077] A siRNA polynucleotide is a RNA nucleic acid molecule that mediates the effect of RNA interference, a post-transcriptional gene silencing mechanism. A siRNA polynucleotide preferably comprises a double-stranded RNA (dsRNA) but is not intended to be so limited and may comprise a single-stranded RNA (see, e.g., Martinez et al. Cell 110:563-74 (2002)). A siRNA polynucleotide may comprise other naturally occurring, recombinant, or synthetic single-stranded or double-stranded polymers of nucleotides (ribonucleotides or deoxyribonucleotides or a combination of both) and/or nucleotide analogues as provided herein (e.g., an oligonucleotide or polynucleotide or the like, typically in 5' to 3' phosphodiester linkage). Accordingly it will be appreciated that certain exemplary sequences disclosed herein as DNA sequences capable of directing the transcription of the subject invention siRNA polynucleotides are also intended to describe the corresponding RNA sequences and their complements, given the well established principles of complementary nucleotide base-pairing. A siRNA may be transcribed using as a template a DNA (genomic, cDNA, or synthetic) that contains a RNA polymerase promoter, for example, a U6 promoter or the H1 RNA polymerase III promoter, or the siRNA may be a synthetically derived RNA molecule. In certain embodiments the subject invention siRNA polynucleotide may have blunt ends, that is, each nucleotide in one strand of the duplex is perfectly complementary (e.g., by Watson-Crick base-pairing) with a nucleotide of the opposite strand. In certain other embodiments, at least one strand of the subject invention siRNA polynucleotide has at least one, and preferably two nucleotides that "overhang" (i.e., that do not base pair with a complementary base in the opposing strand) at the 3' end of either strand, or preferably both strands, of the siRNA polynucleotide. In a preferred embodiment of the invention, each strand of the siRNA polynucleotide duplex has a two-nucleotide overhang at the 3' end. The two-nucleotide overhang is preferably a thymidine dinucleotide (TT) but may also comprise other bases, for example, a TC dinucleotide or a TG dinucleotide, or any other dinucleotide. The overhang dinucleotide may also be complementary to the two nucleotides at the 5' end of the sequence of the polynucleotide that is targeted for interference. For a discussion of 3' ends of siRNA polynucleotides see, e.g., WO 01/75164.

[0078] Preferred siRNA polynucleotides comprise doublestranded oligomeric nucleotides of about 18-30 nucleotide base pairs, preferably about 18, 19, 20, 21, 22, 23, 24, 25, 26, or 27 base pairs, and in other preferred embodiments about 19, 20, 21, 22 or 23 base pairs, or about 27 base pairs, whereby the use of "about" indicates, as described above, that in certain embodiments and under certain conditions the processive cleavage steps that may give rise to functional siRNA polynucleotides that are capable of interfering with expression of a selected polypeptide may not be absolutely efficient. Hence, siRNA polynucleotides, for instance, of "about" 18, 19, 20, 21, 22, 23, 24, or 25 base pairs may include one or more siRNA polynucleotide molecules that may differ (e.g., by nucleotide insertion or deletion) in length by one, two, three or four base pairs, by way of non-limiting theory as a consequence of variability in processing, in biosynthesis, or in artificial synthesis. The contemplated siRNA polynucleotides of the present invention may also comprise a polynucleotide sequence that exhibits variability by differing (e.g., by nucleotide substitution, including transition or transversion) at one, two, three or four nucleotides from a particular sequence, the differences occurring at any of positions 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, or 19 of a particular siRNA polynucleotide sequence, or at positions 20, 21, 22, 23, 24, 25, 26, or 27 of siRNA polynucleotides depending on the length of the molecule, whether situated in a sense or in an antisense strand of the double-stranded polynucleotide. The nucleotide substitution may be found only in one strand, by way of example in the antisense strand, of a double-stranded polynucleotide, and the complementary nucleotide with which the substitute nucleotide would typically form hydrogen bond base pairing may not necessarily be correspondingly substituted in the sense strand. In preferred embodiments, the siRNA polynucleotides are homogeneous with respect to a specific nucleotide sequence. As described herein, preferred siRNA polynucleotides interfere with expression of a DSP-3, SHP-2, KAP, PRL-3, cdc14 or cdc25 polypeptide. These polynucleotides may also find uses as probes or primers.

[0079] Polynucleotides that are siRNA polynucleotides of the present invention may in certain embodiments be derived from a single-stranded polynucleotide that comprises a single-stranded oligonucleotide fragment (e.g., of about 18-30 nucleotides, which should be understood to include any whole integer of nucleotides including and between 18 and 30) and its reverse complement, typically separated by a spacer sequence. According to certain such embodiments, cleavage of the spacer provides the single-stranded oligonucleotide fragment and its reverse complement, such that they may anneal to form (optionally with additional processing steps that may result in addition or removal of one, two, three or more nucleotides from the 3' end and/or the 5' end of either or both strands) the double-stranded siRNA polynucleotide of the present invention. In

certain embodiments the spacer is of a length that permits the fragment and its reverse complement to anneal and form a double-stranded structure (e.g., like a hairpin polynucleotide) prior to cleavage of the spacer (and, optionally, subsequent processing steps that may result in addition or removal of one, two, three, four, or more nucleotides from the 3' end and/or the 5' end of either or both strands). A spacer sequence may therefore be any polynucleotide sequence as provided herein that is situated between two complementary polynucleotide sequence regions which, when annealed into a double-stranded nucleic acid, comprise a siRNA polynucleotide. Preferably a spacer sequence comprises at least 4 nucleotides, although in certain embodiments the spacer may comprise 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21-25, 26-30, 31-40, 41-50, 51-70, 71-90, 91-110, 111-150, 151-200 or more nucleotides. Examples of siRNA polynucleotides derived from a single nucleotide strand comprising two complementary nucleotide sequences separated by a spacer have been described (e.g., Brummelkamp et al., 2002 Science 296:550; Paddison et al., 2002 Genes Develop. 16:948; Paul et al. Nat. Biotechnol. 20:505-508 (2002); Grabarek et al., BioTechniques 34:734-44 (2003)).

[0080] Polynucleotide variants may contain one or more substitutions, additions, deletions, and/or insertions such that the activity of the siRNA polynucleotide is not substantially diminished, as described above. The effect on the activity of the siRNA polynucleotide may generally be assessed as described herein, or using conventional methods. Variants preferably exhibit at least about 75%, 78%, 80%, 85%, 87%, 88% or 89% identity and more preferably at least about 90%, 92%, 95%, 96%, or 97% identity to a portion of a polynucleotide sequence that encodes a native DSP-3, SHP-2, KAP, PRL-3, cdc14 or cdc25. The percent identity may be readily determined by comparing sequences of the polynucleotides to the corresponding portion of the target polynucleotide, using any method including using computer algorithms well known to those having ordinary skill in the art, such as Align or the BLAST algorithm (Altschul, J. Mol. Biol. 219:555-565, 1991; Henikoff and Henikoff, Proc. Natl. Acad. Sci. USA 89:10915-10919, 1992), which is available at the NCBI website (see [online] Internet:<URL:http:// www/ncbi.nlm.nih.gov/cgi-bin/BLAST). Default parameters may be used.

[0081] Certain siRNA polynucleotide variants are substantially homologous to a portion of a native gene that encodes a desired target polypeptide. Single-stranded nucleic acids derived (e.g., by thermal denaturation) from such polynucleotide variants are capable of hybridizing under moderately stringent conditions to a naturally occurring DNA or RNA sequence encoding a native target polypeptide. In a preferred embodiment of the invention, a siRNA polynucleotide that detectably hybridizes under moderately stringent conditions to a target polypeptide-encoding polynucleotide comprises a nucleotide sequence other than SEQ ID NO:10, which is disclosed in Mailand et al. (2002 Nature Cell Biol. 4:317). A siRNA polynucleotide that detectably hybridizes under moderately stringent conditions may have a nucleotide sequence that includes at least 10 consecutive nucleotides, more preferably 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or 30 consecutive nucleotides that are complementary to a particular target polynucleotide. In certain preferred embodiments such a siRNA sequence (or its complement) will be unique to a single particular target

polypeptide for which interference with expression is desired, and in certain other embodiments the sequence (or its complement) may be shared by two or more related target polypeptides for which interference with polypeptide expression is desired.

[0082] Suitable moderately stringent conditions include, for example, pre-washing in a solution of 5×SSC, 0.5% SDS, 1.0 mM EDTA (pH 8.0); hybridizing at 50° C.-70° C., 5×SSC for 1-16 hours (e.g., overnight); followed by washing once or twice at 22-65° C. for 20-40 minutes with one or more each of 2x, 0.5x and 0.2xSSC containing 0.05-0.1% SDS. For additional stringency, conditions may include a wash in 0.1×SSC and 0.1% SDS at 50-60° C. for 15-40 minutes. As known to those having ordinary skill in the art, variations in stringency of hybridization conditions may be achieved by altering the time, temperature, and/or concentration of the solutions used for pre-hybridization, hybridization, and wash steps. Suitable conditions may also depend in part on the particular nucleotide sequences of the probe used, and of the blotted, proband nucleic acid sample. Accordingly, it will be appreciated that suitably stringent conditions can be readily selected without undue experimentation when a desired selectivity of the probe is identified, based on its ability to hybridize to one or more certain proband sequences while not hybridizing to certain other proband sequences.

[0083] Sequence specific siRNA polynucleotides of the present invention may be designed using one or more of several criteria. For example, to design a siRNA polynucleotide that has 19 consecutive nucleotides identical to a sequence encoding a polypeptide of interest (e.g., PTP1B and other polypeptides described herein), the open reading frame of the polynucleotide sequence may be scanned for 21-base sequences that have one or more of the following characteristics: (1) an A+T/G+C ratio of approximately 1:1 but no greater than 2:1 or 1:2; (2) an AA dinucleotide or a CA dinucleotide at the 5' end; (3) an internal hairpin loop melting temperature less than 55° C.; (4) a homodimer melting temperature of less than 37° C. (melting temperature calculations as described in (3) and (4) can be determined using computer software known to those skilled in the art); (5) a sequence of at least 16 consecutive nucleotides not identified as being present in any other known polynucleotide sequence (such an evaluation can be readily determined using computer programs available to a skilled artisan such as BLAST to search publicly available databases). Alternatively, a siRNA polynculeotide sequence may be designed and chosen using a computer software available commercially from various vendors (e.g., OligoEngine™ (Seattle, Wash.); Dharmacon, Inc. (Lafayette, Colo.); Ambion Inc. (Austin, Tex.); and QIAGEN, Inc. (Valencia, Calif.)). (See also Elbashir et al., Genes & Development 15:188-200 (2000); Elbashir et al., Nature 411:494-98 (2001); and [online] Internet: URLhttp://www.mpibpc.g- wdg.de/abteilungen/100/105/Tusch1 MIV2(3) 2002.pdf.) The siRNA polynucleotides may then be tested for their ability to interfere with the expression of the target polypeptide according to methods known in the art and described herein. The determination of the effectiveness of an siRNA polynucleotide includes not only consideration of its ability to interfere with polypeptide expression but also includes consideration of whether the siRNA polynucleotide manifests undesirably toxic effects, for example, apoptosis of a cell for which cell death is not a desired effect of RNA interference (e.g., interference of PTP1B expression in a cell).

[0084] It should be appreciated that not all siRNAs designed using the above methods will be effective at silencing or interfering with expression of a desired target polypeptide. And further, that the siRNAs will effect silencing to different degrees. Such siRNAs must be tested for their effectiveness, and selections made therefrom based on the ability of a given siRNA to interfere with or modulate (e.g., decrease in a statistically significant manner) the expression of the target. Accordingly, identification of specific siRNA polynucleotide sequences that are capable of interfering with expression of a desired target polypeptide requires production and testing of each siRNA, as demonstrated in greater detail below (see Examples).

[0085] Furthermore, not all siRNAs that interfere with protein expression will have a physiologically important effect. The inventors here have designed, and describe herein, physiologically relevant assays for measuring the influence of modulated target polypeptide expression, for instance, cellular proliferation, induction of apoptosis, and/ or altered levels of protein tyrosine phosphorylation (e.g., insulin receptor phosphorylation), to determine if the levels of interference with target protein expression that were observed using the siRNAs of the invention have clinically relevant significance. Additionally, and according to nonlimiting theory, the invention contemplates altered (e.g., decreased or increased in a statistically significant manner) expression levels of one or more polypeptides of interest, and/or altered (i.e., increased or decreased) phosphorylation levels of one or more phosphoproteins of interest, which altered levels may result from impairment of target protein expression and/or cellular compensatory mechanisms that are induced in response to RNAi-mediated inhibition of a specific target polypeptide expression.

[0086] Persons having ordinary skill in the art will also readily appreciate that as a result of the degeneracy of the genetic code, many nucleotide sequences may encode a polypeptide as described herein. That is, an amino acid may be encoded by one of several different codons and a person skilled in the art can readily determine that while one particular nucleotide sequence may differ from another (which may be determined by alignment methods disclosed herein and known in the art), the sequences may encode polypeptides with identical amino acid sequences. By way of example, the amino acid leucine in a polypeptide may be encoded by one of six different codons (TTA, TTG, CTT, CTC, CTA, and CTG) as can serine (TCT, TCC, TCA, TCG, AGT, and AGC). Other amino acids, such as proline, alanine, and valine, for example, may be encoded by any one of four different codons (CCT, CCC, CCA, CCG for proline; GCT, GCC, GCA, GCG for alanine; and GTT, GTC, GTA, GTG for valine). Some of these polynucleotides bear minimal homology to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically contemplated by the present invention.

[0087] Polynucleotides, including target polynucleotides, may be prepared using any of a variety of techniques, which will be useful for the preparation of specifically desired siRNA polynucleotides and for the identification and selec-

tion of desirable sequences to be used in siRNA polynucleotides. For example, a polynucleotide may be amplified from cDNA prepared from a suitable cell or tissue type. Such polynucleotides may be amplified via polymerase chain reaction (PCR). For this approach, sequence-specific primers may be designed based on the sequences provided herein and may be purchased or synthesized. An amplified portion may be used to isolate a full-length gene, or a desired portion thereof, from a suitable library (e.g., human skeletal muscle cDNA) using well known techniques. Within such techniques, a library (cDNA or genomic) is screened using one or more polynucleotide probes or primers suitable for amplification. Preferably, a library is size-selected to include larger molecules. Random primed libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences. Suitable sequences for a siRNA polynucleotide contemplated by the present invention may also be selected from a library of siRNA polynucleotide sequences.

[0088] For hybridization techniques, a partial sequence may be labeled (e.g., by nick-translation or end-labeling with ³²P) using well known techniques. A bacterial or bacteriophage library may then be screened by hybridizing filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe (see, e.g., Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratories, Cold Spring Harbor, N.Y., 2001). Hybridizing colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. Clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a primer from the partial sequence and a primer from the vector. Restriction maps and partial sequences may be generated to identify one or more overlapping clones. A full-length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

[0089] Alternatively, numerous amplification techniques are known in the art for obtaining a full-length coding sequence from a partial cDNA sequence. Within such techniques, amplification is generally performed via PCR. One such technique is known as "rapid amplification of cDNA ends" or RACE. This technique involves the use of an internal primer and an external primer, which hybridizes to a polyAregion or vector sequence, to identify sequences that are 5' and 3' of a known sequence. Any of a variety of commercially available kits may be used to perform the amplification step. Primers may be designed using, for example, software well known in the art. Primers (or oligonucleotides for other uses contemplated herein, including, for example, probes and antisense oligonucleotides) are preferably 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31 or 32 nucleotides in length, have a GC content of at least 40% and anneal to the target sequence at temperatures of about 54° C. to 72° C. The amplified region may be sequenced as described above, and overlapping sequences assembled into a contiguous sequence. Certain oligonucleotides contemplated by the present invention may, for some preferred embodiments, have lengths of 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33-35, 35-40, 41-45, 46-50, 56-60, 61-70, 71-80, 81-90 or more nucleotides.

[0090] A number of specific siRNA polynucleotide sequences useful for interfering with target polypeptide expression, and are presented in the Examples, the Drawings, and the Sequence Listing. SiRNA polynucleotides may generally be prepared by any method known in the art, including, for example, solid phase chemical synthesis. Modifications in a polynucleotide sequence may also be introduced using standard mutagenesis techniques, such as oligonucleotide-directed site-specific mutagenesis. Further, siRNAs may be chemically modified or conjugated to improve theur serum stability and/or delivery properties. Included as an aspect of the invention are the siRNAs described herein wherein the ribose has been removed therefrom. Alternatively, siRNA polynucleotide molecules may be generated by in vitro or in vivo transcription of suitable DNA sequences (e.g., polynucleotide sequences encoding a target polypeptide, or a desired portion thereof), provided that the DNA is incorporated into a vector with a suitable RNA polymerase promoter (such as T7, U6, H1, or SP6). In addition, a siRNA polynucleotide may be administered to a patient, as may be a DNA sequence (e.g., a recombinant nucleic acid construct as provided herein) that supports transcription (and optionally appropriate processing steps) such that a desired siRNA is generated in vivo.

[0091] Accordingly, a siRNA polynucleotide that is complementary to at least a portion of a target polypeptideencoding sequence may be used to modulate gene expression, or as a probe or primer. Identification of siRNA polynucleotide sequences and DNA encoding genes for their targeted delivery involves techniques described herein. Identification of such siRNA polynucleotide sequences and DNA encoding genes for their targeted delivery involves techniques that are also described herein. As discussed above, siRNA polynucleotides exhibit desirable stability characteristics and may, but need not, be further designed to resist degradation by endogenous nucleolytic enzymes by using such linkages as phosphorothioate, methylphosphonate, sulfone, sulfate, ketyl, phosphorodithioate, phosphoramidate, phosphate esters, and other such linkages (see, e.g., Agrwal et al., Tetrahedron Lett. 28:3539-3542 (1987); Miller et al., J. Am. Chem. Soc. 93:6657-6665 (1971); Stec et al., Tetrahedron Lett. 26:2191-2194 (1985); Moody et al., Nucleic Acids Res. 12:4769-4782 (1989); Uznanski et al., Nucleic Acids Res. (1989); Letsinger et al., Tetrahedron 40:137-143 (1984); Eckstein, Annu. Rev. Biochem. 54:367402 (1985); Eckstein, Trends Biol. Sci. 14:97-100 (1989); Stein, In: Oligodeoxynucleotides. Antisense Inhibitors of Gene Expression, Cohen, ed., Macmillan Press, London, pp. 97-117 (1989); Jager et al., Biochemistry 27:7237-7246 (1988)).

[0092] Any polynucleotide of the invention may be further modified to increase stability in vivo. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than phosphodiester linkages in the backbone; and/or the inclusion of nontraditional bases such as inosine, queosine, and wybutosine and the like, as well as acetyl- methyl-, thio- and other modified forms of adenine, cytidine, guanine, thymine, and uridine.

[0093] Nucleotide sequences as described herein may be joined to a variety of other nucleotide sequences using established recombinant DNA techniques. For example, a polynucleotide may be cloned into any of a variety of

cloning vectors, including plasmids, phagemids, lambda phage derivatives, and cosmids. Vectors of particular interest include expression vectors, replication vectors, probe generation vectors, and sequencing vectors. In general, a suitable vector contains an origin of replication functional in at least one organism, convenient restriction endonuclease sites, and one or more selectable markers. (See, e.g., WO 01/96584; WO 01/29058; U.S. Pat. No. 6,326,193; U.S. 2002/0007051). Other elements will depend upon the desired use, and will be apparent to those having ordinary skill in the art. For example, the invention contemplates the use of siRNA polynucleotide sequences in the preparation of recombinant nucleic acid constructs including vectors for interfering with the expression of a desired target polypeptide such as a PTP polypeptide, a MAP kinase kinase polypeptide, or a chemotherapeutic target polypeptide in vivo; the invention also contemplates the generation of siRNA transgenic or "knock-out" animals and cells (e.g., cells, cell clones, lines or lineages, or organisms in which expression of one or more desired polypeptides (e.g., a target polypeptide) is fully or partially compromised). An siRNA polynucleotide that is capable of interfering with expression of a desired polypeptide (e.g., a target polypeptide) as provided herein thus includes any siRNA polynucleotide that, when contacted with a subject or biological source as provided herein under conditions and for a time sufficient for target polypeptide expression to take place in the absence of the siRNA polynucleotide, results in a statistically significant decrease (alternatively referred to as "knockdown" of expression) in the level of target polypeptide expression that can be detected. Preferably the decrease is greater than 10%, more preferably greater than 20%, more preferably greater than 30%, more preferably greater than 40%, 50%, 60%, 70%, 75%, 80%, 85%, 90%, 95% or 98% relative to the expression level of the polypeptide detected in the absence of the siRNA, using conventional methods for determining polypeptide expression as known to the art and provided herein. Preferably, the presence of the siRNA polynucleotide in a cell does not result in or cause any undesired toxic effects, for example, apoptosis or death of a cell in which apoptosis is not a desired effect of RNA interference.

[0094] Within certain embodiments, siRNA polynucleotides may be formulated so as to permit entry into a cell of a mammal, and expression therein. Such formulations are particularly useful for therapeutic purposes, as described below. Those having ordinary skill in the art will appreciate that there are many ways to achieve expression of a polynucleotide in a target cell, and any suitable method may be employed. For example, a polynucleotide may be incorporated into a viral vector using well known techniques (see also, e.g., U.S. 2003/0068821). A viral vector may additionally transfer or incorporate a gene for a selectable marker (to aid in the identification or selection of transduced cells) and/or a targeting moiety, such as a gene that encodes a ligand for a receptor on a specific target cell, to render the vector target specific. Targeting may also be accomplished using an antibody, by methods known to those having ordinary skill in the art.

[0095] Other formulations for therapeutic purposes include colloidal dispersion systems, such as macromolecule complexes, nanocapsules, microspheres, beads, and lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, and liposomes. A preferred colloidal system for use as a delivery vehicle in vitro and in vivo is a liposome

(i.e., an artificial membrane vesicle). The preparation and use of such systems is well known in the art.

[0096] Within other embodiments, one or more promoters may be identified, isolated and/or incorporated into recombinant nucleic acid constructs of the present invention, using standard techniques. The present invention provides nucleic acid molecules comprising such a promoter sequence or one or more cis- or trans-acting regulatory elements thereof. Such regulatory elements may enhance or suppress expression of a siRNA. A 5' flanking region may be generated using standard techniques, based on the genomic sequence provided herein. If necessary, additional 5' sequences may be generated using PCR-based or other standard methods. The 5' region may be subcloned and sequenced using standard methods. Primer extension and/or RNase protection analyses may be used to verify the transcriptional start site deduced from the cDNA.

[0097] To define the boundary of the promoter region, putative promoter inserts of varying sizes may be subcloned into a heterologous expression system containing a suitable reporter gene without a promoter or enhancer. Suitable reporter genes may include genes encoding luciferase, betagalactosidase, chloramphenicol acetyl transferase, secreted alkaline phosphatase, or the Green Fluorescent Protein gene (see, e.g., Ui-Tei et al., FEBS Lett. 479:79-82 (2000). Suitable expression systems are well known and may be prepared using well known techniques or obtained commercially. Internal deletion constructs may be generated using unique internal restriction sites or by partial digestion of non-unique restriction sites. Constructs may then be transfected into cells that display high levels of siRNA polynucleotide and/or polypeptide expression. In general, the construct with the minimal 5' flanking region showing the highest level of expression of reporter gene is identified as the promoter. Such promoter regions may be linked to a reporter gene and used to evaluate agents for the ability to modulate promoter-driven transcription.

[0098] Once a functional promoter is identified, cis- and trans-acting elements may be located. Cis-acting sequences may generally be identified based on homology to previously characterized transcriptional motifs. Point mutations may then be generated within the identified sequences to evaluate the regulatory role of such sequences. Such mutations may be generated using site-specific mutagenesis techniques or a PCR-based strategy. The altered promoter is then cloned into a reporter gene expression vector, as described above, and the effect of the mutation on reporter gene expression is evaluated.

[0099] In general, polypeptides and polynucleotides as described herein are isolated. An "isolated" polypeptide or polynucleotide is one that is removed from its original environment. For example, a naturally occurring protein is isolated if it is separated from some or all of the coexisting materials in the natural system. Preferably, such polypeptides are at least about 90% pure, more preferably at least about 95% pure and most preferably at least about 99% pure. A polynucleotide is considered to be isolated if, for example, it is cloned into a vector that is not a part of the natural environment. A "gene" includes the segment of DNA involved in producing a polypeptide chain; it further includes regions preceding and following the coding region "leader and trailer," for example promoter and/or enhancer

and/or other regulatory sequences and the like, as well as intervening sequences (introns) between individual coding segments (exons).

[0100] As noted above, according to certain embodiments of the invention compositions and methods are provided that relate to altering or altered expression of a PTP as described herein (including DSPs) or of other target polypeptides as disclosed herein, and/or to a PTP associated disorder. A PTP associated disorder includes any disease, disorder, condition, syndrome, pathologic or physiologic state, or the like, wherein at least one undesirable deviation or departure from a physiological norm causes, correlates with, is accompanied by or results from an inappropriate alteration (i.e., a statistically significant change) to the structure, activity, function, expression level, physicochemical or hydrodynamic property, or stability of a PTP or of a molecular component of a biological signal transduction pathway that comprises a PTP, for instance, a MAP kinase such as JNK (e.g., Shen et al., 2001 Proc. Nat. Acad. Sci. USA 98:13613; see also U.S. Pat. No. 6,342,595), TYK2 or Jak2 (e.g., Myers et al., 2001. J. Biol. Chem. 276:47771), or a MAP kinase kinase MKK4 or MKK7 (e.g., Shen et al., Proc. Natl. Acad. Sci. USA 98:13613-18 (2001) and references cited therein), a receptor such as IR (Salmeen et al., 2000), or leptin receptor (e.g., Kalman et al. 2000 and references cited therein) or other such pathways comprising PTPs as known to the art. In preferred embodiments the molecular component may be a protein, peptide or polypeptide, and in certain other preferred embodiments the alteration may be an altered level of PTP expression. In certain other preferred embodiments the alteration may be manifest as an a typical or unusual phosphorylation state of a protein under particular conditions, for example, hypophosphorylation or hyperphosphorylation of a phosphoprotein, wherein those familiar with the art will appreciate that phosphorylated proteins typically comprise one or more phosphotyrosine, phosphoserine, or phosphothreonine residues.

[0101] PTP associated disorders therefore include, for example, diabetes mellitus, obesity, impaired glucose tolerance and other metabolic disorders wherein alteration of a biological signaling pathway component is associated with the disorder The effect of siRNA interference with expression of a component in the signal transduction pathway induced by insulin, for example, may be evaluated by determining the level of tyrosine phosphorylation of insulin receptor beta (IR-β) and/or of the downstream signaling molecule PKB/Akt and/or of any other downstream polypeptide that may be a component of a particular signal transduction pathway as provided herein. The invention is not intended, however, to be so limited and contemplates other disorders, such as JNK-associated disorders (e.g., cancer, cardiac hypertrophy, ischemia, diabetes, hyperglycemia-induced apoptosis, inflammation, neurodegenerative disorders), and other disorders associated with different signal transduction pathways, for instance, cancer, autoimmunity, cellular proliferative disorders, neurodegenerative disorders, and infectious diseases (see, e.g., Fukada et al., 2001 J. Biol. Chem. 276:25512; Tonks et al., 2001 Curr. Opin. Cell Biol. 13:182; Salmeen et al., 2000 Mol. Cell 6:1401; Hu et al., J. Neurochem. 85:432-42 (2003); and references cited therein).

[0102] Cancer is also associated with other dual specificity phosphatases, such as DSP-3, PRL-3 (see, e.g., Saha et al.,

Science 294:1343-46 (2001), PTP ϵ (Elson, Oncogene 18:7535-42 (1999)), and the cell cycle dual specificity phosphatases cdc25 (see, e.g., Donzelli et al., EMBO 21:4875-84 (2002), cdc14 (Wong et al., Genomics 59:248-51 (1999)), and KAP (see, e.g., Lee et al., Mol. Cell Biol. 20:1723-32 (2000); Yeh et al., Cancer Res. 60:4697-700 (2000); see also, e.g., Donato et al., J. Clin. Invest. 109:51-58 (2002)). Another dual specificity phosphatase believed to be involved in the cell cycle, cdc14, is reported to interact with the tumor suppressor protein p53 (Li et al., J. Biol. Chem. 275:2410014 (2000); see also Agami et al., Cell 102:55-66 (2000)). In normal cells, cdc14 is reported to be a part of the mitotic exit network, which involves intricate regulatory pathways that coordinate chromosome segregation and mitotic exit with physical separation of two nascent cells, and in cytokineses (see, e.g., Gruneberg et al., J. Cell Biol. 158:901-14 (2002); Trautman et al., Curr. Biol. 12:R733-R735 (2002); Visintin et al., Mol. Cell 2:709-18 (1998); see also Mailand et al., supra). Persons skilled in the art will be familiar with an array of criteria according to which it may be recognized what are, for instance, biological, physiological, pathological and/or clinical signs and/or symptoms of PTP associated and other disorders as provided herein (see, e.g., Irie-Sasaki et al., Curr. Top. Med. Chem. 3:783-96 (2003) (discussing role of CD45 in signal transduction pathways); Oh et al., Mol. Cell Biol. 19:3205-15 (1999) (describing regulation of early events in integrin signaling by SHP-2); Musante et al., Eur. J. Hum. Genet. 11:201-206 (2003), Tartaglia et al., Nat. Genet. 29:465-68 (2001), and Ion et al., Hum. Genet. 111:421-27 (2002) (discussing correlation between mutations in the PTPN11 gene that encodes SHP-2 and Noonan Syndrome)); Tanuma et al., Blood 98:3030-34 (2001) (reporting that PTPe inhibits IL-6 and IL-10 induced JAK-STAT signaling)).

[0103] Also contemplated by the invention are disorders associated with the NF-kappaB signaling pathway, for example, in cancer cells in which NF-kappaB is overexpressed or constitutively activated (see, e.g., Bayon et al., Mol. Cell Biol. 23:1061-74 (2003); Arsura et al., Oncogene 22:412-25 (2003)). Other disorders associated with the NF-kappaB signaling pathway include those associated with other components of the pathway, for example, inflammation associated with IkappaB kinase gamma (IKKgamma), which is an upstream regulator of NF-kappaB that is required for NF-kappaB activation by various stimuli (see, e.g., Makris et al., Mol. Cell Biol. 22:6573-81 (2002); Li et al., J. Biol. Chem. 277:45129-40 (2002); Sadikot et al., J. Immunol. 170:1091-98 (2003)).

[0104] As noted above, regulated tyrosine phosphorylation contributes to specific pathways for biological signal transduction, including those associated with cell division, cell survival, apoptosis, proliferation and differentiation, and "biological signal transduction pathways," or "inducible signaling pathways" in the context of the present invention include transient or stable associations or interactions among molecular components involved in the control of these and similar processes in cells. Depending on the particular pathway of interest, an appropriate parameter for determining induction of such pathway may be selected. For example, for signaling pathways associated with cell proliferation, a variety of well known methodologies are available for quantifying proliferation, including, for example, incorporation of tritiated thymidine into cellular DNA, monitoring of detectable (e.g., fluorimetric or calorimetric)

indicators of cellular respiratory activity (for example, conversion of the tetrazolium salts (yellow) 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) or 3-(4, 5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4sulphophenyl)-2H-tetrazolium (MTS) to formazan dyes (purple) in metabolically active cells), or cell counting, or the like. Similarly, in the cell biology arts, multiple techniques are known for assessing cell survival (e.g., vital dyes, metabolic indicators, etc.) and for determining apoptosis (for example, annexin V binding, DNA fragmentation assays, caspase activation, marker analysis, e.g., poly(ADP-ribose) polymerase (PARP), etc.). Other signaling pathways will be associated with particular cellular phenotypes, for example specific induction of gene expression (e.g., detectable as transcription or translation products, or by bioassays of such products, or as nuclear localization of cytoplasmic factors), altered (e.g., statistically significant increases or decreases) levels of intracellular mediators (e.g., activated kinases or phosphatases, altered levels of cyclic nucleotides or of physiologically active ionic species, etc.), altered cell cycle profiles, or altered cellular morphology, and the like, such that cellular responsiveness to a particular stimulus as provided herein can be readily identified to determine whether a particular cell comprises an inducible signaling pathway.

[0105] In addition, according to certain embodiments of the invention compositions and methods are provided that relate to altering or altered expression of chemotherapeutic target polypeptides. Sequence specific siRNA polynucleotides may be used as a conjunctive therapy with chemotherapeutic drugs or may provide an alternative therapy in circumstances when a cancer becomes refractory to chemotherapeutic treatment regimens. Resistance to chemotherapeutic drugs may develop when a chemotherapeutic target polypeptide is overexpressed or when its expression becomes constitutive. Overexpression or amplified expression of such a target polypeptide could be reduced by introducing a specific siRNA polynucleotide into the cell. In particular, chemotherapeutic target polypeptides that may become resistant to drug therapies include, for example, components of the thymidylate biosynthesis pathway, thymidylate synthetase and DHFR, which become refractory to anti-neoplastic drugs such as 5-FU and methotrexate, respectively, and contribute to a drug resistance phenotype. Also contemplated by the invention are sequence specific siRNA polynucleotides that interfere with expression of DNA-processing enzymes such as topoisomerase I and that would have anti-cancer or anti-bacterial effects. The effect of siRNA interference on expression of such chemotherapeutic target polypeptides may alter cell division, cell survival, apoptosis, proliferation, and differentiation, which may be assessed by any of the techniques and methods described herein.

[0106] PTPs

[0107] As used herein, a phosphatase is a member of the PTP family if it contains the signature motif CX₅R (SEQ ID NO: ______). Dual specificity PTPs, i.e., PTPs that dephosphorylate both phosphorylated tyrosine and phosphorylated serine or threonine, are also suitable for use in the invention. PTPs for use in the present invention include PTP1B (e.g., GenBank Accession Nos. M31724 (SEQ ID NOS: _______); NM_002827 (SEQ ID NOS: _______); M31724 (SEQ ID NOS: _______); M31724 (SEQ ID NOS: _______); M33689 (SEQ ID NOS: ________)

M33962 (SEQ ID)). In certain preferred embodiments, TC-PTP (e.g., GenBank Accession Nos. M25393 (SEQ ID NOS: _); M81478 (SEQ ID NO:); M80737 (SEQ ID NO:); M81477 (SEQ ID NOS:); X58<u>828</u> (SEQ ID NOS: NM_002828 (SEQ ID NOS: TC45 (e.g., NM_080422 (SEQ ID NOS:)) may be used. In certain other embodiments PTPs and DSPs for use in the present invention include DSP-3 (WO00/60092); SHP2, (e.g., GenBank Accession Nos. D13540 (SEQ ID NOS:); L03535 (SEQ ID _); L07527 (SEQ ID NOS:); L0<u>880</u>7); X70766 (SEQ ID NOS: (SEQ ID NO:); S78088 (SEQ ID NOS: _); S39383 (SEQ ID NO: _); D84372 (SEQ ID _); U09307 (SEQ ID NOS: 15-16)); cdc14 (which includes cdc14a (e.g., GenBank Accession _); AF064102 Nos. AF122013 (SEQ ID NOS:); AF064103 (SEQ ID NOS: (SEQ ID NOS:); Li et al., 1997 J. Biol. Chem. 272:29403; U.S. Pat. No. 6,331,614) and cdc14b (e.g., GenBank Accession Nos. AF064104 (SEQ ID NOS: AF064105 (SEQ ID NOS: ((e.g., GenBank Accession Nos. NM_001789 (SEQ ID _), AF527417 (SEQ ID NOS:]), NM_133571 (SEQ ID NOS: CDC25B (e.g., GenBank Accession Nos. NM_133572 _), NM_023117 (SEQ ID (SEQ ID NOS: _), NM_021872 (SEQ ID NOS: NOS:); NM_021872; M81934) (SEQ ID NOS:); and CDC25C (e.g., GenBank Accession Nos. NM_001790 (SEQ ID NOS: NM_022809 (SEQ ID NOS:)); CD45 (Charbonneau et al., Proc. Natl. Acad. Sci. USA 85:7182-86 (1988); Genbank Accession Nos. NM_080922 (SEQ ID _), NM_080921 (SEQ ID NOS: NOS:), NM_002838 (SEQ ID NOS: NM_080923) (SEQ and ID); GenBank Acc. No. XM_16748; SEQ ID NO:32 encoded by SEQ ID NO:31; KAP (Genbank Accession No. L27711 (SEQ ID NOS: et al., Proc. Natl. Acad. Sci. USA 91:1731-35 (1994)); PTP€ (e.g., Genbank Accession Nos. NM_006504 (SEQ ID NOS:) and NM_130435 (SEQ ID NOS:)); and PRL-3 (e.g., Zhao et al., Genomics 35:172-81 (1996); Genbank Accession Nos. (NM 003479 (SEQ ID _), NM_080392 (SEQ ID NOS: NOS: <u>), NM</u>_080391 (SEQ ID NOS:), NM_032611 (SEQ ID NOS: _ and NM_007079 (SEQ ID NOS: tain preferred embodiments PTPs and DSPs include, but are not limited to, U.S. application Ser. No. 10/151,320 (DSP18); WO 01/05983 (DSP-11); U.S. application Ser. No. 09/775,925 (DSP-12 and DSP-13); U.S. application Ser. No. 09/847,519 and WO 01/46394 (DSP-14); The invention also contemplates using mutated forms of the PTPs and DSPs, which may include PTPs and DSPs that contain single nucleotide polymorphisms (SNPs), or may include allelic forms.

[0108] Specific substitutions of individual amino acids through introduction of site-directed mutations are well-known and may be made according to methodologies with which those having ordinary skill in the art will be familiar.

The effects on catalytic activity of the resulting mutant PTP may be determined empirically by testing the resulting modified protein for the preservation of the Km and reduction of Kcat to less than 1 per minute as provided herein and as previously disclosed (e.g., WO98/04712; Flint et al., 1997 *Proc. Nat. Acad. Sci. USA* 94:1680). In addition, the effect on phosphorylatation of one or more tyrosine residues of the resulting mutant PTP molecule can also be determined empirically merely by testing such a mutant for the presence of phosphotyrosine, as also provided herein, for example, following exposure of the mutant to conditions in vitro or in vivo where it may act as a phosphate acceptor for a protein tyrosine kinase.

[0109] In particular, portions of two PTP polypeptide sequences are regarded as "corresponding" amino acid sequences, regions, fragments or the like, based on a convention of numbering one PTP sequence according to amino acid position number, and then aligning the sequence to be compared in a manner that maximizes the number of amino acids that match or that are conserved residues, for example, that remain polar (e.g., D, E, K, R, H, S, T, N, Q), hydrophobic (e.g., A, P, V, L, I, M, F, W, Y) or neutral (e.g., C, G) residues at each position. Similarly, a DNA sequence encoding a candidate PTP that is to be mutated as provided herein, or a portion, region, fragment or the like, may correspond to a known wildtype PTP-encoding DNA sequence according to a convention for numbering nucleic acid sequence positions in the known wildtype PTP DNA sequence, whereby the candidate PTP DNA sequence is aligned with the known PTP DNA such that at least 70%, preferably at least 80% and more preferably at least 90% of the nucleotides in a given sequence of at least 20 consecutive nucleotides of a sequence are identical. In certain preferred embodiments, a candidate PTP DNA sequence is greater than 95% identical to a corresponding known PTP DNA sequence. In certain particularly preferred embodiments, a portion, region or fragment of a candidate PTP DNA sequence is identical to a corresponding known PTP DNA sequence. As is well known in the art, an individual whose DNA contains no irregularities (e.g., a common or prevalent form) in a particular gene responsible for a given trait may be said to possess a wildtype genetic complement (genotype) for that gene, while the presence of irregularities known as mutations in the DNA for the gene, for example, substitutions, insertions or deletions of one or more nucleotides, indicates a mutated or mutant genotype. The invention need not be so limited, however, and contemplates other embodiments wherein two or more non-PTP polypeptides of interest (e.g., as siRNA targets), such as MAP kinase kinases or chemotherapeutic target polypeptides, are structurally related and have portions of polypeptide sequences that may be regarded as "corresponding" amino acid sequences, regions, fragments or the like, according to the alignment and identity criteria discussed above.

[0110] Modification of DNA may be performed by a variety of methods, including site-specific or site-directed mutagenesis of DNA encoding the polypeptide of interest (e.g., a siRNA target polypeptide) and the use of DNA amplification methods using primers to introduce and amplify alterations in the DNA template, such as PCR splicing by overlap extension (SOE). Site-directed mutagenesis is typically effected using a phage vector that has single-and double-stranded forms, such as M13 phage vectors, which are well-known and commercially available. Other

suitable vectors that contain a single-stranded phage origin of replication may be used (see, e.g., Veira et al., Meth. Enzymol. 15:3, 1987). In general, site-directed mutagenesis is performed by preparing a single-stranded vector that encodes the protein of interest (e.g., a member of the PTP family, a MAP kinase kinase, or a chemotherapeutic target polypeptide). An oligonucleotide primer that contains the desired mutation within a region of homology to the DNA in the single-stranded vector is annealed to the vector followed by addition of a DNA polymerase, such as E. coli DNA polymerase I (Klenow fragment), which uses the double stranded region as a primer to produce a heteroduplex in which one strand encodes the altered sequence and the other the original sequence. Additional disclosure relating to sitedirected mutagenesis may be found, for example, in Kunkel et al. (Methods in Enzymol. 154:367, 1987) and in U.S. Pat. Nos. 4,518,584 and 4,737,462. The heteroduplex is introduced into appropriate bacterial cells, and clones that include the desired mutation are selected. The resulting altered DNA molecules may be expressed recombinantly in appropriate host cells to produce the modified protein.

[0111] SiRNAs of the invention may be fused to other nucleotide molecules, or to polypeptides, in order to direct their delivery or to accomplish other functions. Thus, for example, fusion proteins comprising a siRNA oligonucleotide that is capable of specifically interfering with expression of a target polypeptide may comprise affinity tag polypeptide sequences, which refers to polypeptides or peptides that facilitate detection and isolation of the such polypeptide via a specific affinity interaction with a ligand. The ligand may be any molecule, receptor, counterreceptor, antibody or the like with which the affinity tag may interact through a specific binding interaction as provided herein. Such peptides include, for example, poly-His or "FLAG®" or the like, e.g., the antigenic identification peptides described in U.S. Pat. No. 5,011,912 and in Hopp et al., (1988 Bio/Technology 6:1204), or the XPRESS™ epitope tag (Invitrogen, Carlsbad, Calif.). The affinity sequence may be a hexa-histidine tag as supplied, for example, by a pBAD/His (Invitrogen) or a pQE-9 vector to provide for purification of the mature polypeptide fused to the marker in the case of a bacterial host, or, for example, the affinity sequence may be a hemagglutinin (HA) tag when a mammalian host, e.g., COS-7 cells, is used. The HA tag corresponds to an antibody defined epitope derived from the influenza hemagglutinin protein (Wilson et al., 1984 Cell 37:767).

[0112] The present invention also relates to vectors and to constructs that include or encode siRNA polynucleotides of the present invention, and in particular to "recombinant nucleic acid constructs" that include any nucleic acids that may be transcribed to yield target polynucleotide-specific siRNA polynucleotides (i.e., siRNA specific for a polynucleotide that encodes a target polypeptide, such as a mRNA) according to the invention as provided above; to host cells which are genetically engineered with vectors and/or constructs of the invention and to the production of siRNA polynucleotides, polypeptides, and/or fusion proteins of the invention, or fragments or variants thereof, by recombinant techniques. SiRNA sequences disclosed herein as RNA polynucleotides may be engineered to produce corresponding DNA sequences using well established methodologies such as those described herein. Thus, for example, a DNA polynucleotide may be generated from any siRNA sequence described herein (including in the Sequence Listing), such that the present siRNA sequences will be recognized as also providing corresponding DNA polynucleotides (and their complements). These DNA polynucleotides are therefore encompassed within the contemplated invention, for example, to be incorporated into the subject invention recombinant nucleic acid constructs from which siRNA may be transcribed.

[0113] According to the present invention, a vector may comprise a recombinant nucleic acid construct containing one or more promoters for transcription of an RNA molecule, for example, the human U6 snRNA promoter (see, e.g., Miyagishi et al, Nat. Biotechnol. 20:497-500 (2002); Lee et al., Nat. Biotechnol. 20:500-505 (2002); Paul et al., Nat. Biotechnol. 20:505-508 (2002); Grabarek et al., Bio-Techniques 34:73544 (2003); see also Sui et al., Proc. Natl. Acad. Sci. USA 99:5515-20 (2002)). Each strand of a siRNA polynucleotide may be transcribed separately each under the direction of a separate promoter and then may hybridize within the cell to form the siRNA polynucleotide duplex. Each strand may also be transcribed from separate vectors (see Lee et al., supra). Alternatively, the sense and antisense sequences specific for a PTP1B sequence may be transcribed under the control of a single promoter such that the siRNA polynucleotide forms a hairpin molecule (Paul et al., supra). In such an instance, the complementary strands of the siRNA specific sequences are separated by a spacer that comprises at least four nucleotides, but may comprise at least 5, 6, 7, 8, 9, 10, 11, 12, 14, 16, 94 18 nucleotides or more nucleotides as described herein. In addition, siRNAs transcribed under the control of a U6 promoter that form a hairpin may have a stretch of about four uridines at the 3' end that act as the transcription termination signal (Miyagishi et al., supra; Paul et al., supra). By way of illustration, if the target sequence is 19 nucleotides, the siRNA hairpin polynucleotide (beginning at the 5' end) has a 19-nucleotide sense sequence followed by a spacer (which as two uridine nucleotides adjacent to the 3' end of the 19-nucleotide sense sequence), and the spacer is linked to a 19 nucleotide antisense sequence followed by a 4-uridine terminator sequence, which results in an overhang. SiRNA polynucleotides with such overhangs effectively interfere with expression of the target polypeptide (see id.). A recombinant construct may also be prepared using another RNA polymerase III promoter, the H1 RNA promoter, that may be operatively linked to siRNA polynucleotide specific sequences, which may be used for transcription of hairpin structures comprising the siRNA specific sequences or separate transcription of each strand of a siRNA duplex polynucleotide (see, e.g., Brummelkamp et al., Science 296:550-53 (2002); Paddison et al., supra). DNA vectors useful for insertion of sequences for transcription of an siRNA polynucleotide include pSUPER vector (see, e.g., Brummelkamp et al., supra); pAV vectors derived from pCWRSVN (see, e.g., Paul et al., supra); and pIND (see, e.g., Lee et al., supra), or the like.

[0114] PTP polypeptides and other target polypeptides of interest can be expressed in mammalian cells, yeast, bacteria, or other cells under the control of appropriate promoters, providing ready systems for evaluation of siRNA polynucleotides that are capable of interfering with polypeptide expression as provided herein. Appropriate cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described, for example, by Sambrook, et al.,

Molecular Cloning: A Laboratory Manual, Third Edition, Cold Spring Harbor, N.Y., (2001).

[0115] Generally, recombinant expression vectors for use in the preparation of recombinant nucleic acid constructs or vectors of the invention will include origins of replication and selectable markers permitting transformation of the host cell, e.g., the ampicillin resistance gene of E. coli and S. cerevisiae TRP1 gene, and a promoter derived from a highly-expressed gene to direct transcription of a downstream structural sequence (e.g., a siRNA polynucleotide sequence). Such promoters can be derived from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), α-factor, acid phosphatase, or heat shock proteins, among others. For PTP polypeptide expression (including PTP fusion proteins and substrate trapping mutant PTPs), and for other expression of other polypeptides of interest, the heterologous structural sequence is assembled in appropriate phase with translation initiation and termination sequences. Optionally, the heterologous sequence can encode a fusion protein including an N-terminal identification peptide imparting desired characteristics, e.g., stabilization or simplified purification of expressed recombinant product.

[0116] Useful expression constructs for bacterial use are constructed by inserting into an expression vector a structural DNA sequence encoding a desired siRNA polynucleotide, together with suitable transcription initiation and termination signals in operable linkage, for example, with a functional promoter. The construct may comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector construct and, if desirable, to provide amplification within the host. Suitable prokaryotic hosts for transformation include *E. coli, Bacillus subtilis, Salmonella typhimurium* and various species within the genera Pseudomonas, Streptomyces, and Staphylococcus, although others may also be employed as a matter of choice. Any other plasmid or vector may be used as long as they are replicable and viable in the host.

[0117] As a representative but nonlimiting example, useful expression vectors for bacterial use can comprise a selectable marker and bacterial origin of replication derived from commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017). Such commercial vectors include, for example, pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden) and GEM1 (Promega Biotec, Madison, Wis., USA). These pBR322 "backbone" sections are combined with an appropriate promoter and the structural sequence to be expressed.

[0118] Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter, if it is a regulated promoter as provided herein, is induced by appropriate means (e.g., temperature shift or chemical induction) and cells are cultured for an additional period. Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification. Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freezethaw cycling, sonication, mechanical disruption, or use of cell lysing agents; such methods are well know to those skilled in the art.

[0119] Thus, for example, the nucleic acids of the invention as described herein (e.g., DNA sequences from which

siRNA may be transcribed) herein may be included in any one of a variety of expression vector constructs as a recombinant nucleic acid construct for expressing a target polynucleotide-specific siRNA polynucleotide. Such vectors and constructs include chromosomal, nonchromosomal and synthetic DNA sequences, e.g., derivatives of SV40; bacterial plasmids; phage DNA; baculovirus; yeast plasmids; vectors derived from combinations of plasmids and phage DNA, viral DNA, such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. However, any other vector may be used for preparation of a recombinant nucleic acid construct as long as it is replicable and viable in the host.

[0120] The appropriate DNA sequence(s) may be inserted into the vector by a variety of procedures. In general, the DNA sequence is inserted into an appropriate restriction endonuclease site(s) by procedures known in the art. Standard techniques for cloning, DNA isolation, amplification and purification, for enzymatic reactions involving DNA ligase, DNA polymerase, restriction endonucleases and the like, and various separation techniques are those known and commonly employed by those skilled in the art. A number of standard techniques are described, for example, in Ausubel et al. (1993 Current Protocols in Molecular Biology, Greene Publ. Assoc. Inc. & John Wiley & Sons, Inc., Boston, Mass.); Sambrook et al. (2001 Molecular Cloning, Third Ed., Cold Spring Harbor Laboratory, Plainview, N.Y.); Maniatis et al. (1982 Molecular Cloning, Cold Spring Harbor Laboratory, Plainview, N.Y.); and elsewhere.

[0121] The DNA sequence in the expression vector is operatively linked to at least one appropriate expression control sequences (e.g., a promoter or a regulated promoter) to direct mRNA synthesis. Representative examples of such expression control sequences include LTR or SV40 promoter, the E. coli lac or trp, the phage lambda PL promoter and other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses. Promoter regions can be selected from any desired gene using CAT (chloramphenicol transferase) vectors or other vectors with selectable markers. Two appropriate vectors are pKK232-8 and pCM7. Particular named bacterial promoters include lac, lacZ, T3, T7, gpt, lambda P_R, P_L and trp. Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus, and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art, and preparation of certain particularly preferred recombinant expression constructs comprising at least one promoter or regulated promoter operably linked to a nucleic acid encoding a polypeptide (e.g., PTP, MAP kinase kinase, or chemotherapeutic target polypeptide) is described herein.

[0122] As noted above, in certain embodiments the vector may be a viral vector such as a retroviral vector. For example, retroviruses from which the retroviral plasmid vectors may be derived include, but are not limited to, Moloney Murine Leukemia Virus, spleen necrosis virus, retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma virus, avian leukosis virus, gibbon ape leukemia virus, human immunodeficiency virus, adenovirus, Myeloproliferative Sarcoma Virus, and mammary tumor virus.

[0123] The viral vector includes one or more promoters. Suitable promoters which may be employed include, but are not limited to, the retroviral LTR; the SV40 promoter; and

the human cytomegalovirus (CMV) promoter described in Miller, et al., *Biotechniques* 7:980-990 (1989), or any other promoter (e.g., cellular promoters such as eukaryotic cellular promoters including, but not limited to, the histone, pol III, and β -actin promoters). Other viral promoters which may be employed include, but are not limited to, adenovirus promoters, thymidine kinase (TK) promoters, and B19 parvovirus promoters. The selection of a suitable promoter will be apparent to those skilled in the art from the teachings contained herein, and may be from among either regulated promoters or promoters as described above.

[0124] The retroviral plasmid vector is employed to transduce packaging cell lines to form producer cell lines. Examples of packaging cells which may be transfected include, but are not limited to, the PE501, PA317, \$\psi\$-2, \$\psi\$-AM, PA12, T19-14X, VT-19-17-H2, \$\psi\$CRE, \$\psi\$CRIP, GP+E-86, GP+envAm12, and DAN cell lines as described in Miller, Human Gene Therapy, 1:5-14 (1990), which is incorporated herein by reference in its entirety. The vector may transduce the packaging cells through any means known in the art. Such means include, but are not limited to, electroporation, the use of liposomes, and calcium phosphate precipitation. In one alternative, the retroviral plasmid vector may be encapsulated into a liposome, or coupled to a lipid, and then administered to a host.

[0125] The producer cell line generates infectious retroviral vector particles that include the nucleic acid sequence(s) encoding the PTP polypeptides or other polypeptide of interest and fusion proteins thereof. Such retroviral vector particles then may be employed, to transduce eukaryotic cells, either in vitro or in vivo. The transduced eukaryotic cells will express the nucleic acid sequence(s) encoding the siRNA polynucleotide that is capable of specifically interfering with expression of a polypeptide or fusion protein. Eukaryotic cells which may be transduced include, but are not limited to, embryonic stem cells, embryonic carcinoma cells, as well as hematopoietic stem cells, hepatocytes, fibroblasts, myoblasts, keratinocytes, endothelial cells, bronchial epithelial cells and various other culture-adapted cell lines.

[0126] In another aspect, the present invention relates to host cells containing the above described recombinant PTP expression constructs and to host cells containing the above described recombinant expression constructs comprising a (non-PTP) polypeptide of interest as described herein. Host cells are genetically engineered (transduced, transformed or transfected) with the vectors and/or expression constructs of this invention that may be, for example, a cloning vector, a shuttle vector, or an expression construct. The vector or construct may be, for example, in the form of a plasmid, a viral particle, a phage, etc. The engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying particular genes such as genes encoding siRNA polynucleotides or fusion proteins thereof. The culture conditions for particular host cells selected for expression, such as temperature, pH and the like, will be readily apparent to the ordinarily skilled artisan.

[0127] The host cell can be a higher eukaryotic cell, such as a mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or the host cell can be a prokaryotic cell, such as a bacterial cell. Representative examples of appropriate host

cells according to the present invention include, but need not be limited to, bacterial cells, such as *E. coli*, Streptomyces, *Salmonella typhimurium*; fungal cells, such as yeast; insect cells, such as Drosophila S2 and Spodoptera S19; animal cells, such as CHO, COS or 293 cells; adenoviruses; plant cells, or any suitable cell already adapted to in vitro propagation or so established de novo. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein.

Various mammalian cell culture systems can also be employed to produce siRNA polynucleotides from recombinant nucleic acid constructs of the present invention. The invention is therefore directed in part to a method of producing a siRNA polynucleotide, by culturing a host cell comprising a recombinant nucleic acid construct that comprises at least one promoter operably linked to a nucleic acid sequence encoding a siRNA polynucleotide specific for a desired target polypeptide. In certain embodiments, the promoter may be a regulated promoter as provided herein, for example a tetracylcine-repressible promoter. In certain embodiments the recombinant expression construct is a recombinant viral expression construct as provided herein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts, described by Gluzman, Cell 23:175 (1981), and other cell lines capable of expressing a compatible vector, for example, the C127, 3T3, CHO, HeLa, HEK, and BHK cell lines. Mammalian expression vectors will comprise an origin of replication, a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking nontranscribed sequences, for example as described herein regarding the preparation of recombinant siRNA polynucleotide constructs. DNA sequences derived from the SV40 splice, and polyadenylation sites may be used to provide the required nontranscribed genetic elements. Introduction of the construct into the host cell can be effected by a variety of methods with which those skilled in the art will be familiar, including but not limited to, for example, liposomes including cationic liposomes, calcium phosphate transfection, DEAF-Dextran mediated transfection, or electroporation (Davis et al., 1986 Basic Methods in Molecular Biology), or other suitable technique.

[0129] The expressed recombinant siRNA polynucleotides may be useful in intact host cells; in intact organelles such as cell membranes, intracellular vesicles or other cellular organelles; or in disrupted cell preparations including but not limited to cell homogenates or lysates, microsomes, uni- and multilamellar membrane vesicles or other preparations. Alternatively, expressed recombinant siRNA polynucleotides can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps.

[0130] Samples

[0131] According to the present invention, a method is provided for interfering with expression of a desired target

polypeptide as provided herein, comprising contacting a siRNA polynucleotide with a cell that is capable of expressing the target polypeptide, typically in a biological sample or in a subject or biological source. A "sample" as used herein refers to a biological sample containing at least one protein tyrosine phosphatase or a MAP kinase kinase or a chemotherapeutic target polypeptide, and may be provided by obtaining a blood sample, biopsy specimen, tissue explant, organ culture or any other tissue or cell preparation from a subject or a biological source. A sample may further refer to a tissue or cell preparation in which the morphological integrity or physical state has been disrupted, for example, by dissection, dissociation, solubilization, fractionation, homogenization, biochemical or chemical extraction, pulverization, lyophilization, sonication or any other means for processing a sample derived from a subject or biological source. In certain preferred embodiments, the sample is a cell that comprises at least one PTP and/or at least one MAP kinase, and/or at least one MAP kinase kinase, and in certain particularly preferred embodiments the cell comprises an inducible biological signaling pathway, at least one component of which is a specific target polypeptidee. In particularly preferred embodiments the cell is a mammalian cell, for example, Rat-1 fibroblasts, COS cells, CHO cells, HEK-293 cells, HepG2, HII4E-C3, L6, and 3T3-L1, or other well known model cell lines, which are available from the American Type Culture Collection (ATCC, Manassas, Va.). In other preferred embodiments, the cell line is derived from PTP-1B knockout animals and which may be transfected with human insulin receptor (HIR), for example, 1BKO mouse embryo fibroblasts.

[0132] In certain other preferred embodiments the sample is a cell that comprises a chemotherapeutic target polypeptide, which includes, for example, a cell line that is derived from a tumor cell. The cell line may be a primary tumor cell line, that is, a cell line prepared directly from a tumor sample removed from a human or a non-human animal. Alternatively, the cell line may be one of several established tumor cell lines known in the art, including but not limited to MCF7, T47D, SW620, HS578T, MDA-MB-435, MDA MB 231, HCT-116, HT-29, HeLa, Raji, Ramos, and the like (see ATCC collection).

[0133] The subject or biological source may be a human or non-human animal, a primary cell culture or culture adapted cell line including but not limited to genetically engineered cell lines that may contain chromosomally integrated or episomal recombinant nucleic acid sequences, immortalized or immortalizable cell lines, somatic cell hybrid cell lines, differentiated or differentiatable cell lines, transformed cell lines and the like. Optionally, in certain situations it may be desirable to treat cells in a biological sample with hydrogen peroxide and/or with another agent that directly or indirectly promotes reactive oxygen species (ROS) generation, including biological stimuli as described herein; in certain other situations it may be desirable to treat cells in a biological sample with a ROS scavenger, such as N-acetyl cysteine (NAC) or superoxide dismutase (SOD) or other ROS scavengers known in the art; in other situations cellular glutathione (GSH) may be depleted by treating cells with L-buthionine-SR-sulfoximine (Bso); and in other circumstances cells may be treated with pervanadate to enrich the sample in tyrosine phosphorylated proteins. Other means may also be employed to effect an increase in the population of tyrosine phosphorylated proteins present in the sample,

including the use of a subject or biological source that is a cell line that has been transfected with at least one gene encoding a protein tyrosine kinase.

[0134] Additionally or alternatively, a biological signaling pathway may be induced in subject or biological source cells by contacting such cells with an appropriate stimulus, which may vary depending upon the signaling pathway under investigation, whether known or unknown. For example, a signaling pathway that, when induced, results in protein tyrosine phosphorylation and/or protein tyrosine dephosphorylation may be stimulated in subject or biological source cells using any one or more of a variety of well known methods and compositions known in the art to stimulate protein tyrosine kinase (PTK) and/or PTP activity. These stimuli may include, without limitation, exposure of cells to cytokines, growth factors, hormones, peptides, small molecule mediators, cell stressors (e.g., ultraviolet light; temperature shifts; osmotic shock; ROS or a source thereof, such as hydrogen peroxide, superoxide, ozone, etc. or any agent that induces or promotes ROS production (see, e.g., Halliwell and Gutteridge, Free Radicals in Biology and Medicine (3rd Ed.) 1999 Oxford University Press, Oxford, UK); heavy metals; alcohol) or other agents that induce PTK-mediated protein tyrosine phosphorylation and/or PTP-mediated phosphoprotein tyrosine dephosphorylation. Such agents may include, for example, interleukins (e.g., IL-1, IL-3), interferons (e.g., IFN-γ), human growth hormone, insulin, epidermal growth factor (EGF), platelet derived growth factor (PDGF), granulocyte colony stimulating factor (G-CSF), granulocyte-megakaryocyte colony stimulating factor (GM-CSF), transforming growth factor (e.g., TGF- β 1), tumor necrosis factor (e.g., TNF- α) and fibroblast growth factor (FGF; e.g., basic FGF (bFGF)), any agent or combination of agents capable of triggering T lymphocyte activation via the T cell receptor for antigen (TCR; TCR-inducing agents may include superantigens, specifically recognized antigens and/or MHC-derived peptides, MHC peptide tetramers (e.g., Altman et al., 1996 Science 274:94-96); TCR-specific antibodies or fragments or derivatives thereof), lectins (e.g., PHA, PWM, ConA, etc.), mitogens, G-protein coupled receptor agonists such as angiotensin-2, thrombin, thyrotropin, parathyroid hormone, lysophosphatidic acid (LPA), sphingosine-1-phosphate, serotonin, endothelin, acetylcholine, platelet activating factor (PAF) or bradykinin, as well as other agents with which those having ordinary skill in the art will be familiar (see, e.g., Rhee et al., [online] Oct. 10, 2000 Science's stke, Internet: URL<www.stke.org/cgl/content/full/OCsigtrans;2000/53/pel>), and references cited therein).

[0135] As noted above, regulated tyrosine phosphorylation contributes to specific pathways for biological signal transduction, including those associated with cell division, cell survival, apoptosis, proliferation and differentiation, and "inducible signaling pathways" in the context of the present invention include transient or stable associations or interactions among molecular components involved in the control of these and similar processes in cells. Depending on the particular pathway of interest, an appropriate parameter for determining induction of such pathway may be selected. For example, for signaling pathways associated with cell proliferation, a variety of well known methodologies are available for quantifying proliferation, including, for example, incorporation of tritiated thymidine into cellular DNA, monitoring of detectable (e.g., fluorimetric or colorimetric)

indicators of cellular respiratory activity, (e.g., MTT assay) or cell counting, or the like. Similarly, in the cell biology arts there are known multiple techniques for assessing cell survival (e.g., vital dyes, metabolic indicators, etc.) and for determining apoptosis (e.g., annexin V binding, DNA fragmentation assays, caspase activation, PARP cleavage, etc.). Other signaling pathways will be associated with particular cellular phenotypes, for example specific induction of gene expression (e.g., detectable as transcription or translation products, or by bioassays of such products, or as nuclear localization of cytoplasmic factors), altered (e.g., statistically significant increases or decreases) levels of intracellular mediators (e.g., activated kinases or phosphatases, altered levels of cyclic nucleotides or of physiologically active ionic species, etc.), altered cell cycle profiles, or altered cellular morphology, and the like, such that cellular responsiveness to a particular stimulus as provided herein can be readily identified to determine whether a particular cell comprises an inducible signaling pathway.

[0136] In preferred embodiments where a siRNA of the invention is being used to interfere with expression of a target polypeptide that is a PTP or that is a component of a biological signaling pathway that comprises a PTP, a PTP substrate may be any naturally or non-naturally occurring phosphorylated peptide, polypeptide or protein that can specifically bind to and/or be dephosphorylated by a PTP (including dual specificity phosphatases) as provided herein, or any other phosphorylated molecule that can be a substrate of a PTP family member as provided herein. Non-limiting examples of known PTP substrates include the proteins VCP (see, e.g., Zhang et al., 1999 J. Biol. Chem. 274:17806, and references cited therein), p130cas, EGF receptor, p210 ber:abl, MAP kinase, She (Tiganis et al., 1998 Mol Cell. Biol. 18:1622-1634), insulin receptor, lck (lymphocyte specific protein tyrosine kinase, Marth et al., 1985 Cell 43:393), T cell receptor zeta chain, and phosphatidylinositol 3,4,5triphosphate (Maehama et al., 1998 J. Biol. Chem. 273:13375).

[0137] Identification and selection of PTP substrates as provided herein, for use in the present invention, may be performed according to procedures with which those having ordinary skill in the art will be familiar, or may, for example, be conducted according to the disclosures of WO 00/75339, U.S. application Ser. No. 09/334,575, or U.S. application Ser. No. 10/366,547, and references cited therein. The phosphorylated protein/PTP complex may be isolated, for example, by conventional isolation techniques as described in U.S. Pat. No. 5,352,660, including salting out, chromatography, electrophoresis, gel filtration, fractionation, absorption, polyacrylamide gel electrophoresis, agglutination, combinations thereof or other strategies. PTP substrates that are known may also be prepared according to well known procedures that employ principles of molecular biology and/or peptide synthesis (e.g., Ausubel et al., Current Protocols in Molecular Biology, Greene Publ. Assoc. Inc. & John Wiley & Sons, Inc., Boston, Mass. (1993); Sambrook et al., Molecular Cloning, Third Ed., Cold Spring Harbor Laboratory, Plainview, N.Y. (2001); Fox, Molec. Biotechnol. 3:249 (1995); Maeji et al., Pept. Res. 8:33 (1995)).

[0138] The PTP substrate peptides of the present invention may therefore be derived from PTP substrate proteins, polypeptides and peptides as provided herein having amino acid sequences that are identical or similar to tyrosine

phosphorylated PTP substrate sequences known in the art. For example by way of illustration and not limitation, peptide sequences derived from the known PTP substrate proteins referred to above are contemplated for use according to the instant invention, as are peptides having at least 70% similarity (preferably 70% identity), more preferably 80% similarity (more preferably 80% identity), more preferably 90% similarity (more preferably 90% identity) and still more preferably 95% similarity (still more preferably 95% identity) to the polypeptides described in references cited herein and in the Examples and to portions of such polypeptides as disclosed herein. As known in the art "similarity" between two polypeptides is determined by comparing the amino acid sequence and conserved amino acid substitutes thereto of the polypeptide to the sequence of a second polypeptide (e.g., using GENEWORKS, Align or the BLAST algorithm, or another algorithm, as described above).

[0139] In certain preferred embodiments of the present invention, the siRNA polynucleotide and/or the PTP substrate is detectably labeled, and in particularly preferred embodiments the siRNA polynucleotide and/or PTP substrate is capable of generating a radioactive or a fluorescent signal. The siRNA polynucleotide and/or PTP substrate can be detectably labeled by covalently or non-covalently attaching a suitable reporter molecule or moiety, for example a radionuclide such as ³²P (e.g., Pestka et al., 1999 Protein Expr. Purif. 17:203-14), a radiohalogen such as iodine [125] or ¹³¹I] (e.g., Wilbur, 1992 *Bioconjug. Chem.* 3:433-70), or tritium [3H]; an enzyme; or any of various luminescent (e.g., chemiluminescent) or fluorescent materials (e.g., a fluorophore) selected according to the particular fluorescence detection technique to be employed, as known in the art and based upon the present disclosure. Fluorescent reporter moieties and methods for labeling siRNA polynucleotides and/or PTP substrates as provided herein can be found, for example in Haugland (1996 Handbook of Fluorescent Probes and Research Chemicals—Sixth Ed., Molecular Probes, Eugene, Oreg.; 1999 Handbook of Fluorescent Probes and Research Chemicals-Seventh Ed., Molecular Probes, Eugene, Oreg., Internet: http://www.probes.com/ lit/) and in references cited therein. Particularly preferred for use as such a fluorophore in the subject invention methods are fluorescein, rhodamine, Texas Red, AlexaFluor-594, AlexaFluor-488, Oregon Green, BODIPY-FL, umbelliferone, dichlorotriazinylamine fluorescein, dansyl chloride, phycoerythrin or Cy-5. Examples of suitable enzymes include, but are not limited to, horseradish peroxidase, biotin, alkaline phosphatase, β-galactosidase and acetylcholinesterase. Appropriate luminescent materials include luminol, and suitable radioactive materials include radioactive phosphorus [32P]. In certain other preferred embodiments of the present invention, a detectably labeled siRNA polynucleotide comprises a magnetic particle, for example a paramagnetic or a diamagnetic particle or other magnetic particle or the like (preferably a microparticle) known to the art and suitable for the intended use. Without wishing to be limited by theory, according to certain such embodiments there is provided a method for selecting a cell that has bound, adsorbed, absorbed, internalized or otherwise become associated with a siRNA polynucleotide that comprises a magnetic particle. For example, selective isolation of a population or subpopulation of cells containing one or more PTP-specific siRNA polynucleotide-magnetic particle conjugates may offer certain advantages in the further characterization or regulation of PTP signaling pathways.

[0140] In certain embodiments of the present invention, particular PTP-specific siRNA polynucleotides of interest may be identified by contacting a candidate siRNA polynucleotide with a sample comprising a cell that comprises a target polypeptide-encoding gene and that is capable of target polypeptide gene transcription or expression (e.g., translation), under conditions and for a time sufficient to detect such gene transcription or expression, and comparing target transcription levels, polypeptide expression and/or functional expression (e.g., PTP catalytic activity) in the absence and presence of the candidate siRNA polynucleotide. Preferably target transcription or expression is decreased in the presence of the siRNA polynucleotide, which in the case of targets that are PTPs provides an alternative to PTP active site directed approaches to modulating PTP activity. (The invention need not be so limited, however, and contemplates other embodiments wherein transcription and/or expression levels of a signal transduction component other than that which is specifically targeted by the siRNA may be increased in the presence of a certain target-specific siRNA polynucleotide. By way of non-limiting theory, such an increase may result from a cellular compensatory mechanism that is induced as a result of the siRNA.)

[0141] Activity of a siRNA target polypeptide of interest may also be measured in whole cells transfected with a reporter gene whose expression is dependent upon the activation of an appropriate substrate. For example, appropriate cells (i.e., cells that express the target polypeptide and that have also been transfected with a target-specific siRNA polynucleotide that is either known or suspected of being capable of interfering with target polypeptide expression) may be transfected with a substrate-dependent promoter linked to a reporter gene. In such a system, expression of the reporter gene (which may be readily detected using methods well known to those of ordinary skill in the art) depends upon activation of the substrate via its interaction with the target polypeptide. For example, dephosphorylation of substrate may be detected based on a decrease in reporter activity in situations where the target polypeptide regulates substrate phosphorylation.

[0142] Within other aspects, the present invention provides animal models in which an animal, by virtue of introduction of an appropriate target polypeptide-specific siRNA polynucleotide, for example, as a transgene, does not express (or expresses a significantly reduced amount of) a functional PTP. Such animals may be generated, for example, using standard homologous recombination strategies, or alternatively, for instance, by oocyte microinjection with a plasmid comprising the siRNA-encoding sequence that is regulated by a suitable promoter (e.g., ubiquitous or tissue-specific) followed by implantation in a surrogate mother. Animal models generated in this manner may be used to study activities of PTP signaling pathway components and modulating agents in vivo.

[0143] Therapeutic Methods

[0144] One or more siRNA polynucleotides capable of interfering with target polypeptide expression and identified according to the above-described methods may also be used to modulate (e.g., inhibit or potentiate) target polypeptide

activity in a patient. As used herein, a "patient" may be any mammal, including a human, and may be afflicted with a condition associated with undesired target polypeptide activity or may be free of detectable disease. Accordingly, the treatment may be of an existing disease or may be prophylactic. Conditions associated with signal transduction and/or with inappropriate activity of specific siRNA target polypeptides described herein include obesity, impaired glucose tolerance and diabetes and cancer, disorders associated with cell proliferation, including cancer, graft-versushost disease (GVHD), autoimmune diseases, allergy or other conditions in which immunosuppression may be involved, metabolic diseases, abnormal cell growth or proliferation and cell cycle abnormalities.

[0145] For administration to a patient, one or more specific siRNA polynucleotides, either alone, with or without chemical modification or removal of ribose, or comprised in an appropriate vector as described herein (e.g., including a vector which comprises a DNA sequence from which a specific siRNA can be transcribed) are generally formulated as a pharmaceutical composition. A pharmaceutical composition may be a sterile aqueous or non-aqueous solution, suspension or emulsion, which additionally comprises a physiologically acceptable carrier (i.e., a non-toxic material that does not interfere with the activity of the active ingredient). Such compositions may be in the form of a solid, liquid or gas (aerosol). Alternatively, compositions of the present invention may be formulated as a lyophilizate or compounds may be encapsulated within liposomes using well known technology. Pharmaceutical compositions within the scope of the present invention may also contain other components, which may be biologically active or inactive. Such components include, but are not limited to, buffers (e.g., neutral buffered saline or phosphate buffered saline), carbohydrates (e.g., glucose, mannose, sucrose or dextrans), mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, chelating agents such as EDTA or glutathione, stabilizers, dyes, flavoring agents, and suspending agents and/or preservatives.

[0146] Any suitable carrier known to those of ordinary skill in the art may be employed in the pharmaceutical compositions of the present invention. Carriers for therapeutic use are well known, and are described, for example, in Remingtons Pharmaceutical Sciences, Mack Publishing Co. (A. R. Gennaro ed. 1985). In general, the type of carrier is selected based on the mode of administration. Pharmaceutical compositions may be formulated for any appropriate manner of administration, including, for example, topical, oral, nasal, intrathecal, rectal, vaginal, sublingual or parenteral administration, including subcutaneous, intraveintramuscular, intrasternal, intracavernous, intrameatal or intraurethral injection or infusion. For parenteral administration, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, kaolin, glycerin, starch dextrins, sodium alginate, carboxymethylcellulose, ethyl cellulose, glucose, sucrose and/or magnesium carbonate, may be employed.

[0147] A pharmaceutical composition (e.g., for oral administration or delivery by injection) may be in the form of a liquid (e.g., an elixir, syrup, solution, emulsion or

suspension). A liquid pharmaceutical composition may include, for example, one or more of the following: sterile diluents such as water for injection, saline solution, preferably physiological saline, Ringer's solution, isotonic sodium chloride, fixed oils such as synthetic mono or diglycerides which may serve as the solvent or suspending medium, polyethylene glycols, glycerin, propylene glycol or other solvents; antibacterial agents such as benzyl alcohol or methyl paraben; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. A parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic. The use of physiological saline is preferred, and an injectable pharmaceutical composition is preferably sterile.

[0148] The compositions described herein may be formulated for sustained release (i.e., a formulation such as a capsule or sponge that effects a slow release of compound following administration). Such compositions may generally be prepared using well known technology and administered by, for example, oral, rectal or subcutaneous implantation, or by implantation at the desired target site. Sustained-release formulations may contain an agent dispersed in a carrier matrix and/or contained within a reservoir surrounded by a rate controlling membrane. Carriers for use within such formulations are biocompatible, and may also be biodegradable; preferably the formulation provides a relatively constant level of active component release. The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

[0149] Within a pharmaceutical composition, a therapeutic agent comprising a polypeptide-directed siRNA polynucleotide as described herein (or, e.g., a recombinant nucleic acid construct encoding a siRNA polynucleotide) may be linked to any of a variety of compounds. For example, such an agent may be linked to a targeting moiety (e.g., a monoclonal or polyclonal antibody, a protein or a liposome) that facilitates the delivery of the agent to the target site. As used herein, a "targeting moiety" may be any substance (such as a compound or cell) that, when linked to an agent enhances the transport of the agent to a target cell or tissue, thereby increasing the local concentration of the agent. Targeting moieties include antibodies or fragments thereof, receptors, ligands and other molecules that bind to cells of, or in the vicinity of, the target tissue. An antibody targeting agent may be an intact (whole) molecule, a fragment thereof, or a functional equivalent thereof. Examples of antibody fragments are F(ab')2, Fab', Fab and F[v] fragments, which may be produced by conventional methods or by genetic or protein engineering. Linkage is generally covalent and may be achieved by, for example, direct condensation or other reactions, or by way of bi- or multifunctional linkers. Targeting moieties may be selected based on the cell(s) or tissue(s) toward which the agent is expected to exert a therapeutic benefit.

[0150] Pharmaceutical compositions may be administered in a manner appropriate to the disease to be treated (or prevented). An appropriate dosage and a suitable duration and frequency of administration will be determined by such

factors as the condition of the patient, the type and severity of the patient's disease, the particular form of the active ingredient and the method of administration. In general, an appropriate dosage and treatment regimen provides the agent(s) in an amount sufficient to provide therapeutic and/or prophylactic benefit (e.g., an improved clinical outcome, such as more frequent complete or partial remissions, or longer disease-free and/or overall survival, or a lessening of symptom severity). For prophylactic use, a dose should be sufficient to prevent, delay the onset of or diminish the severity of a disease associated with cell proliferation.

[0151] Optimal dosages may generally be determined using experimental models and/or clinical trials. In general, the amount of siRNA polynucleotide present in a dose, or produced in situ by DNA present in a dose (e.g., from a recombinant nucleic acid construct comprising a siRNA polynucleotide), ranges from about 0.01 µg to about 1001 g per kg of host, typically from about 0.1 µg to about 10 µg. The use of the minimum dosage that is sufficient to provide effective therapy is usually preferred. Patients may generally be monitored for therapeutic or prophylactic effectiveness using assays suitable for the condition being treated or prevented, which will be familiar to those having ordinary skill in the art. Suitable dose sizes will vary with the size of the patient, but will typically range from about 10 mL to about 500 mL for 10-60 kg animal.

[0152] The following Examples are offered by way of illustration and not by way of limitation.

EXAMPLE 1

Interference of Dual Specificity Phosphatase Expression by Small Interfering RNA

[0153] This example describes the effect on dual specificity phosphatase (DSP) expression in cells transfected with sequence-specific small interfering RNA (siRNA) polynucleotides. Interference with expression of MKP-1 and DSP-3 was examined by transfecting sequence-specific siR-NAs into mammalian cells expressing the DSP polypeptide and then detecting expression by immunoblot.

[0154] The siRNA nucleotide sequences specific for each DSP were chosen by first scanning the open reading frame of the target cDNA for 21-base sequences that were flanked on the 5' end by two adenine bases (AA) and that had A+T/G+C ratios that were nearly 1:1. Twenty-one-base sequences with an A+T/G+C ratio greater than 2:1 or 1:2 were excluded. If no 21-base sequences were identified that met this criteria, the polynucleotide sequence encoding the DSP was searched for a 21-base sequence having the bases CA at the 5' end. The polynucleotide sequences examined were the sequences encoding DSP-3 polypeptide (SEQ ID) and MKP-1 (SEQ ID NO: __). For the selection of sequences for some of the siRNA polynucleotides, the sense and antisense sequences of each 21-mer that met the above criteria were then analyzed to determine if the sequence had the potential to form an internal hairpin loop or homodimer. Such an analysis can be performed using computer software programs known to those in the art. Any 21-mer that had an internal hairpin loop melting temperature of greater than 55° C. and a homodimer melting temperature of greater than 37° C. was excluded. The specificity of each 21-mer was determined by performing a BLAST search of public databases. Sequences that contained at least 16 of 21 consecutive nucleotides with 100% identity with a polynucleotide sequence other than the target sequence were not used in the experiments. In each of the Examples provided herein, each siRNA sequence represents the sense strand of the siRNA polynucleotide and its corresponding sequence identifier. "Related sequence identifiers" referred to in the Examples identify sequences in the sequence listing that contain the same nucleotides at positions 1-19 of the siRNA sequence with and without two additional nucleotides (NN) at the 3' end (which would correspond to a two-nucleotide overhang in a double stranded polynucleotide), and the reverse complement of each. Unless otherwise stated, it is to be understood that the siRNA transfected into a cell is composed of the sense strand and its complementary antisense strand, which form a duplex siRNA polynucleotide. The sequences chosen for these experiments were as follows.

[0155] DSP-3 Specific:

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DSP3.1:
5'-cgauagugccaggccuaugtt-3' [SEQ ID NO:__]
DSP3.2:
5'-gcaugagguccaucaguautt-3' [SEQ ID NO:__]
DSP3.3:
5'-cgauacugccaggccaugtt-3' [SEQ ID NO:__]
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[0156] MKP-1 Specific:

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MKP.1: 5'-auccugeccuuucuguacett-3' [SEQ ID NO:___]

MKP.2: 5'-gcagaggcaaagcaucauctt-3' [SEQ ID NO:___]
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[0157] Sense and antisense oligonucleotides for MKP.1, MKP.2, DSP3.1, DSP3.2, and DSP3.3 were synthesized according to the standard protocol of the vendor (Dharmacon Research, Inc., Lafayette, Colo.). For some experiments described in this and other examples, the vendor gel-purified the double-stranded siRNA polynucleotide, which was then used. In the instances when the vendor did not prepare double-stranded siRNA, just before transfection, double-stranded siRNAs were prepared by annealing the sense and anti-sense oligonucleotides in annealing buffer (100 mM potassium acetate, 30 mM HEPES-KOH, pH 7.4, 2 mM magnesium acetate) for 1 minute at 90° C., followed by a 60 minute incubation at 37° C.

[0158] Recombinant nucleic acid expression vectors containing encoding sequences for the MKP-1 polypeptide and DSP-3 polypeptide were prepared according to standard molecular biology techniques. Polynucleotides comprising the MKP-1 coding sequence of SEQ ID NO: ____ and comprising the DSP-3 coding sequence of SEQ ID NO: ____ were cloned into recombinant expression vectors according to methods known to those skilled in the molecular biology art.

[0159] HeLa cells (ATCC, Manassas, Va.) were maintained in Dulbecco's modified Eagle's medium (DMEM, Life Technologies, Inc., Gaithersburg, Md.) plus 10% fetal bovine serum (FBS), 100 units/ml penicillin, and 100 μ g/ml streptomycin. Cells were plated in 6-well tissue culture plates at a density of approximately 5×10^4 cells per well at the time of transfection.

[0160] HeLa cells were transfected with 60 pmoles of MKP.1, MKP.2, or CD45.1 (SEQ ID NO: For each cell culture well, the siRNA polynucleotides were diluted into 250 μ l of O_{PTI}MEM® Reduced Serum Medium (Gibco™, Life Technologies), and 15 µl Oligofectamine™ (Invitrogen Life Technologies, Carlsbad, Calif.) was diluted into 250 μ l of O_{PTI}MEM®. A control solution without siRNA was also prepared. Each solution was incubated at room temperature for 5 minutes. The two solutions were mixed and then incubated for 20 minutes at room temperature to allow the liposome-nucleic acid complexes to form. FBS-containing media was removed from the HeLa cell cultures and replaced with OPTIMEM®. The liposomenucleic acid mixture then was added to the HeLa cell culture, and the transfected cells incubated at 37° C. for 22-24 hours. Media were removed from the cell cultures and replaced with DMEM containing 10% FBS. Cells were incubated at 37° C. in the media plus FBS solution for 0, 1, or 4 hours.

[0161] Expression of MKP-1 was analyzed by immunoblotting HeLa cell extracts. The cells were rinsed twice in phosphate buffered saline (PBS) (4° C.) and then lysed in 250 µl of ice-cold RIPA buffer RIPA buffer (150 mM NaCl, 10 mM NaPO₄, 2 mM EDTA, 1% deoxycholate, 1% Nonidet® P40, 0.1% SDS, 5 mM NaF, 14.3 mM beta-mercaptoethanol, and Complete Protease Inhibitor (Roche Applied Bioscience, Indianapolis, Ind.). The lysates were centrifuged and aliquots of supernatant (10 μ l) from each transfected cell culture sample were combined with 10 μ l of 2×SDS-PAGE reducing sample buffer. The samples were heated at 95° C. for five minutes, and then applied to a 14% Tris-glycine SDS-PAGE gel (NOVEX® from Invitrogen Life Technologies, Carlsbad, Calif.). After electrophoresis, the separated proteins were electrophoretically transferred from the gel onto an Immobilon-P polyvinylidene fluoride (PVDF) membrane (Millipore, Bedford, Mass.). The PVDF membrane was blocked in 5% milk in TBST (20 mM Tris pH 7.5, 150 mM NaCl, 0.05% Tween-20), incubated with an anti-MKP-1 antibody (Santa Cruz Biotechnology, Inc., Santa Cruz, Calif.) for 2-16 hours at room temperature, washed 3×10 minutes with TBST, and then incubated with an appropriate horseradish peroxidase (HRP) conjugate IgG (1:10,000) (Amersham Biosciences, Piscataway, N.J.) for 30 minutes at room temperature. Binding was detected with the ECL chemiluminescent reagent used according to the manufacturer's instructions (Amersham Biosciences, Piscataway, N.J.) as shown in FIG. 1 (upper). A second SDS-PAGE gel in which the HeLa cell extracts were separated was stained with Coomassie Blue (FIG. 1, lower).

[0162] Interference with DSP-3 polypeptide expression was analyzed in HeLa cells transfected with siRNA polynucleotides. To determine the transfection efficiency of a siRNA polynucleotide, HeLa cells cultured as described above were plated at different cell densities and then transfected with a sequence-specific siRNA. DSP3.1 siRNA (SEQ ID NO:) was synthesized and conjugated to fluorescein isothiocyanate (FITC) according to the vendor's standard methods (Synthetic Genetics, San Diego, Calif.). HeLa cells plated at varying cell densities to achieve approximately 1×10⁴ cells/well, 3×10⁴ cells/well, 5×10⁴ cells/well, 1×10⁵ cells/well, 2×10⁵ cells/well, and 4×10⁵ cells/well were transfected with FITC-DSP3.1 as described above. Controls included HeLa cells exposed to Lipofectamine™ 2000 alone and to media alone. The transfected cells were harvested after 24-48 hours and analyzed by a fluorescence-activated cell sorter (FACS). Transfection was more efficient at cell densities of 5×10^4 cells/well or less.

[0163] Interference of DSP-3 expression by two different DSP-3 sequence specific siRNA polynucleotides, DSP3.1 (SEQ ID NO: _____) and DSP3.2 (SEQ ID NO: _____). Transfection of HeLa cells was performed as described for MKP-1. As controls, HeLa cells were transfected with non-specific MKP.1 (SEQ ID NO: _____) and with transfection solution not containing the expression vector or siRNA.

[0164] Twenty-four hours after transfection, cell extracts were prepared either using RIPA buffer (see above) or 1% Triton X-100®. The extracts were analyzed by immunoblot (see above) using an anti-DSP-3 monoclonal antibody, clone 17, diluted 1:10,000 in TBST and binding was detected with HRP-conjugated anti-mouse IgG. DSP3.1 effectively decreased expression of DSP-3, whereas the level of expression in cells transfected with siRNA DSP3.2 was comparable to expression in the cells transfected with the nonspecific MKP.1 siRNA. The cell extracts were also immunoblotted against an anti-PTP1B antibody, which demonstrated that protein expression of another protein expressed in the cells was not affected by the presence of siRNA polynucleotides. The data suggest that the decrease in the level of DSP-3 expression varies depending upon the particular sequence of the siRNA.

[0165] To evaluate the sensitivity of interference by specific siRNA polynucleotides, DSP3.1 siRNA (SEQ ID) was titrated in HeLa cells. HeLa cells were transfected as described above with DSP3.1 siRNA (SEQ ID NO:1) at a concentration of 1, 2, 5, 10, 20, and 100 nM. HeLa cells were also transfected at the same concentrations with non-specific siRNAs, cdc14a.1 (5'-caucgugcgaagguuccugtt-3' (SEQ ID NO:6)) and CD45.2 (5'-gccgagaacaaaguggaugtt-3' (SEQ ID NO: _)). An immunoblot of cell extracts prepared using RIPA buffer was probed with anti-DSP-3 monoclonal antibody clone 17. A second immunoblot was probed with an anti-JNK2 antibody. DSP-3 expression decreased to approximately the same level in cells transfected with 5, 10, 20, and 100 nM of the specific siRNA DSP3.1. The level of expression of DSP-3 also decreased in the presence of the lowest concentrations of siRNA DSP3.1 compared with DSP-3 expression in cells transfected with non-specific siRNAs. Expression of JNK2 was not affected.

[0166] The specificity of siRNA interference was demonstrated by co-transfecting HeLa cells with the DSP-3 expression vector and an siRNA, DSP3.3 (SEQ ID NO:____) that had two base differences from siDSP3.1. Transfection and immunoblotting were performed as described above for the titration experiment. The expression levels of DSP-3 polypeptide was effectively decreased in the presence of 1, 5, 10, 20, or 100 nM of DSP3.1 but not in cells transfected with DSP3.3. The level of expression of JNK2 was not affected.

EXAMPLE 2

Interference with Expression of Protein Tyrosine Phosphatases by Sequence-Specific Small Interfering RNA

[0167] This example describes RNA interference of transient and endogenous expression of various protein tyrosine phosphatases (PTPs).

[0168] Co-Transfection Assays to Determine Interference of PTP Expression by siRNA

[0169] DSP-11 and DSP-18

[0170] Interference of expression of FLAG®-tagged DSP-11 polypeptide and FLAG®-tagged DSP-18pr polypeptide (DSP-18) by sequence specific siRNA polynucleotides was determined. (FLAG® sequence: Asp-Tyr-Lys-Asp-Asp-Asp-Asp-Lys (SEQ ID NO:_____)) (Sigma Aldrich, St. Louis, Mo.). Two siRNA sequences that were specific for DSP-11 polynucleotide (SEQ ID NO: encoding a DSP-11 polypeptide (SEQ ID NO:) and two siRNA sequences specific for DSP-18pr polynucleotide (DSP-18, SEQ ID NO:_ ____) encoding a DSP-18 polypeptide (SEQ) were designed using the criteria described in Example 1. The following sequences were used in the experiments.

[0171] DSP-11 Specific:

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DSP11.2:
5'-cuggcaccaugcuggccugtt-3' [SEQ ID NO:__]

DSP11.4:
5'-agcagucuuccaguucuactt-3' [SEQ ID NO:__]
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[0172] DSP-18 Specific:

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DSP18.2:
5'-cugccuugugcacugcuuutt-3' [SEQ ID NO:___]
DSP18.4:
5'-gaguuuggcugggccaguutt-3' [SEQ ID NO:___]
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[0173] Vectors for expression of DSP-18 and DSP-11 were prepared as follows. Vector pCMVTag2B (Stratagene, La Jolla, Calif.) was digested with restriction endonuclease BamHI (New England Biolabs, Beverly, Mass.) for 3 hours at 37° C. The digested vector was then incubated with Klenow polymerase (New England Biolabs) for 15 minutes at 25° C. to fill in the recessed 3' termini, followed by an incubation of 30 minutes at 37° C. with calf intestinal phosphatase (New England Biolabs). The GATEWAYTM Reading Frame Cassette B (Invitrogen, Carlsbad, Calif.) was inserted into the pCMVTag2B vector by ligation with T4 DNA ligase (Invitrogen) overnight at 16° C. according to the supplier's instructions. DB3.1TM competent E. coli cells were transformed with the ligated vector (GWpCMVTag2), and DNA was isolated by standard molecular biology methods. DSP-11 and DSP-18 constructs were prepared by ligating a polynucleotide encoding DSP-11 (SEQ ID NO:25) and a polynucleotide encoding DSP-18 (SEQ ID NO:27) into a modified bacterial pGEX-6PKG expression vector (Amersham Biosciences), referred to as pGEX-6P1, according to standard methods known in the molecular biology art. DSP-11 and DSP18 constructs and the pENTR™ 1A entry vector (Invitrogen) were digested with EcoRI (New England Biolabs) for 3 hours at 37° C. The pENTR™ 1A clone was treated with calf intestinal phosphatase for 30 minutes at 37° C., and then DSP-11 and DSP-18 constructs were inserted into separate pENTRTM vectors by ligation overnight at 16° C. with T4 DNA ligase. Vector DNA was prepared from LIBRARY EFFICIENCY® DH5αTM cells (Invitrogen) that were transformed with each construct according to the supplier's recommendation.

[0174] FLAG® epitope-tagged DSP-11 and DSP-18 polypeptides were prepared by cloning the pENTR™ 1A-DSP-18 and substrate trapping mutant constructs into the GWpCMVTag2 vector. The pENTR™ 1A constructs containing the DSP-11 and the DSP-18 polynucleotides were linearized by digesting the constructs with Vsp I (Promega Corp., Madison, Wis.) for 2 hours at 37° C. The DNA was purified using a QIAGEN PCR Purification kit (QIAGEN, Inc., Valencia, Calif.), and $30 \mu l$ (100 ng/ μl) was combined in a GATEWAYTM LR reaction with 6 µl linearized pENTRTM 1A-DSP-11, pENTRTM 1A-DSP-18, 3 μ l TE buffer, 4 μ l ClonaseTM Enzyme, and 4 μ l LR reaction buffer (Invitrogen) for 1 hour at room temperature. After addition of Proteinase K (Invitrogen) to each reaction for 10 minutes, LIBRARY EFFICIENCY® DH5α[™] cells were transformed with each expression vector. For controls, FLAG®-DSP-3 and FLAG®-cdc14b were also prepared according to the above method.

[0175] 293-HEK cells, maintained in DMEM, 10% FBS at 37° C. and 5% CO₂, were co-transfected with the FLAG®-DSP-11, FLAG®-DSP-18, FLAG®-DSP-3, and FLAG®cdc14b expression vectors and DSP11.2, DSP11.4, DSP18.2, and DSP18.4 siRNAs (20 nM) (double-stranded RNA was prepared as described in Example 1) using the Lipofectamine™ 2000 reagent (Invitrogen). After incubating the transfected cells for 22-24 hours at 37° C., cells were rinsed twice in phosphate buffered saline (PBS) (4° C.) and then lysed in 250 μ l of ice-cold RIPA buffer (see Example 1). The cell debris was pelleted and aliquots of each supernatant were separated by SDS-PAGE and immunoblotted as described in Example 1. DSP-11 and DSP-18 polypeptides were detected by probing the immunoblots with an anti-FLAG® antibody (Sigma-Aldrich, St. Louis, Mo.) followed by probing with an HRP-conjugated goat anti-mouse reagent (see Example 1). Binding of the anti-FLAG® antibody was detected by chemiluminescence development (see Example 1). FIG. 2 shows that expression of FLAG®-DSP-11 and FLAG®-DSP-18 was inhibited in the presence of sequencespecific siRNA.

[0176] DSP-13 and DSP-14

[0177] Expression constructs of DSP-13 (SEQ ID) and DSP-14 (SEQ ID NO: FLAG® epitope-tagged DSP-13 and DSP-14 polypeptides (SEQ ID NO: and SEQ ID NO: _, respectively) were prepared essentially as described above. Four siRNA sequences specific for DSP-13 polynucleotide and four siRNA sequences specific for DSP-14 were designed according to the criteria described in Example 1 except that melting temperatures were not necessarily calculated. After performing the BLAST search to analyze the specificity of a sequence, sequences that contained at least 16 consecutive nucleotides with 100% identity with a polynucleotide sequence other than the target sequence were not used in the experiments. The siRNA polynucleotides were manufactured by Dharmacon Research Inc. The sequences of the siRNA polynucleotides are as follows.

[0178] DSP-13 Specific:

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DSP13.1:
5'-cuugcgggaauucaaggaatt-3' (SEQ ID NO:____
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-continued

DSP13.2: 5'-ccgagggguacgguauauctt-3'	(SEQ ID NO:)
DSP13.3: 5'-caucaggcuggcuguaagatt-3'	(SEQ ID NO:)
DSP13.4: 5'-cauggaucuaaaugccuugtt-3'	(SEQ ID NO:)

[0179] DSP-14 Specific:

DSP-14.1: 5'-gugaagacaagccucaagatt-3'	(SEQ	ID	NO:)
DSP-14.2: 5'-gcucuacauuggcgaugagtt-3'	(SEQ	ID	NO:)
DSP-14.3: 5'-gcgacgaccacaguaagautt-3'	(SEQ	ID	NO:)
DSP-14.4: 5'-ggacaugacccugguggactt-3'	(SEQ	ID	NO:)

[0180] 293-HEK cells were co-transfected with 1-2 µg of the FLAG®-DSP-13 or FLAG®-DSP-14 expression vector and 20 nM of siRNA and expression detected by immunoblot as described above. As controls, cells co-transfected with a DSP expression vector and a non-specific siRNA and untransfected 293-HEK cells were included in the analysis.

[0181] The amount of of FLAG®-DSP-13 polypeptide expressed in 293-HEK cells co-transfected with the FLAG®-DSP-13 construct and either DSP13.3 or DSP13.4 siRNA decreased more than 95% compared with cells transfected with the DSP-13 expression constructs only. Expression of the DSP-13 polypeptide in cells co-transfected with DSP13.2 siRNA was comparable to expression in cells co-transfected with a non-specific siRNA(DSP14.1). Expression of FLAG®-DSP-14 polypeptide decreased 70% in 293-HEK cells when the cells were co-transfected with DSP14.1 siRNA and decreased 90% when the cells were co-transfected with DSP-14.3 siRNA. Expression of DSP-14 in the presence of siRNA 14.4 was only slightly lower than observed with a non-specific siRNA (DSP13.1).

[0182] DSP-3

[0183] Transient co-transfection experiments in 293-HEK cells were also performed with DSP3.1 siRNA (SEQ ID NO:1) and a DSP-3 polypeptide recombinant expression vector (prepared according to standard molecular biology techniques). Expression of DSP-3 was determined by immunoblot probed with anti-DSP-3 monoclonal antibody clone 17. The results showed that the amount of DSP-3 polypeptide expressed in the 293-HEK cells decreased 80% in the presence of sequence specific siRNA.

[0184] SHP-2

[0185] Inhibition of expression of the protein tyrosine phosphatase (PTP) SHP-2 (src homology protein-2) was also examined in the 293-HEK co-transfection assay. Four different siRNAs specific for the polynucleotide sequence (SEQ ID NO:_____) encoding SHP-2 (SEQ ID NO:_____) were co-transfected with a FLAG®-SHP-2 expression construct prepared according to the molecular biology methods described above. SHP-2 specific siRNAs had the following sequences.

SHP2.1: 5'-gauucagaacacuggugautt-3'	(SEQ ID NO:
SHP2.2: 5'-gaauauggcgucaugcgugtt-3'	(SEQ ID NO:
SHP2.3: 5'-cggucuggcaauaccacuutt-3'	(SEQ ID NO:
SHP2.4: 5'-ugacggcaagucuaaagugtt-3'	(SEQ ID NO:

[0186] The siRNA SHP2.1 effectively impaired expression of SHP-2 in transfected 293-HEK cells, decreasing the amount of FLAG®-SHP-2 polypeptide detected by more than 95%. In the presence of siRNA SHP2.2, FLAG®-SHP-2 polypeptide expression decreased by 85%. SHP2-4 had no specific effect on SHP-2 expression.

[0187] PRL-3 and KAP

[0188] Inhibition of expression of the human protein tyrosine phosphatases (PTP) PRL-3 and KAP were also examined in the 293-HEK co-transfection assay. Four different siRNAs specific for the polynucleotide sequence (SEQ ID NO:) encoding PRL-3 (SEQ ID) were co-transfected with a FLAG®-PRL-3 expression construct prepared according to the molecular biology methods described above. Similarly, four different siRNAs specific for the polynucleotide sequence (SEQ ID) encoding KAP (SEQ ID NO: co-transfected with a FLAG®-KAP expression construct. The siRNA sequences and the percent decrease in the level of expression of the PTP in cells transfected with the each siRNA is presented in Table 1 below, and it is noted that each 21-mer sequence below contains a dinucleotide "overhang" at the 3' end, and that the invention herein should be considered to include the 19-mer polynucleotide sequences beginning at the 5' end therein as well as the 21-mer polynucleotide shown in the Table.

TABLE 1

		siRNA				E WITH E		 KAP	
Target	: siRNA Se	quence	(SEQ	ID	NO)	siRNA	Name	 lated ID NO	Decrease in Expression
KAP KAP KAP	5 '-GAGCC 5 '-GAGCU 5 '-GAGCU	GUGGUAU	ACAAG	ACT	'T-3'	KAP KAP KAP	.2		>90% >90% >90%

TABLE 1-continued

sirna interference with prL-3 and kap in co-transfection assays

Target siRNA Sequence (SEQ ID NO)	siRNA Name	Decrease in Expression
KAP 5'-UACACUGCUAUGGAGGACUTT-3' PRL-3 5'-GUGACCUAUGACAAAACGCTT-3' PRL-3 5'-GGCCAAGUUCUGUGAGGCCTT-3' PRL-3 5'-GUACGAGGACGCCAUCCAGTT-3' PRI-3 UACCGGCCCAAACAGAGGCTT	KAP.4 Pr13.1 Pr13.2 Pr13.3 Pr13.4	<10% 50% 50% 50% <10%

[0189] PTP€

[0190] Inhibition of expression of human PTP€ is examined in the 293-HEK co-transfection assay. Four different siRNAs specific for the polynucleotide sequence (SEQ ID NO:_____) encoding PTP€ (SEQ ID NO:_____) are co-transfected with a FLAG®-PTP€ expression construct prepared according to the molecular biology methods described above. The siRNA sequences that are analyzed have AA leader sequences (not included in the siRNA polynucleotide transfected into HEK cells) and the following sequences.

RPTPE.1:5'GCAGAGGAAAGCUGUGGUCTT3' (SEQ ID NO:___)

RPTPE.2:5'GUCUGCGACCAUCGUCAUGTT3' (SEQ ID NO:___)

RPTPE.3:5'GCCUUACUCGAGUACUACCTT3' (SEQ ID NO:___)

RPTPE.4:5'GGACUAUUUCAUCGCCACCTT3' (SEQ ID NO:___)

[0191] Interference by siRNA Polynucleotides of Endogenous PTP Expression

[0192] The effect of sequence specific siRNA polynucleotides on expression of protein tyrosine phosphatases endogenously expressed in cells was also determined. Inhibition of expression of SHP-2 in HeLa cells by specific siRNAs was examined. HeLa cells were transfected with 10 nM of SEQ ID NO:_____); SHP2.2 (SEQ ID _____); DSP13.3 (SEQ ID NO:______); DSP14.1 SHP2.1 (SEQ ID NO: (SEQ ID NO:); and DSP14.3 (SEQ ID NO: Each siRNA was diluted in 50 µl OptiMEM® to provide a final concentration of 10 nM per well of cells in six well tissue culture plate. In a separate tube, 3 μ l of LipofectamineTM was combined with 10 μ l OptiMEM®. Each solution was incubated for 7 minutes. The two solutions were then mixed and incubated at room temperature for 22 minutes. The final volume of the mixed solution was adjusted to 500 μ l and then was added to the HeLa cells. Cells were transfected with the siRNAs or with annealing buffer alone. The transfected cells were incubated with siRNAs for 60 hours.

[0193] Cell lysates were prepared by extracting the cells in RIPA buffer as described in Example 1. The lysates were separated by SDS-PAGE gel and analyzed by immunoblot according to the procedures described in Examples 1 and above in Example 2 using an anti-SHP-2 murine monoclonal antibody (Santa Cruz Biotechnology, Inc., Santa Cruz, Calif.). The levels of expression of endogenous SHP-2 decreased by 75% in the presence of SHP2.2 and by 90% in

the presence of SHP2.1. The expression of SHP-2 in the siRNAs presence of DSP13.3, DSP14.1, or DSP14.3 was comparable to the level of expression observed in cells treated with buffer only.

[0194] A similar experiment was performed to determine the level of endogenous expression of DSP-3 in HeLa cells and in MDA-MB-435 cells (ATCC) in the presence of sequence specific siRNA. DSP3.1 siRNA (SEQ ID NO:1) was transfected into each cell line as described above, and the level of expression of DSP-3 polypeptide was analyzed by immunoblot (see Example 1 for immunoblot procedure to detect DSP-3). Expression of DSP-3 polypeptide decreased 70-100% in HeLa cells and decreased 100% in MDA-MB-435 cells in the presence of the specific mRNA.

[0195] Particular siRNA polynucleotide sequences that are specific for CD45, SHP2, cdc14a, cdc14b, cdc25A, cdc25B, cdc25C, PRL-3, KAP, DSP-3, and PTP ϵ are provided below. The level of expression of each PTP and DSP in cells that are capable of expressing the PTP or DSP and that are transfected with any one of the following specific siRNA polynucleotides is determined according to methods and procedures described above. The siRNA sequences that are incorporated into a vector from which a hairpin vector is transcribed and/or that are transfected via liposomes according to methods described in Examples 1 and 2 are presented in the following tables. The human TCPTP target sequences were derived from a human TCPTP nucleotide sequence (see GenBank Accession No. M25393, NM 002828, NM_080422 (SEQ ID NOs:)); the CD45 target sequences were derived from a human CD45 nucleotide sequence, (see Charbonneau et al. (SEQ ID NO: the SHP-2 target sequences were derived from a human SHP-2 nucleotide sequence (see GenBank Accession No. L03535 and L07527 (SEQ ID NO:)); the cdc14a target sequences were derived from a human cdc14a nucleotide sequence (see GenBank Accession No. AF122013 _)); the cdc14b target sequences were (SEO ID NO: derived from a human cdc14b nucleotide sequence (Gen-Bank Accession No. AF023158 (SEQ ID NO:_ cdc25A target sequences were derived from a human cdc25A nucleotide sequence (see GenBank Accession No. NM_133571 and AF527417 (SEQ ID NO: cdc25B target sequences were derived from a human cdc25B nucleotide sequence (see GenBank Accession No. M81934 (SEQ ID NO:____ __)); the cdc25C target sequences were derived from a human cdc25C nucleotide sequence (see GenBank Accession No. NM_001790 (SEQ ID NO: _); the PRL-3 target sequences are derived from the human PRL-3 nucleotide sequence (see GenBank Accession No. NM_032611 and NM_003479 (SEQ ID

NO:______); the KAP target sequences are derived from the human KAP nucleotide sequence (see GenBank Accession No. L2711 (SEQ ID NO:_____)); the DSP-3 target sequences were derived from the human DSP-3 nucleotide sequence set forth in (SEQ ID NO:778); and the PTPε target sequences were derived from the human PTPε nucleotide sequence (see GenBank Accession No. NM_006504 and NM_130435 (SEQ ID NO:_____)).

[0196] siRNA polynucleotide sequences were selected using the Dharmacon siDESIGN system (Dharmacon Research). These sequences were generated using the following parameters: (1) leader sequences included dinucleotides AA, CA, TA, and GA; (2) the coding region (CR) was scanned; (4) the G+C content varied from approximately 31-63%; (5) overlaps of sequences within different 19 nucleotide sequences were permitted. These sequences were then compared to known human genome sequences using the BLAST program. Potential target sequences were eliminated if 16 or more consecutive nucleotides within the 19-nucleotide target sequence were identified in another human polynucleotide sequence. The remaining 19-nucleotide siRNA sequences are presented in the tables below. Each siRNA sequence represented in Tables 2-12 lists the sequence of the sense strand of the siRNA and its corresponding sequence identifier. For PRL-3, only one sequence (AGACCCGGUGCUGCGUUAU, SEQ ID NO: was identified by this method. An siRNA polynucleotide as described herein is understood to be composed of the 19 nucleotide sense strand and its complementary (or antisense) strand. In addition, a siRNA polynucleotide of the present invention typically has a dinucleotide overhang at the 3' end of each strand, which may be any two nucleotides. Accordingly, it is noted that each 21-mer sequence below contains a dinucleotide "overhang" at the 3' end, and that the invention herein should be considered to include the 19-mer polynucleotide sequences beginning at the 5' end therein as well as the 21-mer polynucleotide shown in the Tables.

TABLE 2

HUMAN CD45 SIRNA POLYNCULEOTIDE SEQUENCES

(POST-BLAST)	~		_
19-Nucleotide Target Sequence	Region	SEQ	ID	NO.
CCACCAUCACAGCGAACAC	CR			
AGCGCUGUCAUUUCAACCA	CR			
ACCACAACAAUAGCUACUA	CR			
GCUACUACUCCAUCUAAGC	CR			
AAUGCGUCUGUUUCCAUAU	CR			
AUGCGUCUGUUUCCAUAUC	CR			
UGCGUCUGUUUCCAUAUCU	CR			
ACCUUUACUUGUGAUACAC	CR			
CAGAUUUCAGUGUGGUAAU	CR			
ACCCGAACAUGAGUAUAAG	CR			
CCCGAACAUGAGUAUAAGU	CR			
CAAGUUUACUAACGCAAGU	CR			
GGAGUAAUUACCUGGAAUC	CR			
CAUGCCUACAUCAUUGCAA	CR			
AUAGUAUGCAUGUCAAGUG	CR			
UGAACGUUACCAUUUGGAA	CR			
AUGAGUCGCAUAAGAAUUG	CR			
UGAGUCGCAUAAGAAUUGC	CR			
GAAUUGCGAUUUCCGUGUA	CR			
AUUGCGAUUUCCGUGUAAA	CR			
GCCAAUCCAUGCAGAUAUU	CR			
UUAUAACCGUGUUGAACUC	CR			
UAACCGUGUUGAACUCUCU	CR			
ACGGAGAUGCAGGGUCAAA	CR			

TABLE 2-continued

19-Nucleotide Target Sequence Region SEQ II	NO.
GAUGCAGGGUCAAACUACA CR	
ACCCAGGAAAUACAUUGCU CR	
UGUCCAGAUUACAUCAUUC CR	
AUGCCUUCAGCAAUUUCUU CR	
CAGGAACCUAUAUCGGAAU CR	
GGAACCUAUAUCGGAAUUG CR	
ACCUAUAUCGGAAUUGAUG CR	
GUGGAUGUUUAUGGUUAUG CR	
GGCGACAGAGAUGCCUGAU CR	
GAGGCCCAGUACAUCUUGA CR	
GGCCCAGUACAUCUUGAUC CR	
GCUACUGGAAACCUGAAGU CR	
ACCUGAAGUGAUUGCU CR	
AGUUGACCUGAAAGACACA CR	
ACUUAUACCCUUCGUGUCU CR	
CUUAUACCCUUCGUGUCUU CR	
GGAAAGACUCUCGAACUGU CR	
ACCCAAGGAAUUAAUCUCU CR	
CCCAAGGAAUUAAUCUCUA CR	
UGAUUCAGGUCGUCAAACA CR	
GGGAUGGAUCUCAGCAAAC CR	
UCUCAGCAAACGGGAAUAU CR	
UUCGAGCAAUAUCAAUUCC CR	
CCUACCCUGCUCAGAAUGG CR	

[0197]

TABLE 3

HUMAN SHP-2 siRNA POLYNCULI (POST-BLAST		EQUEN	CES	.
19-Nucleotide Target Sequence	Region	SEQ	ID	NO.
AUGGAGCUGUCACCCACAU	CR			
UGGAACAUCACGGGCAAUU	CR			
GCAAUGACGGCAAGUCUAA	CR			
AUGACGGCAAGUCUAAAGU	CR			
UGACGGCAAGUCUAAAGUG	CR			
GUCUAAAGUGACCCAUGUU	CR			
UGAUUCGCUGUCAGGAACU	CR			
CGACGUUGGUGGAGGAGAA	CR			
ACGGUUUGAUUCUUUGACA	CR			
UUCUUUGACAGAUCUUGUG	CR			
GAAUCCUAUGGUGGAAACA	CR			
AUCCUAUGGUGGAAACAUU	CR			
UCCUAUGGUGGAAACAUUG	CR			
CAGUACUACAACUCAAGCA	CR			
UUUGAGACACUACAACAAC	CR			
AACUUCUCUACAGCCGAAA	CR			
ACAUCCUGCCCUUUGAUCA	CR			
UCAUACCAGGGUUGUCCUA	CR			
UACCAGGGUUGUCCUACAC	CR			
UUUGAAACCAAGUGCAACA	CR			
AGAGUUACAUUGCCACACA	CR			
GAGUUACAUUGCCACACAA	CR			
AAACACGGUGAAUGACUUU	CR			
CUGGCCUGAUGAGUAUGCU	CR			
UGGCGUCAUGCGUGUUAGG	CR			
UGCGUGUUAGGAACGUCAA	CR			
UGACUAUACGCUAAGAGAA	CR			
CUAUACGCUAAGAGAACUU	CR			
GGUUGGACAAGGGAAUACG	CR			
GAACGGUCUGGCAAUACCA	CR			
CGGUCUGGCAAUACCACUU	CR			
AAGGUGUUGACUGCGAUAU	CR			
AGGUGUUGACUGCGAUAUU	CR			
GGUGUUGACUGCGAUAUUG	CR			

31

TABLE 3-continued

TIBEE 9-cence	inaca			
HUMAN SHP-2 siRNA POLYNCULI (POST-BLAST		EQUEN	ICES	<u>. </u>
19-Nucleotide Target Sequence	Region	SEQ	ID	NO.
UAUGGCGGUCCAGCAUUAU	CR			
UGGCGGUCCAGCAUUAUAU	CR			
AACACUACAGCGCAGGAUU	CR			
ACACUACAGCGCAGGAUUG	CR			
GCGCAGGAUUGAAGAAGAG	CR			
GAGGAAAGGGCACGAAUAU	CR			
GGAAAGGGCACGAAUAUAC	CR			
GGGCACGAAUAUACAAAUA	CR			
AAACGUGGGCCUGAUGCAA	CR			
ACGUGGGCCUGAUGCAACA	CR			

[0198]

TABLE 4

HUMAN CDC14A siRNA POLYNCUL (POST-BLAST		EQUE	NCE	s
19-Nucleotide Target Sequence	Region	SEQ	ID	NO.
GCACAGUAAAUACCCACUA	CR			
CUAUUUCUCCAUCGAUGAG	CR			
ACUUGGCAAUGGUGUACAG	CR			
GGUGCCUAUGCAGUAAUCU	CR			
UCUCACCAUUCUCGACUGU	CR			
AAGGGAUUACAACAUGGAU	CR			
AGGGAUUACAACAUGGAUU	CR			
GGGAUUACAACAUGGAUUU	CR			
GAAUGGUUAUCCUCUUCAC	CR			
GCAUAAUGUGACUGCAGUU	CR			
CGCUGGCUUCGAGCACUAU	CR			
GCACACCCAGUGACAACAU	CR			
ACAUCGUGCGAAGGUUCCU	CR			
AGAACAGGGACAUUGAUAG	CR			
GAACAGGGACAUUGAUAGC	CR			
GGGACAUUGAUAGCCUGUU	CR			
CAUUGAUAGCCUGUUAUGU	CR			
CUACAGGUUUACACAUGCU	CR			
AAAUCGACCAUCCAGUGAA	CR			
AAUCGACCAUCCAGUGAAG	CR			
UCGACCAUCCAGUGAAGGA	CR			
AAAUUCUUUCUGGCCUAGA	CR			
UGUCUAUUGGUGGAAAUCU	CR			
ACGAUUUGGAGAGGUAAGU	CR			
CGAUUUGGAGAGGUAAGUU	CR			

[0199]

TABLE 5

HUMAN CDC14B SIRNA POLYNCULEOTIDE SEQUENCES (POST-BLAST)						
19-Nucteotide Target Sequence	Region	SEQ ID NO.				
GAGACAUCCUAUAUUCCUU	CR					
AUACCAGACCGAUUUAUUG	CR					
UACCAGACCGAUUUAUUGC	CR					
GACCGAUUUAUUGCCUUCU	CR					
AAGGAUGUAUGAUGCCAAA	CR					
AGGAUGUAUGAUGCCAAAC	CR					
GGAUGUAUGAUGCCAAACG	CR					
CGGAUGCUGGCUUCGAUCA	CR					
UGCCAUUGUCAAAGAAUUC	CR					
GGGUGCCAUUGCAGUACAU	CR					
GACCUGGCUCGGUGAUUGG	CR					

TABLE 5-continued

HUMAN CDC14B siRNA POLYNCULEOTIDE SEQUENCES (POST-BLAST)					
19-Nucteotide Target Sequence	Region	SEQ ID NO.			
CCCGAACCGUACAGUGAUG	CR				
ACCGUACAGUGAUGAC	CR				
UAGACUUCGGGCCUUGAAA	CR				
ACAAACGCUAUUCCUCUCA	CR				

[0200]

TABLE 6

HUMAN CDC25A siRNA POLYNCUI (POST-BLAST		SEQUENCE	es —
19-Nucleotide Target Sequence	Region	SEQ ID	NO.
GGGUCUGGGCAGUGAUUAU	CR		
GCAACCACUGGAGGUGAAG	CR		
AUCCUAUGAGAAGAAUACA	CR		
UCCUAUGAGAAGAAUACAU	CR		
AAAGCUGUUGGGAUGUAGU	CR		
UUCUGAUUCUCUUGACCAU	CR		
GAAGCCAGUAAGACCUGUA	CR		
CAGCCACUUUGUCUGAUGA	CR		
AACCUUGACAACCGAUGCA	CR		
CAACCGAUGCAAGCUGUUU	CR		
ACCGAUGCAAGCUGUUUGA	CR		
CUCGGUCAGUGUUGAAGAG	CR		
ACGUUCUCAAGAGGAGUCU	CR		
GUCAACUAAUCCAGAGAAG	CR		
AGGCCCAUGAGACUCUUCA	CR		
AGGGACCUUAUAGGAGACU	CR		
GGGACCUUAUAGGAGACUU	CR		
GACUUCUCCAAGGGUUAUC	CR		
GUUUGUUAUCAUCGACUGU	CR		
CUGUCGAUACCCAUAUGAA	CR		
GAAGCCCAUUGUACCUACU	CR		
AGCCCAUUGUACCUACUGA	CR		
GCCCAUUGUACCUACUGAU	CR		
UGGCAAGCGUGUCAUUGUU	CR		
AGCGUGUCAUUGUUGUUU	CR		
UGUGCCGGUAUGUGAGAGA	CR		
GAGAGAUCGCCUGGGUAAU	CR		
GAGAUCGCCUGGGUAAUGA	CR		
GAUCGCCUGGGUAAUGAAU	CR		

[0201]

TABLE 7		
HUMAN CDC25B siRNA POLYNCUL (POST-BLAST		SEQUENCES
19-Nucleotide Target Sequence	Region	SEQ ID NO.
AUCCUCCCUGUCGUCUGAA	CR	
UCCUCCCUGUCGUCUGAAU	CR	
UGGCGGAGCAGACGUUUGA	CR	
CGUUUGAACAGGCCAUCCA	CR	
GCCGGAUCAUUCGAAACGA	CR	
UCAUUCGAAACGAGCAGUU	CR	
GUCUAUGCCGGAUGGAUUU	CR	
UGCCGGAUGGAUUUGUCUU	CR	
AAAGGACCUCGUCAUGUAC	CR	
AAUCACUGUGUCACGAUGA	CR	
AUCACUGUGUCACGAUGAG	CR	
GAGCUGAUUGGAGAUUACU	CR	
GCUGAUUGGAGAUUACUCU	CR	

TABLE 7-continued

HUMAN CDC25B siRNA POLYNCUL (POST-BLAST		EQUENCES
19-Nucleotide Target Sequence	Region	SEQ ID NO.
CUCUAAGGCCUUCCUCCUA	CR	
CAGACAGUAGACGGAAAGC	CR	
AGCACCAAGACCUCAAGUA	CR	
GAAACGAUGGUGGCCCUAU	CR	
AACGAUGGUGGCCCUAUUG	CR	
CGCCGAGAGCUUCCUACUG	CR	

[0202]

TABLE 8

HUMAN CDC25C siRNA POLYNCUL (POST-BLAST		EQUEN	CES
19-Nucleotide Target Sequence	Region	SEQ :	ID NO.
GAACUCCAGUGGGCAAAUU	CR		
UUUAGCUGGGAUGACAAUG	CR		
UUCAAGGACAACACAAUAC	CR		
ACACAAUACCAGAUAAAGU	CR		
CACAAUACCAGAUAAAGUU	CR		
GGAAGGGCUUAUGUUUAAA	CR		
CACCAAGAUCUGAAGUAUG	CR		
AGUAUGUCAACCCAGAAAC	CR		
GUAUGUCAACCCAGAAACA	CR		
UGUCAUUGAUUGUCGCUAU	CR		
UUGAUUGUCGCUAUCCAUA	CR		
UUGUCGCUAUCCAUAUGAG	CR		
UCCAGGGAGCCUUAAACUU	CR		
GGGAGCCUUAAACUUAUAU	CR		
GUCAGGAAGAACUGUUUAA	CR		
AGAAGCCCAUCGUCCCUUU	CR		
GAAGCCCAUCGUCCCUUUG	CR		
AGCCCAUCGUCCCUUUGGA	CR		
CACCCAGAAGAAUAAUC	CR		
UUGUACUACCCAGAGCUAU	CR		
CUACCCAGAGCUAUAUAUC	CR		
CCCAGAGCUAUAUAUCCUU	CR		
UAUAUGGAACUGUGUGAAC	CR		
UAUGGAACUGUGUGAACCA	CR		
CAGAGCUACUGCCCUAUGC	CR		
GAGCUACUGCCCUAUGCAU	CR		
GCUACUGCCCUAUGCAUCA	CR		

[0203]

TABLE 9

HUMAN KAP siRNA POLYNCULE (POST-BLAST		QUENCES
19-Nucleotide Target Sequence	Region	SEQ ID NO.
GAUGAAGAGCCUAUUGAAG	CR	
AGAUGAACAGACUCCAAUU	CR	
GAUGAACAGACUCCAAUUC	CR	
UCACCCAUCAUCAUCCAAU	CR	
GAGCUUACAACCUGCCUUA	CR	
CACUGCUAUGGAGGACUUG	CR	
UCACCAGAGCAAGCCAUAG	CR	
CCAGAGCAAGCCAUAGACA	CR	
CAGCCUGCGAGACCUAAGA	CR	
GUUUCGGGACAAAUUAGCU	CR	
AAUUAGCUGCACAUCUAUC	CR	

TABLE 9-continued

	HUMAN KAP		POLYNCULE OST-BLAST		QUENCES	
19-N	Nucleotide	Target	Sequence	Region	SEQ ID	NO.
	AUUAGCUG	CACAUCU	AUCA	CR		
	UUAGCUGC	ACAUCUA	UCAU	CR		

[0204]

TABLE 10

HUMAN DSP-3 siRNA POLYNCULEOTIDE SEQUENCES (POST-BLAST)			
19-Nucleotide Target Sequence	Region	SEQ ID NO.	
GAGACGCGGAACAAUUGAG	CR		
AGAACAAGGUGACACAUAU	CR		
GAACAAGGUGACACAUAUU	CR		
GCAGCGGAUUCACCAUCUC	CR		
GCGGAUUCACCAUCUCAAA	CR		
CACUGGUGAUCGCAUACAU	CR		
GUAUCGGCAGUGGCUGAAG	CR		

[0205]

TABLE 11

HUMAN PTP EPSILON SIRNA POLYN (POST-BLAST		E SEÇ	UEN	ICES
19-Nucleotide Target Sequence	Region	SEQ	ID	NO.
GAUCCGCCGACGACUGCAA	CR			
GUUUCGGGAGGAGUUCAAC	CR			
AUGACCAUUCUAGGGUGAU	CR			
CCAUUCUAGGGUGAUUCUG	CR			
CAUAGAUGGUUACAAAGAG	CR			
AACAGGAAACGGUUAACGA	CR			
GGAAACGGUUAACGACUUC	CR			
CCAUCGUCAUGUUAACAAA	CR			
CUACACCAUCCGGAAGUUC	CR			
UCCGGAAGUUCUGCAUACA	CR			
GAAAGUAAAGACGCUCAAC	CR			
GCGCCCUCAGAUGGUUCAA	CR			
CGGAUAUGCAGUACACGUU	CR			
CCACCCACUUCGACAAGAU	CR			
CAAAUGUCCGGAUCAUGAA	CR			
CAUGAGGACGGCAACUUG	CR			
UGACUUCAACCGAGUGAUC	CR			
ACCGAGUGAUCCUUUCCAU	CR			
AGAAUACACAGACUACAUC	CR			
GACUACAUCAACGCAUCCU	CR			
UCAACGCAUCCUUCAUAGA	CR			
CACACGGUUGAGGACUUCU	CR			
AAUCCCACACUAUCGUGAU	CR			
AUCCCACACUAUCGUGAUG	CR			
ACCGAGGGCUCAGUUACUC	CR			
CCGAGGGCUCAGUUACUCA	CR			
CUCAUGGAGAAAUAACGAU	CR			
UGGAGAAAUAACGAUUGAG	CR			
GCCAUCAGUAUACGAGACU	CR			
UCAGUAUACGAGACUUUCU	CR			
GGGCAAAGGCAUGAUUGAC	CR			
GCUGGGCGAACAGGUACAU	CR			
CUUCAGAGACCACAUAUGG	CR			

EXAMPLE 3

Decreased Activation of JNK in the Presence of siRNA Specific for DSP-3

[0206] This Example describes the effect on JNK activation by sequence-specific siRNA interference of DSP-3 polypeptide expression.

[0207] HeLa cells were transfected with 60 pmoles of DSP3.1 siRNA (SEQ ID NO:1) or 60 pmoles CD45.2 (SEQ ID NO:13) as described in Example 1. After the incubation following transfection, cells were stimulated with 10 ng/ml TNF-α or 10 ng/ml EGF for 10 minutes or with 500 mM sorbitol for 30 minutes, which are known stimulators of the JNK signal transduction pathway (WO 01/21812; Shen et al. Proc. Natl. Acad. Sci. 98:13613-18 (2001)). After the stimulators were decanted, the 6-well plate of cells was frozen. The cells were treated with 0.5 ml Extraction Buffer (20 mM Tris, pH 8, 136 mM NaCl, 50 mM NaF; 1 mM V04; 0.2 mM EDTA, 0.2 mM EGTA, 20 nM Calyculin, 10% glycerol, 0.5% nonidet P40, 1 μ g/ml of aprotinin, pepstatin, and leupeptin; and 1 mM Benzamidine) (4° C.). When the cells had partially thawed, the wells of the plates were scraped and the cells were collected. The wells were washed 3x with Extraction Buffer and the washes were combined with the cells. After centrifugation of the extracted cells, the supernatants were decanted. The protein concentration of each extract was determined by the Bradford protein assay. Volumes of the different extracts were adjusted with Extraction Buffer to the concentration of the extract having the lowest protein concentration.

[0208] JUN, a substrate of JNK, conjugated to glutathione (GSH) (GST-cJUN) (Shen et al., supra) in 20 mM Tris, pH 7.2, 1 nM EDTA, and 150 mM NaCl was combined with 200-250 μ l of Glutathione-Sepharose (Amersham Biosciences, Piscataway, N.J.). After mixing for 45 minutes at 4° C., the conjugated sepharose beads were washed twice in Extraction Buffer and then resuspended in 1 ml of Extraction Buffer.

[0209] cJUN-Sepharose (20 μ l) was added to each cell extract sample. The mixtures were gently mixed for 2 hours at 4° C., followed by one wash in 1 ml Extraction Buffer and once in 1 ml kinase buffer (20 mM Pipes, pH 7.2, 10 mM MgCl₂, 1 mM DTT, 0.1% Triton X-100, and 1 MM sodium vanadate). The mixtures were centrifuged and the pellets were kept on ice. ATP mix (300° C./ml of $[\gamma^{-32}P]$ ATP (3000 Ci/mmole) in kinase buffer) was incubated in a heat block to bring the solution to 30° C. ATP mix (15 µl) was added to each cold cJUN-Sepharose pellet at time intervals of 20 seconds. After the ATP mix was added, each sample was vortexed gently for 5 seconds and then placed in the 30° C. heat block. Each sample was gently mixed again for 5 seconds at 20-second intervals. After 20 minutes, the reactions were stopped at 20-second intervals with 15 µl 2×SDS-PAGE sample buffer. The samples were immediately heated at 100° C. for 5 minutes, then mixed and frozen at -20° C. The extracts were thawed and applied to 8-16% NOVEX® gels. After electrophoresis, the gels were dried and the cJUN band was cut from the gel and the radioactivity was counted (Cerenkov measurement). As shown in FIGS. 3 and 4, JNK activation as measured by the presence of phosphorylated JUN was mediated less by cells transfected with siRNA specific for DSP-3 than in cells transfected with a nonspecific siRNA.

[0210] Because EGF induces a signaling pathway involving the ERK MAP kinase family, the effect on ERK phosphorylation in HeLa cells transfected with DSP-3 specific siRNA was determined. Transfection of HeLa cells and stimulation of the JNK signaling pathway was performed as in the previous experiment. Additional transfected cell cultures were stimulated with anisomycin. Phosphorylation of ERK was determined in a similar manner as described above for cJUN except that after electrophoresis of the cell extract samples, the proteins separated in the gel were transferred to a PVDF membrane. The immunoblot was probed with an anti-phospho-ERK antibody (1:1000) followed by incubation with the appropriate HRP-conjugated reagent and detection by chemiluminescence. As shown in FIG. 5, phosphorylation of ERK induced by stimulation of the cells with EGF and sorbitol was not affected by interference of DSP-3 polypeptide expression by specific siRNA DSP3.1.

EXAMPLE 4

Interference of Expression and Function of Cell Division Cycle Proteins by Specific siRNA

[0211] This example describes interference of expression of cell division cycle (cdc) proteins, cdc14a, cdc14b, and cdc25A, cdc25B, and cdc25C polypeptides by sequence specific siRNA polynucleotides. The effect on the function of these polypeptides in the presence of siRNA was also determined.

[0212] Interference with Cell Division Cycle Protein Expression by Specific siRNA

[0213] Two siRNA sequences that were specific for cdc14a polynucleotide (SEQ ID NO:33) encoding a cdc14a polypeptide (SEQ ID NO: 34) and two siRNA sequences specific for cdc14b polynucleotide (SEQ ID NO:35) encoding a cdc14b polypeptide (SEQ ID NO:36) were designed using the criteria described in Example 1. Recombinant expression vectors containing polynucleotide sequences encoding FLAG®-tagged cdc14a polypeptide and FLAG®tagged cdc14b polypeptide were prepared essentially according to methods described in Example 2 with the following exceptions. 293-HEK cells were cultured in 35 mm culture dishes and were transfected with FLAG vectors at a concentration of 1 µg per well. 293-HEK cells were co-transfected with FLAG®-tagged cdc14a expression vector and the following siRNAs at 20 nM per well: cdc14a.2 (5'-caucugugagaacaccgaatt-3', SEQ ID NO: cdc14a.3 (5'-cuuggcaaugguguacagatt-3', SEQ), cdc14a.5 (5'-); cdc14a.4, SEQ ID NO: NO: gcacaguaaauacccacuatt-3', SEQ ID NO:); DSP3.1 (SEQ ID NO:); DSP3.2 (SEQ ID NO: cdc14b.3 (5'-caagcaaaugcugccuucctt-3', **SEQ** IDNO: _); cdc14b.4 (5'-gagccagacuugaaaguggtt-3', SEQ); and CD45.3); MKP.2 (SEQ ID NO: (negative control). Controls included 293-HEK cells that were not transfected with any vector or siRNA and 293-HEK cells transfected with FLAG®-tagged cdc14a in the presence of siRNA annealing buffer. The level of expression in each sample was analyzed by immunoblot as described in Example 2 using an anti-FLAG® antibody. As shown in FIG. 6, specific siRNAs, cdc14a.2, cdc14a.3, and cdc14a.5 interfered with expression of cdc14a polypeptide most effectively.

[0214] Specificity of cdc14a.3 siRNA for interfering with expression of cdc14a and not other dual specificity phos-

phatases was shown by co-transfecting cdc14a.3 siRNA with FLAG®-tagged cdc14a (1 µg per 35 mm well of cells), FLAG®-tagged DSP-3, FLAG®tagged cdc14b, and FLAG®-tagged DSP-11. A FLAG® recombinant expression construct containing a polynucleotide sequence (SEQ ID NO:_____) encoding a DSP-3 polypeptide (SEQ ID NO:_____) was prepared as described for constructing other FLAG vectors. 293-HEK cell transfections and analysis of polypeptide expression levels were performed as described in Example 2. FIG. 7 shows that siRNA cdc14a.3 interfered with expression of only the cdc14a dual specificity phosphatase.

[0215] 293-HEK cells were co-transfected with FLAG®tagged cdc14b expression vector (2 μ g/35 mm well) and the following siRNAs at 20 nM per well: cdc14b.3 (SEQ ID NO:); cdc14b.4 (SEQ ID NO:_ (SEQ ID NO: ____)); cdc14a.5 (SEQ ID NO: DSP3.1 (SEQ ID NO: ____); DSP3.2 (SEQ ID NO: _____);); DSP3.2 (SEQ ID); MKP.2 (SEQ ID NO:); and CD45.3. Controls included 293-HEK cells that were not transfected with any vector or siRNA and 293-HEK cells transfected with FLAG®-tagged cdc14b in the presence of siRNA annealing buffer. The level of expression in each sample was analyzed by immunoblot as described in Example 2 using an anti-FLAG® antibody. As shown in FIG. 8, only specific siRNAs, cdc14b.3 and cdc14b.4 interfered with expression of cdc14b polypeptide.

[0216] Specificity of cdc14b.3 and cdc14b.4 siRNAs for interfering with expression of cdc14b and not other dual specificity phosphatases was shown by co-transfecting the siRNAs with FLAG®-tagged cdc14b (2 µg per 35 mm well), FLAG®-tagged DSP-3, and FLAG®-tagged DSP-11. Cells transfected with FLAG®-tagged DSP-3 and FLAG®-tagged DSP-11 were also co-transfected with cdc14a.5 siRNA. 293-HEK cell transfections and analysis of polypeptide expression levels were performed as described in Example 2. FIG. 9 shows that cdc14b.3 and cdc14b.4 siRNAs interfered with expression of only the cdc14b dual specificity phosphatase.

[0217] Expression of cdc14b polypeptide co-transfected with cdc14b.4 siRNA in HeLa cells was analyzed by immunocytochemistry. HeLa cells were co-transfected with a cdc14b recombinant expression vector and siRNA. Expression of cdc14b was detected by standard immunocytochemistry methods. As shown in FIG. 10, cdc14b.4 siRNA interfered with expression of cdc 14b polypeptide (top and bottom right panels).

[0218] The efficacy of RNAi against FLAG®-tagged Cdc25A expression in 293-HEK cells was also determined. Cells were co-transfected with a FLAG®-Cdc25A expression construct (prepared as described in Example 2) and specific siRNAs 25A.1, 25A.2, 25A.3, and 25A.4 (20 nM) and non-specific siRNAs (25B.1-0.4 and 25C.1-0.4). The level of expression of Cdc25A was determined by immunoblotting with an anti-FLAG® antibody. Only siRNA 25A.2 (5'-gaggagccauucugauucutt-3' (SEQ ID NO:_____)) effectively inhibited expression of Cdc25A.

[0219] The effect of RNAi on endogenous expression of Cdc25B and Cdc25C was examined in HeLa cells. The experiments were performed essentially as described in Example 2, except that HeLa cells were exposed to 10 nM siRNA polynucleotides for 48 hours. Four siRNAs specific

for Cdc25A: 25A.1, 25A.2, 25A.3, and 25A.4 (20 nM); four siRNAs specific for Cdc25B: 25B.1, 25B.2, 25B.3, and 25B.4 (20 nM); and four siRNAs specific for Cdc25C: 25C.1, 25C.2, 25C.3, and 25C.4 (20 nM) were transfected into HeLa cells and expression was analyzed by immunoblotting cell lysates separated by SDS-PAGE using a Cdc25B antibody (Santa Cruz Biotechnololgy, Cat. No. c-20) and a Cdc25C antibody (Santa Cruz Biotechnololgy, Cat. No. h-85). The level of expression of Cdc25B was decreased 40-50% in HeLa cells transfected with siRNA cdc25B.2 (5'-aggcggcuacaaggaguuctt-3')), and 50-60% in cells transfected with NO: cdc25B.4 siRNA 5'-gaugecauggaageceacatt-3' (SEQ ID)). In HeLa cells transfected with siRNAs spe-NO: cific for Cdc25C, the level of expression of Cdc25C decreased 90% in HeLa cells transfected with cdc25C.1 (5'-cugceacucageuuaceactt-3' (SEQ ID NO: decreased 70-80% in cells transfected with cdc25C.3 (5'cccagaaacaguggcugcctt-3' (SEQ ID NO: decreased 70-80% in cells transfected with Cdc25C.4 (5'aggcggcuacagagacuuctt-3' (SEQ ID NO:_

[0220] The ability of cancer cell lines to mediate RNA interference was examined by co-transfecting several cancer cell lines with a FLAG® cdc14b expression construct and specific siRNAs. The cell lines included SW620 (colon cancer); MCF7 (breast cancer); HS578T (breast cancer); MDA MB 231 (breast cancer); and T47D (breast cancer) (ATCC, NCI 60 panel). The FLAG® cdc14b expression construct (1-2 µg) was co-transfected with 20 nM of 14b.3 siRNA (SEQ ID NO:); 14b.4 siRNA (SEQ ID); or MKP.2 siRNA (SEQ ID NO:) (nonspecific control) into each cell line as described in Example 2. The level of expression was analyzed by immunoblotting with an anti-FLAG® antibody according to the method described in Example 2. Expression of cdc14b was decreased in each of the five cell lines that were cotransfected with a cdc14b specific siRNA polynucleotide.

[0221] Effect of CDC-Specific siRNA on Cell Prolifera-

[0222] Proliferation of cancer cells in the presence of siRNA polynucleotides specific for cdc14a, cdc14b, and Cdc25A, Cdc25B, and Cdc25C was determined. Cell proliferation was assessed according to a quantitative metabolic assay that measures the enzymatic conversion by cellular dehydrogenase in viable cells of a yellow tetrazolium salt (methylthiazoletetrazolium (MTT)) to purple formazan crystals. MDA-MB-231, SW620, and HeLa cell lines were transfected according to the procedures described in Examples 1 and 2 with the following siRNA polynucleotides (5 nM): cdc14a.3 (5'-cuuggcaaugguguacagatt-3' (SEQ ID)); cdc14a.5 (5'-gcacaguaaauacccacuatt-3' (SEQ ID $\overline{\text{NO}}$: _)); cdc14b.3 (5'-caagcaaaugcugccuucctt-3' SEQ ID \overline{NO} :); cdc14b.4 (5'-gagccagacuugaaaguggtt-); cdc25A.2 (SEQ ID NO: 3' SEQ ID NO: __); cdc25C.1 (SEQ ID cdc25B.4 (SEQ ID NO:). The transfected cells were seeded at in a tissue culture plate and maintained for 5 days. A MTT assay was performed according to manufacturer's instructions (ATCC MTT Cell Proliferation Assav Kit, Cat. NO. 30-1010K, ATCC). The MTT-containing media was removed from the wells and was added to solubilize the formazan. The amount of formazan formed was determined by measuring absorbance at 570 m. Compared to the buffer only control, a

significant decrease in proliferation was observed in MDA-MB-231 cells transfected with cdc14a.3, cdc14a.5, cdc14b.3, cdc14b.4, and cdc25B.4, and in HeLa cells transfected with cdc14a.3, cdc14a.5, cdc14b.4, and cdc25B.4. A significant decrease in cell proliferation of SW620 cells transfected with cdc14a.3 or cdc14a.5 was also observed.

[0223] Effect of CDC-Specific siRNA on Proapoptotic Signaling

[0224] Poly(ADP-ribose) polymerase (PARP) is a nuclear DNA binding protein that participates in genome repair, DNA replication, and the regulation of transcription. Cleavage of PARP (approximately 115 kDa) by members of the caspase family into polypeptide fragments of approximately 85 kDa and 25 kDa prevents PARP from performing its normal repair functions and appears to be an early event in apoptotic cell death. The cleaved PARP fragments can be detected by a variety of immunodetection methods.

[0225] HeLa cells were transfected with cdc14a.5 (SEO _); cdc14b.4 (SEQ ID NO:_ _); cdc25A.2 ID NO: (SEQ ID NO: ____); cdc25B.4 (SEQ ID NO:____ __); and cdc25C.1 (SEQ ID NO:_____) at a concentration of 10 nM. After incubating the transfected cells for at 37° C., cell lysates were prepared and an immunoblot performed an antibody that that specifically binds to cleaved PARP and an antibody that binds to PARP (Cell Signaling Technology, Beverly, Mass.). The results are presented in FIG. 24. The data indicated that inhibiting expression of cdc14a by specific siRNA induces proapoptotic signaling to a greater extent than inhibition of expression of the other cell division cycle proteins.

EXAMPLE 5

Interference of PTP-1B and TC-PTP Expression by Specific siRNA

[0226] This Example describes interference with expression of two protein tyrosine phosphatases, PTP-1B and TC-PTP, using sequence specific siRNA polynucleotides.

[0227] Interference of Endogenous Expression of Murine PTP-1B in Mouse Fibroblasts by Sequence Specific siRNA Polynucleotides

[0228] Three siRNA sequences that were specific for murine PTP-1B polynucleotide (GenBank Acc. No. NM_011201, SEQ ID NO:) encoding a murine PTP-1B polypeptide (GenBank Acc. No. NM_011201, SEQ ID NO: and one siRNA sequences specific for human PTP-1B polynucleotide (GenBank Ace. No. NM_02827, SEQ ID NO:) encoding a human PTP-1B polypeptide (GenBank Ace. No. NM_02827, SEQ ID) were designed using the criteria described in Examples 1 and 2. Mouse C57B16 #3 cells, clones 3 and 10, were maintained in cell culture according to standard cell culture methods. Each C57B16 #3 clone was transfected with 200 nM of the following siRNAs: mPTP1B.1 (SEQ ID _), mPTP1B.2 (SEQ ID NO:_ _), mPTP1B.3 (SEQ ID NO:), and hPTP1B.1 (SEQ ID). Each siRNA was diluted in 50 μ l O_{PTI}MEM® to provide a final concentration of 200 nM per well. In a separate tube, 3 μ l of LipofectamineTM was combined with 10 μ l O_{PTI}MEM®. Each solution was incubated for 7 minutes. The two solutions were then mixed and incubated at room temperature for 22 minutes. The final volume of the mixed solution was adjusted to $100 \,\mu$ l and then the C57B16 #3 cells were added. Cells were transfected with the specific siRNAs, the human PTP1B siRNA, or annealing buffer alone. The transfected cells were incubated with siRNAs for six days.

[0229] Cell lysates were prepared by extracting the cells in ELISA extraction buffer (50 mM Tris-HCl, pH 7.5 (room temperature); 2 mM EDTA, pH 7-8; 1 mM phosphate (polyphosphate); 1 mM NaVO4 (monomeric), pH 10; 0.1% Triton X-100; Protease Inhibitor Cocktail set III, (Calbiochem, San Diego, Calif., catalog #539134)). The lysates were separated by SDS-PAGE gel and analyzed by immunoblot according to the procedures described in Examples 1 and 2 using an anti-PTP1B murine monoclonal antibody (Dr. Ben Neel, Harvard University, Cambridge, Mass.). As shown in FIG. 11, the levels of expression of endogenous PTP1B were decreased only in C57B16 cells transfected with the murine PTP1B sequence specific siRNAs.

[0230] The effect of RNAi on endogenous expression of murine PTP1B in a second murine cell line was examined. Mouse PTP1B:3T31R fibroblasts were transfected with 20 nM mPTP1B1.1 (SEQ ID NO:_____); mPTP1B1.6 (SEQ ID NO:_____); and mPTP1B1.8 (SEQ ID NO:_____) according to the method described above. The level of murine PTP1B expression in the cells transfected with mPTPB11.1 decreased approximately 80% compared with cells transfected with a non-specific siRNA (hPTP1B1.3 (SEQ ID NO:_____)); cells transfected with mPTP1B1.6 decreased approximately 40%; and cells transfected with mPTP1B1.8 decreased approximately 60%.

[0231] Interference with Murine PTP1B Expression by siRNA in Co-Transfection Assays

[0232] A recombinant expression construct was prepared that encodes wild-type murine PTP1B (mPTP1B) (GenBank Accession No. NM_011201, SEQ ID NOs:_____ and _____). The following oligonucleotide primers were used for the wild-type construct. The sequences of the BamHI and EcoRI restriction sites are underlined.

mPTP1B-sense (mPTP1B 5'BamHI)

(SEQ ID NO:___)
5'-GGGGGGGATCCATGGAGATGGAGAAGGAGTTCGAGG-3'

mPTP1B anti sense (mPTP1B 3'EcoRI)

(SEQ ID NO:___)
5'-GGGGGAATTCTCAGTGAAAACACACCCGGTAGCAC-3'

[0233] Vector pCMVTag2B (Stratagene, La Jolla, Calif.) was digested with restriction endonuclease BamHI (New England Biolabs, Beverly, Mass.) for 3 hours at 37° C. The digested vector was then incubated with Klenow polymerase (New England Biolabs) for 15 minutes at 25° C. to fill in the recessed 3' termini, followed by an incubation of 30 minutes at 37° C. with calf intestinal phosphatase (New England Biolabs). The GATEWAYTM Reading Frame Cassette B (Invitrogen Life Technologies, Carlsbad, Calif.) was inserted into the pCMVTag2B vector by ligation with T4 DNA ligase (Invitrogen Life Technologies) overnight at 16° C. according to the supplier's instructions. DB3.1TM competent *E. Coli* cells were transformed with the ligated vector (GWpCMVTag2) and DNA was isolated by standard molecular biology methods.

[0234] Vectors for expression of mPTP1 B wild type were prepared as follows. The mPTP1B construct was subcloned into a GATEWAYTM entry vector pENTR3 CTM (Invitrogen Life Technologies) by digesting $20 \mu l$ of the mPTP1B cDNA or 20 µl of the pENTR3CTM vector with 1 µl of BamHI (New England Biolabs); 1 μl of EcoRI (New England Biolabs); 5 μl 10×EcoRI buffer (New England Biolabs); 5 μl 10×BSA (New England Biolabs); and $18 \mu l$ distilled water for 3 hours at 37° C. Digested DNA was run on a 1% agarose gel, digested bands were excised, and the DNA was gel-purified using a QIAGEN Gel Extraction kit (QIAGEN, Inc., Valencia, Calif.). Four microliters of the mPTP1B cDNA was ligated into 2 μ l of the pENTR3CTM vector overnight at 16° C. with 1 μ l 10× Ligation Buffer (Invitrogen Life Technologies), 1 µl T4 DNA Ligase (4U/µl) (Invitrogen, Carlsbad, Calif.), and 2 μ l distilled water. The construct was transformed into LIBRARY EFFICIENCY® DH5α™ cells. The FLAG® epitope-tagged mPTP1B construct was prepared by cloning the pENTR3 CTM mPTP1B WT construct into the GWpCMVTag2 vector. The pENTR3C™ construct containing the mPTP1B polynucleotide was linearized by digesting the construct with Vsp I (Promega Corp., Madison, Wis.) at 37° C. for 2 hours. The DNA was purified using a QIAGEN PCR Purification kit (QIAGEN, Inc.). Three microliters (100 ng/µl) of the GWpCMVTag2 vector were combined in a GATEWAYTM LR reaction with 6 μl linearized pENTR3CTM mPTP1B WT, 3 μl TE buffer, 4 μl ClonaseTM Enzyme, and 4 μ l LR reaction buffer (Invitrogen Life Technologies) for 1 hour at room temperature. After addition of Proteinase K (Invitrogen Life Technologies) to the reaction for 10 minutes, LIBRARY EFFICIENCY® DH5a™ cells were transformed with the expression construct.

[0235] The murine PTP1B expression vector (0.5 µg) was co-transfected with 20 nM murine PTP1B sequence-specific siRNA polynucleotides into PTP1B knockout mouse fibroblasts (PTP1B KO mouse embryonic fibroblasts were prepared from 13-day embryos from PTP1B knock out mice to establish the cell line, which was then transfected with human insulin receptor (1BKO+HIR) (HIR, Julie Moyers, Eli Lilly and Company, Indianapolis, Ind.)). Transfections were performed as described in Example 1. After incubating the transfected cells for 18 hours at 37° C., cell lysates were prepared, separated by 4-12% SDS-PAGE, and immunoblotted using the anti-PTP1B murine monoclonal antibody (see above). The results are summarized in Table 13.

[0236] Interference with Rat PTP1B Expression by siRNA in Co-Transfection Assays

[0237] A co-transfection assay was performed as described above in which 1BKO+HIR mouse fibroblasts were co-transfected with an expression vector containing the sequence encoding the peptide FLAG® in frame with a nucleotide sequence (SEQ ID NO: __) that encoded a rat PTP1B polypeptide (SEQ ID NO:) (GenBank Accession No. NM_102637) and a sequence specific siRNA, rPTP1B1.1 (5'-agaagaaaaagagaugguctt-3' (SEQ ID NO:)) (20 nM). Additional rat PTP1B specific siRNA polynucleotides examined in the co-transfection assay included rPTP1B1.2 (5'-cggauggugguggagguctt-3' (SEQ ID NO:)); rPTP1B1.3 (5'-uggcaagugcaaggagcuett-3' (SEQ ID NO:)); and rPTP1B1.4 (5'cuacaccaccuggccugactt-3' (SEQ ID NO: of expression of the rat PTP1B polypeptide was determined by immunoblotting cell lysates with an anti-human PTP1B antibody that also specifically binds to rat PTP1B ((PHO2, Oncogene Research Products™, Inc. San Diego, Calif.). Expression of rat PTP1B decreased approximately 50% in cells transfected with rPTP1B1.1.

[0238] Interference with Human PTP-1B Expression by siRNA in Co-Transfection Assays

[0239] Human PTP1B encoding sequence was cloned into a Pmt vector according to standard molecular biology procedures (see Flint et al., *EMBO J.* 12:1937-46 (1993)). 1BKO+HIR cells were co-transfected with the human PTP-1B expression vector and siRNA polynucleotides (20 nM) specific for human PTP-1B sequences overnight using Lipofectamine 2000. Cells were lysed as described above, and the lysates were separated by 4-12% SDS-PAGE and transferred onto a PDVF membrane. The level of expression of human PTP-1B was determined by immunoblotting with an anti-human PTP-1B antibody (PHO2, Oncogene Research Products™, Inc. San Diego, Calif.). Interference with expression of human PTP-1B was observed with four siRNA polynucleotides as indicated in Table 14.

TABLE 12

_	sirna interference with muri		(PRESSION	_
Target	siRNA Sequence (SEQ ID NO)	siRNA Name	Related SEQ ID NO:	Decrease in Expression
Murine PTP1B	5'-gaagccagaggagcuauatt-3' 5'-cuacaccacauggccugactt-3' 5'-gacugccgaccagcugcgctt-3' 5'-gguaccgagaugucagccctt-3' 5'-ugacuauaucaaugccagctt-3' 5'-agaagaaaaggagaugguctt-3' 5'-cgggaagugcaaggagcuctt-3'	mPTP1B1.1 mPTP1B1.2 mPTP1B1.3 mPTP1B1.4 mPTP1B1.5 mPTP1B1.6 mPTP1B1.7		95% Not analyzed Not analyzed 25% Not analyzed 80% Not analyzed 80%

TABLE 13

	siRNA INTERFERENCE WITH HUM IN CO-TRANSFECTION		EXPRESSIO	N
Target	siRNA Sequence (SEQ ID NO)	siRNA Name	Related SEQ ID NO:	Decrease in Expression
Human PTP1B	5'-cuauaccacauggccugactt-3' 5'-gccaaaggaguuacauuctt-3' 5'-ggaagaaaaaggaagccctt-3' 5'-caaugggaaaugcagggagtt-3' 5'-ggaucaguggaaggagcuutc-3'	hPTP1B1.3 hPTP1B1.3	2 3 1	Not analyzed >95% >95% >95% >95%

[0240] Interference of Endogenous Expression of Human PTP-1B by siRNA

[0241] The effect of sequence specific siRNA on endogenous expression of human PTP-1B was examined in two different cell lines. HeLa cells were transfected as described above with HPTP1B1.1, hPTP1B1.2, hPTP1B1.3, hPTP1B1.4, and hPTP1B1.5 at 20 nM using Lipofectamine 2000, and after three days, the level of expression of PTP1B was analyzed by immunoblot. No significant decrease in

[0244] Interference with Expression of Human TCPTP by siRNA in Co-Transfection Assays

[0245] Co-transfection assays were performed essentially as described above for PTP1B expression analysis to determine siRNA inhibition of human TCPTP expression. A recombinant expression construct was prepared that encodes wild-type human TC45. The following oligonucleotide primers were used for the wild-type construct. The sequences of the BamHI and EcoRi restriction sites are underlined.

```
Human TC45 sense (TC45 5'BamHI)

5'-GGGGGGATCCATGCCCACCACCACCATGAGCGGGAGTT-3'

(SEQ ID NO___)

Human TC45 antisense (TC45 3'EcoRI)

5'-GGGGAATTCTTAGGTGTCTGTCAATCTTGGCCTTTTTCTTTTCGTTCA-3' (SEQ ID NO:___)
```

expression of human PTP-1B was observed in HeLa cells transfected with the siRNA hPTP1B1.1. In HeLa cells transfected with hPTP1B1.2 and hPTP1B1.4, the level of expression of human PTP-1B decreased 80%, and in cells transfected with hPTP1B1.3, the level of expression decreased 90%. Endogenous expression of human PTP-1B in the second cell line, 293-HEK-HIR, (gift from Julie Moyers, Eli Lilly and Company) transfected with sequence specific siRNAs hPTP1B1.2, hPTP1B1.3, hPTP1B1.4, hPTP1B1.5 (20 nM) was reduced by 90%.

[0242] Interference with Expression of Murine TCPTP by siRNA in Co-Transfection Assays

[0243] A co-transfection assay was performed in which 1BKO+HIR murine fibroblasts were co-transfected as described above with an expression vector comprising a polynucleotide sequence (SEQ ID NO:_) encoding murine TCPTP (SEQ ID NO:) and siRNA (SEQ mTCPTP1.1 (5'-guugucaugcuaaaccgaact-3')) (1 nM) or mTCPTP1.2 (5'-cagaacagagugaugguugag-3' (SEQ ID NO: __)) (20 nM). The level of TCPTP expression was determined by immunoblotting with an anti-human TCPTP antibody (Curt Diltz, CEPTYR, Inc.). The siRNA mTCPTP1.2 did not interfere with expression of murine TCPTP. Expression of murine TCPTP decreased more than 95% in cells transfected with siRNA, mTCPTP1.1.

[0246] Vector pCMVTag2B (Stratagene, La Jolla, Calif.) was digested with restriction endonuclease BamHI (New England Biolabs, Beverly, Mass.) for 3 hours at 37° C. The digested vector was then incubated with Klenow polymerase (New England Biolabs) for 15 minutes at 25° C. to fill in the recessed 3' termini, followed by an incubation of 30 minutes at 37° C. with calf intestinal phosphatase (New England Biolabs). The GATEWAYTM Reading Frame Cassette B (Invitrogen Life Technologies) was inserted into the pCMVTag2B vector by ligation with T4 DNA ligase (Invitrogen Life Technologies) overnight at 16° C. according to the supplier's instructions. DB3.1TM competent *E. coli* cells were transformed with the ligated vector (GWpCMVTag2) and DNA was isolated by standard molecular biology methods.

[0247] Vectors for expression of TC45 wild type were prepared as follows: The TC45 construct was subcloned into a GATEWAYTM entry vector pENTR3CTM (Invitrogen Life Technologies) by digesting 10 μ l of the TC45 cDNA with 1 μ l of BamHI (New England Biolabs), 1 μ l of EcoRI (New England Biolabs), 3 μ l 10×EcoRI buffer (New England Biolabs), 3 μ l 10×BSA (New England Biolabs), and 12 μ l distilled water for 3 hours at 37° C. Two microliters of the pENTR3CTM vector was digested with 0.5 μ l of BamHI (New England Biolabs), 0.5 μ l of EcoRI (New England Biolabs), 2 μ l 10×EcoRI buffer (New England Biolabs), 2 μ l 10×BSA (New England Biolabs), and 13 μ l distilled water for 3 hours at 37° C., followed by an incubation of 30

minutes at 37° C. with calf intestinal phosphatase (New England Biolabs). Digested DNA was run on a 1% agarose gel, digested bands were excised and gel purified using a QIAGEN Gel Extraction kit (QIAGEN, Inc.). Four microliters of the TC45 cDNA was ligated into 2 μ l of the pENTR3CTM vector overnight at 16° C. with 11 μl 10× Ligation Buffer (Invitrogen Life Technologies), 1 µl T4 DNA Ligase $(4U/\mu l)$ (Invitrogen Life Technologies), and 2 μ l distilled water. The construct was transformed into LIBRARY EFFICIENCY® DH5αTM cells. The FLAG® epitope-tagged TC45 construct was prepared by cloning the pENTR3CTM TC45 WT construct into the GWpCMVTag2 vector. The pENTR3C™ construct containing the TC45 polynucleotide was linearized by digesting the construct with Pvu I (New England Biolabs)) at 37° C. for 2 hours. The DNA was purified using a QIAGEN PCR Purification kit (QIAGEN, Inc.). Two microliters (150 ng/ μ l) of the GWpCMVTag2 vector were combined in a GATEWAY™ LR reaction with 3 μl linearized pENTR3CTM TC45 WT, 5 μ l TE buffer, 4 μ l ClonaseTM Enzyme, and 4 μ l LR reaction buffer (Invitrogen Life Technologies) overnight at room temperature. After addition of Proteinase K (Invitrogen Life Technologies) to the reaction for 10 minutes, LIBRARY EFFICIENCY® DH5 α^{TM} cells were transformed with the expression construct.

[0248] Cells (1BKO+HIR murine embryo fibroblasts) were co-transfected with an expression vector containing a nucleotide sequence encoding human TCPTP (SEQ ID NO:____) and siRNAs, hTCPTP1.4 (5'-guugu-caugcugaaccgcatt-3' (SEQ ID NO:____)) (20 nM); hTCPTP1.5 (5'-gcccauaugaucacagucgtg-3' (SEQ ID

human TCPTP was not affected by siRNA hTCPTP1.7. Expression levels decreased more than 95% in the cells co-transfected with hTCPTP1.4; 80% in cells co-transfected with hTCPTP1.5; and greater than 90% in cells transfected with hTCPTP1.6.

[0249] Interference of Endogenous Expression of Human TCPTP by siRNA

[0250] 293-HEK HIR cells were transfected with either hTCPTP1.4 (SEQ ID NO:_____) or rPTP1B1.2, a rat PTP1B sequence specific siRNA (5'-cggaugguggguggagguctt-3' (SEQ ID NO:_____), which was included as a nonspecific siRNA control, at concentrations of 2, 5, 10, 20 and 50 nM. Endogenous expression of human TCPTP in the cells transfected with sequence specific hTCPTP1.4 decreased 90%.

[0251] Transient Transfection of Human PTP1B and Sequence Specific Hairpin Vectors

[0252] Effectiveness of a human PTP1B sequence-specific siRNA in the form of a hairpin insert was examined in a transient co-transfection assay. Cells (1BKO+HIR mouse fibroblasts) were transfected with a human PTP1B expression vector (see above) and co-transfected with hPTP1B hairpin vectors (1, 0.5, and 0.25 µg) according to the transfection method described above. The human PTP1B specific sequences were inserted in frame with a human U6 small nuclear RNA promoter into a vector, which was a gift from David Engelke (University of Michigan, Ann Arbor, Mich.) (see also Paul et al., Nat. Biotechnol. 20:446-48 (2002)). The sequences of each strand inserted into the hairpin vectors are as follows.

hPTP1B H1.2-HP4
5'-tttGCCCAAAGGAGTTACATTCGTAAGAATGTAACTCCTTTGGGCtttt-3' (SEQ ID NO:___)
3' GGGTTTCCTCAATGTAAGCATTCTTACATTGAGGAAACCCGaaaaagatc-5' (SEQ ID NO:___)
hPTP1B H1.2-HP9
5'-tttGCCCAAAGGAGTTACATTCCCTGGGTAAGAATGTAACTCCTTTGGGCtttt-3' (SEQ ID NO:___)
3' GGGTTTCCTCAATGTAAGGGACCCATTCTTACATTGAGGAAACCCGaaaaagatc-5' (SEQ ID NO:___)

NO:______)) (10 nM); hTCPTP1.6 (5'-uegguuaaaugugca-caguac-3' (SEQ ID NO:_____)) (10 nM); or hTCPTP1.7 (5'-ugacuauccucauagaguggg-3' (SEQ ID NO:____)) (20 nM). Additional human TCPTP specific siRNA polynucle-otides were prepared; the sequences of each are as follows: hTCPTP1.1 (5'-agugagagaaucuggcucctt-3' (SEQ ID NO:____)); hTCPTP1.2 (5'-ggagagacuuaucuccugcctt-3' (SEQ ID NO:____)); and hTCPTP1.3 (5'-ggugac-cgauguacaggactt-3' (SEQ ID NO:____)). The level of TCPTP expression was determined by immunoblotting with an anti-human TCPTP antibody. The level of expression of

[0253] Twenty-four hours after the cells were transfected, cell lysates were prepared and expression of human PTP1B was determined by immunoblotting with an antihuman PTP1B antibody (see above). Cell lysates were also immunoblotted with an antibody specific for human insulin receptor beta chain (IR β) (Cat. No. C-19, Santa Cruz Biotechnology). The results are presented in **FIG. 19**.

[0254] Hairpin vectors are also prepared that contain sequences specific for murine PTP1B. The following sequences of each strand are inserted into a hairpin vector.

```
mPTP1BM1.1-HP4
5'-tttGAAGCCCAGAGGAGCTATAAGAATATAGCTCCTCTGGGCTTCtttt-3' (SEQ ID NO:__)
3' TTCGGGTCTCCTCGATATTCTTATATCGAGGAGACCCGAAGaaaaagatc-5' (SEQ ID NO:__)
mPTP1BM1.1-HP9
5'-tttGAAGCCCAGAGGAGCTATAGGGTGAGAATATAGCTCCTCTGGGCTTCtttt-3' (SEQ ID NO:__)
3' TTCGGGTCTCCTCGATATCCCACTCTTATATCGAGGAGACCCGAAGaaaaagatc-5' (SEQ ID NO:__)
```

EXAMPLE 6

Regulatory Role of TCPTP in Insulin Signaling

[0255] The protein tyrosine phosphatase TC-PTP exists in two alternatively spliced forms, TC45 and TC48, that share the same catalytic domain but differ at their extreme carboxy-termini (Mosinger et al., *Proc. Natl. Acad. Sci. USA* 89:499-503 (1992)). Insulin-induced oxidation and inactivation of TC45 suggested that it functions as a negative regulator of insulin signaling (see U.S. Ser. No. 10/366,547). This Example examines the regulatory role of TC45 in insulin signaling by inhibiting expression of the PTP by RNAi.

[0256] The specific siRNA duplexes were designed by first scanning through the open reading frame of TC45 mRNA and selecting sequences of 5'AA(N₁₉)3' (N=any nucleotide) for further characterization. The following 2 oligonucle-5'-AACAGAUACAwere chosen: GAGAUGUAAGC-3' (TCPTP1) (SEQ ID NO: 5'-AAGCCCA UAUGAUC ACAGUCG-3' (TCPTP2) (SEQ). These sequences were submitted to a BLAST search against human, rat, and mouse genome databases to ensure specificity for TC-PTP. The 21-nt siRNA duplexes were obtained in a deprotected and desalted form (Dharmacon Research). Rat-1 fibroblasts (Fischer rat fibroblast 3T3 like cell line) and HepG2 (human hepatocellular carcinoma) cells (American Type Culture Collection (ATCC), Manasass, Va.) were transfected with each siRNA at 100 nM. Both siRNA oligonucleotides suppressed expression of endogenous TC45 in the transfected HepG2 cells and Rat-1 fibroblasts, with TCPTP1 being more efficient.

[0257] Rat-1 (fibroblasts) and HepG2 (human hepatocellular carcinoma) cells were routinely maintained in DMEM supplemented with 10% FBS, 1% glutamine, 100 U/ml penicillin and 100 μ g/ml streptomycin. For stimulation with insulin, cells were plated in media containing 10% FBS for 48 hours, then serum-starved for 16 hours before treatment. For transient transfection, cells were plated in DMEM supplemented with 10% FBS for 16 hours, then in Opti-MEM (Invitrogen) without serum, after which the plasmid (5 μg/dish for Rat-1,30 μg/dish for HepG2) was introduced by LipofectAMINE and PLUS reagents (Invitrogen), according to the manufacture's recommendations. The transfection efficiency was routinely 40%. For RNAi experiments, cells were plated as above and the TCPTP siRNA duplexes were introduced by Oligofectamine (Invitrogen) according to the guidelines provided by Dharmacon Research Inc.

[0258] The potential regulatory role of TC45 in insulin signaling was investigated by examining the phosphorylation status of PKB/Akt, which is a critical effector in the P13 kinase pathway that mediates various intracellular responses to insulin, following ablation of the PTP by RNAi. The human hepatoma cell line HepG2 has been used extensively as a model to study insulin signaling (see Huang et al., *J. Biol. Chem.* 277:18151-60 (2002); Haj et al., *Science* 295 1708-11 (2002)). Serum-deprived Rat-I and HepG2 cells were exposed to 10 or 50 nM insulin for 5 min and lysed. The insulin receptor (IR) was immunoprecipitated from 500 μ g of cell lysate with anti-IR- β antibody 29B4 (Santa Cruz Biotechnology), then immunoblotted with anti-phosphotyrosine, anti-pYpY^{1162/1162}-IR- β (Biosource International,

Camarillo, Calif.) and anti-IR- β (C-19) (Santa Cruz Biotechnology) antibodies. HepG2 cells expressed higher levels of IR- β than Rat-1 cells as shown in **FIG. 20**A and displayed a robust response to insulin stimulation, as shown by the overall tyrosine phosphorylation level of IR- β and autophosphorylation of the activation loop tyrosines 1162 and 1163 (see **FIG. 20A**).

[0259] For the RNAi experiment, HepG2 cells were untransfected (control) or transfected (+siRNA) with 100 nM siRNA TCPTP1 oligonucleotide. Two days after transfection, cells were serum-starved for 16 hours and then stimulated with 10 nM insulin for 0, 1, 2, 5, 10, and 20 minutes. Total lysates (30 μ g) were immunoblotted with anti-phospho-PKB/Akt (Cell Signaling Technology, Beverly, Mass.); anti-PKB/Akt (Cell Signaling Technology); anti-TC45 (1910H (Lorenzen et al., J. Cell. Biol. 131:631-43 (1995))); and anti-PTP1B (FG6 (LaMontagne et al., Mol. Cell. Biol. 18:2965-75 (1998))) antibodies. The results presented in FIG. 20B indicate that depletion of TC45 enhanced both the intensity and duration of the signaling response. FIG. 20C illustrates a densitometric analysis of the gel image to show the ratio of phosphorylated PKB/Akt relative to total PKB/Akt. Similar results were observed in three independent experiments.

[0260] The role of TC45 in insulin signaling was further investigated by preparing a TC45 substrate trapping mutant. Substitution of an alanine residue for the invariant aspartate, which functions as a general acid in catalysis, into the vector expressing TC45 and into a vector expressing PTP1B was performed by standard site-directed mutagenesis protocols. HepG2 cells overexpressing wild type (WT) or trapping mutant (DA) forms of PTP1B and TC45 were either left untreated (-INS) or stimulated with 10 nM insulin for 5 min (+INS), then lysed in trapping buffer (20 mM Tris (pH 7.4), 1% NP-40, 150 mM NaCl, 10% glycerol, 10 mM IAA and 25 μ g/ml each of aprotinin and leupeptin). Aliquots (1 mg) of cell lysate were incubated with anti-PTP1B antibody (FG6) or anti-TC45 antibody (CF4). The immunocomplexes were washed with lysis buffer, subjected to SDS-PAGE then immunoblotted with anti-IR-β (C-19) antibody. An aliquot of lysate (30 μ g) was immunoblotted with anti-PTP1B antibody (FG6) or anti-TC-PTP antibody (CF4) to verify PTP expression. The data are shown in FIG. 21A and are representative of three independent experiments. These data suggest that TC45 recognizes IR-β as a substrate.

[0261] Serum starved, untransfected (control) or TC45 siRNA (100 nM) transfected (+siRNA) HepG2 cells were stimulated with 10 nM insulin for 0, 1, 2, 5, 10, and 20 minutes. The insulin receptor was immunoprecipitated from 750 μ g of cell lysate with anti-IR- β antibody 29B4 and immunoblotted with anti-phosphotyrosine (G104), anti-pY⁹⁷²- β (Biosource), anti-pYpY^{1162/1163}-IR- β , and anti-IR- β (C-19) antibodies as shown in FIG. 21B. FIG. 21C illustrates densitometric analyses of the gel image to show the ratio of phosphorylated IR- β relative to total IR- β for total phosphotyrosine (upper panel), phosphorylation of Tyr 972 (middle panel), and phosphorylation of the activation loop tyrosines 1162 and 1163 (lower panel). Similar results were observed in two independent experiments.

EXAMPLE 7

Effect of siRNAs Specific for PTP1B and TCPTP on Insulin Receptor Tyrosine Phosphorylation

[0262] This example illustrates the effect of RNAi on the function of components in a cell signaling pathway. The role of PTP1B in the down regulation of insulin signaling has been illustrated by data derived from a variety of approaches (Cheng et al., *Eur. J. Biochem.* 269:1050-59 (2002)), including the phenotype of the PTP1B knockout mouse (Elchebly et al., *Science* 283:1544-48 (1999); Klaman et al., *Mol. Cell Biol* 20:5479-89 (2000); see also U.S. patent application Ser. No. 10/366,547).

[0263] The effect of human PTP1B siRNA and of human TCPTP siRNA on the level of phosphorylation of IR-β was evaluated by ELISA. 292-HEK HIR cells were transfected with 0, 0.5, 3, or 10 nM siRNAs. The siRNA polynucleotides transfected into the cells included hPTP1B1.2 (SEQ), hPTP1B1.3 (SEQ ID NO: mPTP1B1.1 (SEQ ID NO:), rPTP1B1.2 (SEQ ID __), hTCPTP1.4 (SEQ ID NO:_ _), and the combination of hPTP1B1.3 and hTCPTP1.4. Seventy-two hours after transfection, cells were exposed to insulin for 7 minutes at concentrations of 0, 25, 50, 75, and 100 nM. Cell lysates were prepared as described in Example 1, and total cell protein was quantified by the Bio-Rad Protein Assay performed according to the manufacturer's instructions (BioRad, Hercules, Calif.). An ELISA was performed as follows. Dynex Immulon HB4X plates were coated with anti-insulin receptor antibody Ab-1 (1 mg/ml; NeoMarkers, Inc., Fremont, Calif.) that was diluted 1:1000 in CMF (calcium magnesium free)-PBS containing 5 µg/ml fatty acid free BSA (faf-BSA). The plates were incubated at 4° C. for at least four hours. The antibody solution was removed by aspiration, followed by the addition of 300 μ l of 3% faf-BSA+CMF-PBS. The plates were incubated for 1 hr with agitation on a vortex platform shaker (setting #5) at room temperature. After aspirating the 3% faf-BSA+CMF-PBS solution, approximately 10-20 µg of lysate were added to the wells and incubated at room temperature for one hour. Plates were washed three times with TBST (20 mM Tris-HCl, pH 7.5 150 mM NaCl; 0.05% Tween 20). An anti-insulin receptor phosphotyrosine specific antibody (pTyr 1162/63, Biosource International, Camarillo, Calif., Catalog #44-804) was diluted 1:2000 in TBST and added to the plates for one hour at room temperature. The plates were washed three times with TBST. HRP-conjugated anti-rabbit antibody (Amersham Biosciences, catalog #NA934V) (1:2000 in TBST) was then added to the wells and incubated at room temperature for one hour. The plates were washed three times with TBST and once with deionized, sterile water. TMB solution (Sigma Aldrich) (100 µl per well) was added and developed until a modest color change (10-30 minutes depending on cell type and insulin response). The reaction was stopped with 100 μ l of 1.8 N H₂SO₄ and then mixed. The optical density of each well was measured at 450 nM in a Spectramax plate reader (Molecular Devices Corp., Sunnyvale, Calif.). The data are presented in FIG. 22. The level of expression of PTP1B in each cell lysates was determined by immunoblot as described above. PTP1B polypeptide was detected using an anti-human PTP-1B antibody (PHO2, Oncogene Research Products™, Inc.). The amount of PTP1B expressed in cells transfected with varying concentrations of either siRNA was quantified by densitometric analysis of the immunoblot. The level of expression of human PTP1B is presented as a percent of the level of expression in cells that were not transfected with hPTP1B1.3 siRNA (i.e., the level of expression in untransfected cells equals 100%) (see tables in FIG. 22).

[0264] In a second experiment, 292-HEK HIR cells were transfected with 0, 0.5, 3, or 10 nM siRNAs. The siRNA polynucleotides transfected into the cells included), hPTP1B1.3 (SEQ ID hPTP1B1.2 (SEQ ID NO: mPTP1B1.1 (SEQ ID NO: hTCPTP1.4 (SEQ ID NO:_ _), and rPTP1B1.2 (SEQ ID). Seventy-two hours after transfection, cells were exposed to insulin for 7 minutes at concentrations of 0, 5, 10, 20, 50, and 100 nM. Cell lysates were prepared and total cell protein was quantified as described above. An ELISA was performed as described above. Cell lysates were coated onto 96-well plates, blocked, and probed with an anti-pYpY^{1162/1163}-IR-β antibody. Binding was detected using an enzyme conjugated secondary reagent. As shown in FIGS. 23 and 24, respectively, increased phosphorylation of the insulin receptor was observed in cells transfected with hPTP1B1.3 and with hTCPTP1.4.

[0265] The percent decrease in the level of PTP1B expression was compared with the level of phosphorylation of the insulin receptor. In three separate experements, 292-HEK HIR cells were transfected with 0, 0.5, 3, or 10 nM hPTP1B1.3 siRNA and then exposed to insulin for 7 minutes at concentrations of 0, 5, 10, 20, 50, and 100 nM. An ELISA and immunoblot of cell lysates were performed as described above. The effect of hPTP1B1.3 siRNA on the phosphorylation state of the insulin receptor is summarized in FIG. 25. Each data point represents the average optical density measured in duplicate wells.

EXAMPLE 8

Identification of Oncology Targets and Decreased Expression of the Targets by Specific siRNAs

[0266] This Example describes validation of DSP-3 as a target for oncology therapeutics. The Example also describes identification of siRNA polynucleotides that effectively interfere with expression of known chemotherapeutic target polypeptides.

[0267] Expression of DSP-3 polypeptide was evaluated in several cancer cell lines transfected with sequence specific DSP-3 siRNA polynucleotides and nonspecific siRNA polynucleotides. Cell lines included HeLa, HS578T; MDA-MB-231; MDA-MB-435 (breast cancer cell line that is ER-, Her²⁺, EGFR⁺, p53^{mut}, and invasive); MCF7 (breast cancer cell line that is ER⁺, Her2^{low}, EGFR^{low}, p53^{WT}, and non-invasive); T47D (breast cancer cell line that is ER⁺, Her2⁻, EGFR⁻, p53^{mut}, and non-invasive); HCT-116 (p53^{WT}); and HT-29 (p53^{mut}). Cells were transfected with 10 nM DSP3.1 ___), DSP3.4 (5'-ggugacacauauucugucutt-(SEQ ID NO: 3', (SEQ ID NO:____)), or Scr.2 (SEQ ID NO:_ (scrambled, a non-specific siRNA sequence not found in a human genome database), and then cell lysates were prepared and evaluated for expression of DSP-3 and inhibition of expression by specific siRNAs, as described in Example 1. Transfection efficiency of some cell lines with siRNA, for example, MC7 and T47D, was improved by using Lipofectamine™ 2000 according to manufacturer's recommendations (Invitrogen Life Technologies) rather than Oligofectamine™ (Invitrogen Life Technologies) for the transfection procedure. The level of expression of DSP-3 polypeptide in the presence of specific siRNA 4compared with the non-specific siRNA control was significantly decreased in MCF7, T47D, MD-MB-435, HCT-116, and HT-29 cells.

[0268] Interference with expression of known chemotherapeutic targets by RNAi was examined, and siRNA polynucleotides that effectively interfere with expression of the targets were identified. Targets included dihydrofolate reductase (DHFR) (GenBank Accession No. NM_000791) (SEQ ID NOs: and); thymidylate synthetase (GenBank Accession No. NM_001071) (SEQ ID); and topoisomerase I (GenBank and Accession No. J03250) (SEQ ID NOs: and The siRNA polynucleotides were designed according to methods described in Examples 1 and 2 and were manufactured by Dharmacon. Each siRNA was transfected into HeLa cells, and the effect of each on the endogenous expression of DHFR, thymidylate synthetase, and topoisomerase I was evaluated by immunoblotting of cell lysates as described in Example 1. The level of expression of the targets was determined by immunoblotting with an anti-DHFR monoclonal antibody (BD monoclonal antibody (diluted 1:250)); an anti-topoisomerase I antibody (Santa Cruz Biotechnology, Cat. No. sc-10783, diluted 1:200); and an anti-thymidylate synthetase antibody (Rockland sheep polyclonal antibody diluted 1:2000). The results are presented in Table 3.

[0271] A cell proliferation assay was also performed using a different cell line, T47D, and the same siRNAs. The data are presented in FIG. 27. The effect of silencing on proliferation was confirmed by cell counting. The number of T47D cells transfected with the nonspecific control siRNA scr.2 was approximately 200×10⁴. In T47D cells transfected with either DSP3.1 or DSP3.4 siRNA, the number of cells was approximately 75% of the negative control, and in the presence of DHFR.1, the number of cells was approximately 50% compared with cells transfected with the nonspecific control. Significantly decreased expression of DSP-3 and DHFR in cells transfected with the respective siRNAs was confirmed by immunoblot.

[0272] Silencing of DSP-3 in HCT-116 and T47D cells also induced proapoptotic signaling. HCT-116 cells and T47D cells were transfected with 10 nM of non-specific si RNA control scrb1.2 (SEQ ID NO:_____) (identical sequence to scr.2 described above), DSP3.1, DSP3.4, or DHFR.1. Three days after transfection of HCT-116 cells and

TABLE 14

SIRNA	siRNA	NA INTERFE	RENCE WIT	H ENDO	SENOUS	S EXPRESS	SION
OF DHFR,	F DHFR,	, THYMIDYL	ATE SYNTE	HETASE,	AND	TOPOISOM	ERASE

Target	siRNA Sequence (SEQ ID NO)	siRNA Name	Related SEQ ID NO:	Decrease in Expression
DHFR	5'-gaccugguucuccauuccutt-3'	DHFR.1		>90%
	5'-gcaguguauuugcuagguctt-3'	DHFR.3		>80%
	5'-gucagcgagcagguucucatt-3'	DHFR.4		>90%
Thymidylate Synthetase	5'-ccaaacguguguucuggaatt-3'	TYMS.1		>95%
	5'-ccaacccugacgacagaagtt-3'	TYMS.2		>90%
	5'-gccaggugacuuuauacactt-3'	TYMS.3		>95%
	5'-cccagaccuuucccaaagctt-3'	$\mathtt{TYMS.4}$		>90%
Topoisomerase I	5'-gauagagccuccuggacuutt-3'	TOP1.1		>90%
	5'-guccggcaugauaacaaggtt-3'	TOP1.2		>90%
	5'-ggagaaacagcggacacugtt-3'	TOP1.3		>80%
	5'-gcagcccgaggaugaucuutt-3'	TOP1.4		>80%

[0269] Interference of expression of another chemotherapeutic polypeptide target IKKgamma is performed according to the same procedures described above. The siRNA polynucleotides that are tested are IKK.1 (5'-gagucucucuggggaagett-3' (SEQ ID NO:_____)); IKK.2 (5'-gaguuccucaugugcaagtt-3' (SEQ ID NO:_____)); IKK.3 (5'-ggcucugugaaagcccagtt-3' (SEQ ID NO:_____)); and IKK.4 (5'-cacgcugcucuugauguggtt-3' (SEQ ID NO:_____)).

[0270] The effect of RNAi silencing on expression of DHFR was compared with silencing of DSP-3, Cdc14a, and SHP-2 polypeptide expression in a HCT-116 cell proliferation assay. HCT-116 cells were transfected with 2.5 nM of the following siRNA oligonucleotides: scr.2 (SEQ ID

five days after transfection of T47D cells, PARP assays were performed as described in Example 4. The results are presented in FIG. 28.

EXAMPLE 9

Inhibition of MAP Kinase Kinase Expression by RNAi

[0273] This Example describes interference of expression of MAP kinase kinases that are involved in the JNK signal transduction pathway in cells transfected with sequence specific siRNA polynucleotides.

[0274] Transient co-transfection experiments were performed as described in Example 2. 293-HEK cells were co-transfected with an expression vector that contained a polynucleotide sequence (GenBank Accession No. L36870)) that encoded FLAG®-tagged human (SEQ ID NO: MKK4 polypeptide (GenBank Accession No. L36870 (SEQ)) or with an expression vector that contained a polynucleotide sequence (GenBank Accession No. _)) that encoded FLAG®-AF013588 (SEQ ID NO:_ tagged human MKK7 polypeptide (GenBank Accession No. AF013588 (SEQ ID NO:)). The siRNA oligonucleotides were designed and prepared as described in Examples 1 and 2. The cells were transfected and the level of expression of each kinase was determined by immunoblotting with an anti-FLAG® monoclonal antibody as described in Example 2. The results are presented in Table 4.

[0283] Clemens et al., Proc. Natl. Acad. Sci. USA 97:6499-6503 (2000)

[0284] Elbashir et al., Genes & Development 15:188-200 (2001)

[0285] Elbashir, et al., *Nature* 411:494-498 (2001)

[**0286**] Fire et al., *Nature* 391:806-11 (1993)

[0287] Flint et al., Proc. Natl. Acad. Sci. USA 94:1680-1685 (1997)

[0288] Fukada et al., J. Biol. Chem. 276:25512-25519 (2001)

[0289] Harborth et al., J. Cell Sci. 114:4557-4565

TABLE 15

	sirna Interference WITH IN CO-TRANSFI			N
Target	siRNA Sequence (SEQ ID NO)	siRNA Name	Related SEQ ID NOs	Decrease in Expression
MKK4	5'-gugggcaaauaauggcagutt-3' 5'-cugugaaagcacuaaaccatt-3' 5'-ggagauccuccgcagcugatt-3' 5'-gcucuuuauacuuuggccutt-3' 5'-cacggacgucuucaucgcctt-3' 5'-gaagcggaugcagggccctt-3' 5'-cugcaagacggacuuugagtt-3'	MKK4 . 1 MKK4 . 2 MKK4 . 3 MKK4 . 4 MKK7 . 1 MKK7 . 2 MKK7 . 3 MKK7 . 4		80% 90% 90% 80% 10% 10%

EXAMPLE 10

Inhibition of Human P53 Expression by RNAi

[0275] An hairpin vector is prepared that contains a polynucleotide insert comprising a sequence that is a portion of a polynucleotide that encodes human p53 as described in Example 5. This sequence may be incorporated into a hairpin vector and transfected into a cell line known to express p53 (see Example 5). The level of expression of p53 is then determined by methods well known in the art, such as immunoblotting using an anti-p53 antibody (see Example 5). The p53 sequence incorporated into a hairpin vector is as follows.

[0276] HP53-HP9

[0277] 5'-tttGACTCCAGTGGTMTCTACTTCM-GAGAGTAGATTACCACTGGAGTCttttt-3' (SEQ ID NO:_____)

[0278] 3' tgaggtcaccattagatgaagttetetcatetaatggtgaceteagAAAAAGATC-5' (SEQ ID NO:)

ADDITIONAL REFERENCES

[0279] Agami et al., Cell 102:55-66 (2000)

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[0281] Brummelkamp et al., Science 296:550-53 (2002)

[0282] Carthew, Richard W., Current Opinion in Cell Biology 13:244-248 (2001)

[0290] Hutvagner et al., Curr. Opin. Gen. & Dev. 12:225-232 (2002)

[0291] Kisielow et al., Biochem. J. 363:1-5 (2002)

[0292] Paddison et al., Genes & Development 16:948-958 (2002)

[**0293**] Salmeen et al., *Moleular Cell* 6:1401-1412 (2000)

[0294] Scadden et al., EMBO Reports 2:1107-1111 (2001)

[**0295**] Sharp, Phillip A., Genes & Development 13:139-141 (1999)

[0296] Sharp, Phillip A., Genes & Development 15:485-490 (2001)

[0297] Shen et al., Proc. Natl. Acad. Sci. USA 24:13613-13618 (2001)

[0298] Sui et al., Proc. Natl. Acad. Sci. USA 99:5515-5520 (2002)

[0299] Tonks et al, Curr. Opin. Cell Biol. 13:182-195 (2001)

[0300] Tuschl, Thomas, *Chembiochem*. 2:239-245 (2001)

[0301] Ui-Tei et al., FEBS Letters 479:79-82 (2000)

[0302] Wen et al., Proc. Natl. Acad. Sci. 98:4622-4627 (2001)

[0303] Zamore et al., Cell 101:25-33 (2000)

[**0311**] WO 01/34815

[0312] WO 01/42443

[0304]	EP1 152 056	[0313] WO 01/68836
[0305]	U.S. Pat. No. 2001/0029617	[0314] WO 01/75164
[0306]	U.S. Pat. No. 2002/0007051	[0315] WO 01/92513
[0307]	U.S. Pat. No. 6,326,193	[0316] WO 01/96584
[0308]	U.S. Pat. No. 6,342,595	[0317] WO 99/32619
[0309]	U.S. Pat. No. 6,506,559	[0318] From the foregoi although specific embodim
[0310]	WO 01/29058	described herein for the

[0318] From the foregoing, it will be appreciated that, although specific embodiments of the invention have been described herein for the purpose of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the present invention is not limited except as by the appended claims.

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139

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12237 OTHER INTORMITORS II - ATTICIO OF O	
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		TION: 1, 2 R INFORMATIO	и• n – A П	C G or II			
		ENCE: 777	N. 11 - A,1,	.c,			
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acatc	ggcaa	cttcaaagat	gccagagacg	cggaacaatt	gagcaagaac	aaggtgacac	180
atatto	ctgtc	tgtccacgat	agtgccaggc	ctatgttgga	gggagttaaa	tacctgtgca	240
tccca	gcagc	ggattcacca	tctcaaaacc	tgacaagaca	tttcaaagaa	agtattaaat	300
tcatto	cacga	gtgccggctc	cgcggtgaga	gctgccttgt	acactgcctg	gccggggtct	360
ccagga	agcgt	gacactggtg	atcgcataca	tcatgaccgt	cactgacttt	ggctgggagg	420
atgcco	ctgca	caccgtgcgt	gctgggagat	cctgtgccaa	ccccaacgtg	ggcttccaga	480
gacago	ctcca	ggagtttgag	aagcatgagg	tccatcagta	tcggcagtgg	ctgaaggaag	540
aatato	ggaga	gagccctttg	caggatgcag	aagaagccaa	aaacattctg	gccgctccag	600
gaatto	ctgaa	gttctgggcc	tttctcagaa	gactgtaatg	tacctgaagt	ttctgaaata	660
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His Ile Leu Ser Val His Asp Ser Ala Arg Pro Met Leu Glu Gly Val 35 40 45			
Lys Tyr Leu Cys Ile Pro Ala Ala Asp Ser Pro Ser Gln Asn Leu Thr 50 55 60			
Arg His Phe Lys Glu Ser Ile Lys Phe Ile His Glu Cys Arg Leu Arg 65 70 75 80			
Gly Glu Ser Cys Leu Val His Cys Leu Ala Gly Val Ser Arg Ser Val 85 90 95			
Thr Leu Val Ile Ala Tyr Ile Met Thr Val Thr Asp Phe Gly Trp Glu 100 105 110			
Asp Ala Leu His Thr Val Arg Ala Gly Arg Ser Cys Ala Asn Pro Asn 115 120 125			
Val Gly Phe Gln Arg Gln Leu Gln Glu Phe Glu Lys His Glu Val His 130 135 140			
Gln Tyr Arg Gln Trp Leu Lys Glu Glu Tyr Gly Glu Ser Pro Leu Gln 145 150 155 160			
Asp Ala Glu Glu Ala Lys Asn Ile Leu Ala Ala Pro Gly Ile Leu Lys 165 170 175			
Phe Trp Ala Phe Leu Arg Arg Leu 180			
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gtgcttccgg gccggctggc gggactggcg ctgccgcggc tccccgccca ctaccagttc	120		
ctgttggacc tgggcgtgcg gcacctggtg tccctgacgg agcgcgggcc ccctcacagc	180		
gacagetgee eeggeeteac eetgeacege etgegeatee eegaettetg eeegeeggee	240		
cccgaccaga tcgaccgctt cgtgcagatc gtggacgagg ccaacgcacg gggagaggct	300		
gtgggagtgc actgtgctct gggctttggc cgcactggca ccatgctggc ctgttacctg	360		
gtgaaggage ggggettgge tgeaggagat geeattgetg aaateegaeg actaegaece	420		
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taaggggcct tagtaccctt ctaccaggcc ctcactcccc ttccccatgt tgtcgatggg	540										
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cactgaagta gcccacccct gcaggcaggt cctgattgaa ggggaggctt gtactgcttt	660										
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Ala Gly Leu Ala Leu Pro Arg Leu Pro Ala His Tyr Gln Phe Leu Leu 20 25 30											
Asp Leu Gly Val Arg His Leu Val Ser Leu Thr Glu Arg Gly Pro Pro 35 40 45											
His Ser Asp Ser Cys Pro Gly Leu Thr Leu His Arg Leu Arg Ile Pro 50 55 60											
Asp Phe Cys Pro Pro Ala Pro Asp Gln Ile Asp Arg Phe Val Gln Ile 65 70 75 80											
Val Asp Glu Ala Asn Ala Arg Gly Glu Ala Val Gly Val His Cys Ala 85 90 95											
Leu Gly Phe Gly Arg Thr Gly Thr Met Leu Ala Cys Tyr Leu Val Lys 100 105 110											
Glu Arg Gly Leu Ala Ala Gly Asp Ala Ile Ala Glu Ile Arg Arg Leu 115 120 125											
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cagoctotgo tgcaggatat cacotacott ogcatocogg togotgatac cootgaggta	300										
cccatcaaaa agcacttcaa agaatgtatc aacttcatcc actgctgccg ccttaatggg	360										
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tatgtgatga ctgtgacggg gctaggctgg cgggacgtgc ttgaagccat caaggccacc	480 540										
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gaggaggagt tgcgcgctt gctgccgctt tgcaagcgct gccggcaggg ctccgcgacc	660										

teggeeteet eegeegggee geacteagea geeteegagg gaacegtgea gegeetggtg	720
ccgcgcacgc cccgggaage ccaccggccg ctgccgctgc tggcgcgcgt caagcagact	780
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His Ile Ile Ser Ile His Glu Ser Pro Gln Pro Leu Leu Gln Asp Ile 35 40 45	
Thr Tyr Leu Arg Ile Pro Val Ala Asp Thr Pro Glu Val Pro Ile Lys 50 55 60	
Lys His Phe Lys Glu Cys Ile Asn Phe Ile His Cys Cys Arg Leu Asn 65 70 75 80	
Gly Gly Asn Cys Leu Val His Cys Phe Ala Gly Ile Ser Arg Ser Thr 85 90 95	
Thr Ile Val Thr Ala Tyr Val Met Thr Val Thr Gly Leu Gly Trp Arg	
Asp Val Leu Glu Ala Ile Lys Ala Thr Arg Pro Ile Ala Asn Pro Asn 115 120 125	
Pro Gly Phe Arg Gln Gln Leu Glu Glu Phe Gly Trp Ala Ser Ser Gln 130 135 140	
Lys Leu Arg Arg Gln Leu Glu Glu Arg Phe Gly Glu Ser Pro Phe Arg 145 150 155 160	
Asp Glu Glu Glu Leu Arg Ala Leu Leu Pro Leu Cys Lys Arg Cys Arg 165 170 175	
Gln Gly Ser Ala Thr Ser Ala Ser Ser Ala Gly Pro His Ser Ala Ala 180 185 190	
Ser Glu Gly Thr Val Gln Arg Leu Val Pro Arg Thr Pro Arg Glu Ala 195 200 205	
His Arg Pro Leu Pro Leu Leu Ala Arg Val Lys Gln Thr Phe Ser Cys 210 215 220	
Leu Pro Arg Cys Leu Ser Arg Lys Gly Gly Lys 225 230 235	
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tggcccgcaa gaggaaggcg cccctcgctt gcacctgcag cctcggtggc cccgacatga	180
ttccttactt ctccgccaac gcggtcatct cgcagaacgc catcaaccag ctcatcagcg	240
agagetttet aactgteaaa ggtgetgeee tttttetace aeggggaaat ggeteateea	300

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caatgttcat	tttactccgc	ccagaagaca	acatcaggct	ggctgtaaga	ctggaaagta	420
cttaccagaa	tcgaacacgc	tatatggtag	tggtttcaac	taatggtaga	caagacactg	480
aagaaagcat	cgtcctagga	atggatttct	cctctaatga	cagtagcact	tgtaccatgg	540
gcttagtttt	gcctctctgg	agcgacacgc	taattcattt	ggatggtgat	ggtgggttca	600
gtgtatcgac	ggataacaga	gttcacatat	tcaaacctgt	atctgtgcag	gcaatgtggt	660
ctgcactaca	gagcttacac	aaggcttgtg	aagtcgccag	agcgcataac	tactacccag	720
gcagcctatt	tctcacttgg	gtgagttatt	atgagagcca	tatcaactca	gatcaatcct	780
cagtcaatga	atggaatgca	atgcaagatg	tacagtccca	ccggcccgac	tctccagctc	840
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atgtcactcg	agagatagat	aacttcttcc	caggagtctt	tgagtatcat	aacattcggg	1200
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ctaaagcaaa	gaaacatgga	tctaaatgcc	ttgtgcactg	caaaatgggg	gtgagtcgct	1320
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gttcaccctc	ttgctgcaac	cctgagaagt	tacttcacat	ttctcatcct	tacctgaccc	1620
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<210> SEQ ID NO 785

<211> LENGTH: 509

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 785

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Leu Thr Val Lys Gly Ala Ala Leu Phe Leu Pro Arg Gly Asn Gly Ser 50 60

Ser Thr Pro Arg Ile Ser His Arg Arg Asn Lys His Ala Gly Asp Leu 65 70 75 80

Gln Gln His Leu Gln Ala Met Phe Ile Leu Leu Arg Pro Glu Asp Asn 85 90 95

Ile Arg Leu Ala Val Arg Leu Glu Ser Thr Tyr Gln Asn Arg Thr Arg

			100					105					110		
Tyr	Met	Val 115	Val	Val	Ser	Thr	Asn 120	Gly	Arg	Gln	Asp	Thr 125	Glu	Glu	Ser
Ile	Val 130	Leu	Gly	Met	Asp	Phe 135	Ser	Ser	Asn	Asp	Ser 140	Ser	Thr	Cys	Thr
Met 145	Gly	Leu	Val	Leu	Pro 150	Leu	Trp	Ser	Asp	Thr 155	Leu	Ile	His	Leu	Asp 160
Gly	Asp	Gly	Gly	Phe 165	Ser	Val	Ser	Thr	Asp 170	Asn	Arg	Val	His	Ile 175	Phe
Lys	Pro	Val	Ser 180	Val	Gln	Ala	Met	Trp 185	Ser	Ala	Leu	Gln	Ser 190	Leu	His
Lys	Ala	Cys 195	Glu	Val	Ala	Arg	Ala 200	His	Asn	Tyr	Tyr	Pro 205	Gly	Ser	Leu
Phe	Leu 210	Thr	Trp	Val	Ser	Tyr 215	Tyr	Glu	Ser	His	Ile 220	Asn	Ser	Asp	Gln
Ser 225	Ser	Val	Asn	Glu	Trp 230	Asn	Ala	Met	Gln	Asp 235	Val	Gln	Ser	His	Arg 240
Pro	Asp	Ser	Pro	Ala 245	Leu	Phe	Thr	Asp	Ile 250	Pro	Thr	Glu	Arg	Glu 255	Arg
Thr	Glu	Arg	Leu 260	Ile	Lys	Thr	Lys	Leu 265	Arg	Glu	Ile	Met	Met 270	Gln	Lys
Asp	Leu	Glu 275	Asn	Ile	Thr	Ser	Lys 280	Glu	Ile	Arg	Thr	Glu 285	Leu	Glu	Met
Gln	Met 290	Val	Cys	Asn	Leu	Arg 295	Glu	Phe	Lys	Glu	Phe 300	Ile	Asp	Asn	Glu
Met 305	Ile	Val	Ile	Leu	Gly 310	Gln	Met	Asp	Ser	Pro 315	Thr	Gln	Ile	Phe	Glu 320
His	Val	Phe	Leu	Gly 325	Ser	Glu	Trp	Asn	Ala 330	Ser	Asn	Leu	Glu	Asp 335	Leu
Gln	Asn	Arg	Gly 340	Val	Arg	Tyr	Ile	Leu 345	Asn	Val	Thr	Arg	Glu 350	Ile	Asp
Asn	Phe	Phe 355	Pro	Gly	Val	Phe	Glu 360	Tyr	His	Asn	Ile	Arg 365	Val	Tyr	Asp
Glu	Glu 370	Ala	Thr	Asp	Leu	Leu 375	Ala	Tyr	Trp	Asn	Asp 380	Thr	Tyr	Lys	Phe
Ile 385	Ser	Lys	Ala	Lys	Lys 390	His	Gly	Ser	Lys	С у в 395	Leu	Val	His	Cys	Lys 400
Met	Gly	Val	Ser	Arg 405	Ser	Ala	Ser	Thr	Val 410	Ile	Ala	Tyr	Ala	Met 415	Lys
Glu	Tyr	Gly	Trp 420	Asn	Leu	Asp	Arg	Ala 425	Tyr	Asp	Tyr	Val	Lys 430	Glu	Arg
Arg	Thr	Val 435	Thr	Lys	Pro	Asn	Pro 440	Ser	Phe	Met	Arg	Gln 445	Leu	Glu	Glu
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Arg 465	Asp	Lys	Pro	Trp	Gly 470	Glu	Lys	Ser	Thr	Glu 475	Phe	Glu	Ser	Val	Asp 480
Leu	Val	Ser	Ile	Pro 485	Gly	Ser	Pro	Ser	Cys 490	Cys	Asn	Pro	Glu	Lys 495	Leu
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60

											-	con	tin	ued	
			100					105					110		
Ala	Asp	Asp 115	Leu	Pro	Thr	Phe	Asp 120	Leu	Ser	Val	Phe	Phe 125	Tyr	Pro	Ala
Ala	Ala 130	Phe	Ile	Asp	Arg	Ala 135	Leu	Ser	Asp	Asp	His 140	Ser	Lys	Ile	Leu
Val 145	His	Сув	Val	Met	Gl y 150	Arg	Ser	Arg	Ser	Ala 155	Thr	Leu	Val	Leu	Ala 160
Tyr	Leu	Met	Ile	His 165	Lys	Asp	Met	Thr	Leu 170	Val	Asp	Ala	Ile	Gln 175	Gln
Val	Ala	Lys	Asn 180	Arg	Сув	Val	Leu	Pro 185	Asn	Arg	Gly	Phe	Leu 190	Lys	Gln
Leu	Arg	Glu 195	Leu	Asp	Lys	Gln	Leu 200	Val	Gln	Gln	Arg	Arg 205	Arg	Ser	Gln
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cgc	ggaga	atg 🤉	gttt	cacco	ca aa	atato	cacto	ggt	gtgga	aggc	agaa	aaac	cta (ctgtt	gacaa
gag	gagti	tga 1	tggca	agtt	tt ti	tggc	aaggo	cta	agtaa	aaag	taad	cact	gga	gactt	cacac
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- 4															+

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Gly Ala Val Thr His Ile Lys Ile Gln Asn Thr Gly Asp Tyr Tyr Asp 50 60

Leu Tyr Gly Gly Glu Lys Phe Ala Thr Leu Ala Glu Leu Val Gln Tyr 65 70 75 80

Tyr Met Glu His His Gly Gln Leu Lys Glu Lys Asn Gly Asp Val Ile $85 \hspace{1cm} 90 \hspace{1cm} 95$

Glu Leu Lys Tyr Pro Leu Asn Cys Ala Asp Pro Thr Ser Glu Arg Trp \$100\$ \$105\$ \$110\$

Phe His Gly His Leu Ser Gly Lys Glu Ala Glu Lys Leu Leu Thr Glu 115 120 125

Lys Gly Lys His Gly Ser Phe Leu Val Arg Glu Ser Gln Ser His Pro $130 \ \ 135 \ \ 140$

Gly Asp Phe Val Leu Ser Val Arg Thr Gly Asp Asp Lys Gly Glu Ser 145 150155155160

Asn Asp Gly Lys Ser Lys Val Thr His Val Met Ile Arg Cys Gln Glu 165 170 175

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Leu Val Glu His Tyr Lys Lys Asn Pro Met Val Glu Thr Leu Gly Thr

Arg

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Glu 225	Ile	Glu	Ser	Arg	Val 230	Arg	Glu	Leu	Ser	Lys 235	Leu	Ala	Glu	Thr	Thr 240
Asp	Lys	Val	Lys	Gln 245	Gly	Phe	Trp	Glu	Glu 250	Phe	Glu	Thr	Leu	Gln 255	Gln
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Lys	Ty r 370	Trp	Pro	Asp	Glu	Tyr 375	Ala	Leu	Lys	Glu	Tyr 380	Gly	Val	Met	Arg
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Val	Arg	Ser	Gln 500	Arg	Ser	Gly	Met	Val 505	Gln	Thr	Glu	Ala	Gln 510	Tyr	Arg
Phe	Ile	Tyr 515	Met	Ala	Val	Gln	His 520	Tyr	Ile	Glu	Thr	Leu 525	Gln	Arg	Arg
Ile	Glu 530	Glu	Glu	Gln	Lys	Ser 535	Lys	Arg	Lys	Gly	His 540	Glu	Tyr	Thr	Asn
Ile 545	Lys	Tyr	Ser	Leu	Ala 550	Asp	Gln	Thr	Ser	Gly 555	Asp	Gln	Ser	Pro	Leu 560
Pro	Pro	Сув	Thr	Pro 565	Thr	Pro	Pro	Сув	Ala 570	Glu	Met	Arg	Glu	Asp 575	Ser
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Ala Trp Leu Leu 50	Leu Pro Leu 55	Leu Leu Leu	Leu Leu Val	Leu Leu Leu
Ala Ala Tyr Phe 65	Phe Arg Phe	Arg Lys Gln	Arg Lys Ala 75	Val Val Ser 80
Thr Ser Asp Lys	Lys Met Pro 85	Asn Gly Ile 90	Leu Glu Glu	Gln Glu Gln 95
Gln Arg Val Met 100	Leu Leu Ser	Arg Ser Pro 105	Ser Gly Pro	Lys Lys Tyr 110
Phe Pro Ile Pro 115	Val Glu His	Leu Glu Glu 120	Glu Ile Arg 125	Ile Arg Ser
Ala Asp Asp Cys 130	Lys Gln Phe 135		Phe Asn Ser 140	Leu Pro Ser
Gly His Ile Gln 145	Gly Thr Phe 150	Glu Leu Ala	Asn Lys Glu 155	Glu Asn Arg 160
Glu Lys Asn Arg	Tyr Pro Asn 165	Ile Leu Pro 170		Ser Arg Val 175
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Ser Tyr Ile Asp 195	Gly Tyr Lys	Glu Lys Asn 200	Lys Phe Ile 205	Ala Ala Gln
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Gln Lys Ser Ala 225	Thr Ile Val 230	Met Leu Thr	Asn Leu Lys 235	Glu Arg Lys 240
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Ala Gly Val Gly 340	Arg Thr Gly	Thr Phe Ile 345	Val Ile Asp	Ala Met Met 350
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Thr Phe Ile Tyr	Gln Ala Leu	Leu Glu Tyr	Tyr Leu Tyr	Gly Asp Thr

385 390 395 400	
Glu Leu Asp Val Ser Ser Leu Glu Lys His Leu Gln Thr Met His Gly 405 410 415	
Thr Thr Thr His Phe Asp Lys Ile Gly Leu Glu Glu Glu Phe Arg Lys 420 425 430	
Leu Thr Asn Val Arg Ile Met Lys Glu Asn Met Arg Thr Gly Asn Leu 435 440 445	
Pro Ala Asn Met Lys Lys Ala Arg Val Ile Gln Ile Ile Pro Tyr Asp 450 455 460	
Phe Asn Arg Val Ile Leu Ser Met Lys Arg Gly Gln Glu Tyr Thr Asp 465 470 475 480	
Tyr Ile Asn Ala Ser Phe Ile Asp Gly Tyr Arg Gln Lys Asp Tyr Phe 485 490 495	
Ile Ala Thr Gln Gly Pro Leu Ala His Thr Val Glu Asp Phe Trp Arg 500 505 510	
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Gln Glu Arg Glu Gln Asp Lys Cys Tyr Gln Tyr Trp Pro Thr Glu Gly 530 535 540	
Ser Val Thr His Gly Glu Ile Thr Ile Glu Ile Lys Asn Asp Thr Leu 545 550 560	
Ser Glu Ala Ile Ser Ile Arg Asp Phe Leu Val Thr Leu Asn Gln Pro 565 570 575	
Gln Ala Arg Gln Glu Glu Gln Val Arg Val Val Arg Gln Phe His Phe 580 585 590	
His Gly Trp Pro Glu Ile Gly Ile Pro Ala Glu Gly Lys Gly Met Ile 595 600 605	
Asp Leu Ile Ala Ala Val Gln Lys Gln Gln Gln Gln Thr Gly Asn His 610 615 620	
Pro Ile Thr Val His Cys Ser Ala Gly Ala Gly Arg Thr Gly Thr Phe 625 630 635 640	
Ile Ala Leu Ser Asn Ile Leu Glu Arg Val Lys Ala Glu Gly Leu Leu 645 650 655	
Asp Val Phe Gln Ala Val Lys Ser Leu Arg Leu Gln Arg Pro His Met 660 665 670	
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240

300

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<210> SEQ ID NO 795 <211> LENGTH: 642 <212> TYPE: PRT <213> ORGANISM: Homo sapiens

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Ser	Gly 50	Pro	Lys	Lys	Tyr	Phe 55	Pro	Ile	Pro	Val	Glu 60	His	Leu	Glu	Glu
Glu 65	Ile	Arg	Ile	Arg	Ser 70	Ala	Asp	Asp	Cys	Lys 75	Gln	Phe	Arg	Glu	Glu 80
Phe	Asn	Ser	Leu	Pro 85	Ser	Gly	His	Ile	Gln 90	Gly	Thr	Phe	Glu	Leu 95	Ala
Asn	Lys	Glu	Glu 100	Asn	Arg	Glu	Lys	Asn 105	Arg	Tyr	Pro	Asn	Ile 110	Leu	Pro
Asn	Asp	His 115	Ser	Arg	Val	Ile	Leu 120	Ser	Gln	Leu	Asp	Gly 125	Ile	Pro	Cys
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Lys 145	Phe	Ile	Ala	Ala	Gln 150	Gly	Pro	Lys	Gln	Glu 155	Thr	Val	Asn	Asp	Phe 160
Trp	Arg	Met	Val	Trp 165	Glu	Gln	Lys	Ser	Ala 170	Thr	Ile	Val	Met	Leu 175	Thr
Asn	Leu	Lys	Glu 180	Arg	Lys	Glu	Glu	Lys 185	Cys	His	Gln	Tyr	Trp 190	Pro	Asp
Gln	Gly	Cys 195	Trp	Thr	Tyr	Gly	Asn 200	Ile	Arg	Val	Сув	Val 205	Glu	Asp	Cys
Val	Val 210	Leu	Val	Asp	Tyr	Thr 215	Ile	Arg	Lys	Phe	Cys 220	Ile	Gln	Pro	Gln
Leu 225	Pro	Asp	Gly	Суѕ	Lys 230	Ala	Pro	Arg	Leu	Val 235	Ser	Gln	Leu	His	Phe 240
Thr	Ser	Trp	Pro	Asp 245	Phe	Gly	Val	Pro	Phe 250	Thr	Pro	Ile	Gly	Met 255	Leu
Lys	Phe	Leu	Lys 260	Lys	Val	Lys	Thr	Leu 265	Asn	Pro	Val	His	Ala 270	Gly	Pro
Ile	Val	Val 275	His	Суѕ	Ser	Ala	Gly 280	Val	Gly	Arg	Thr	Gly 285	Thr	Phe	Ile
Val	Ile 290	Asp	Ala	Met	Met	Ala 295	Met	Met	His	Ala	Glu 300	Gln	Lys	Val	Asp
Val 305	Phe	Glu	Phe	Val	Ser 310	Arg	Ile	Arg	Asn	Gln 315	Arg	Pro	Gln	Met	Val 320
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Tyr	Leu	Tyr	Gly 340	Asp	Thr	Glu	Leu	Asp 345	Val	Ser	Ser	Leu	Glu 350	Lys	His
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Met 385	Arg	Thr	Gly	Asn	Leu 390	Pro	Ala	Asn	Met	Lys 395	Lys	Ala	Arg	Val	Ile 400
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Ile	Lys	Asn	Asp 500	Thr	Leu	Ser	Glu	Ala 505	Ile	Ser	Ile	Arg	Asp 510	Phe	Leu	
Val	Thr	Leu 515	Asn	Gln	Pro	Gln	Ala 520	Arg	Gln	Glu	Glu	Gln 525	Val	Arg	Val	
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Gln	Gln	Thr	Gly	Asn 565	His	Pro	Ile	Thr	Val 570	His	Cys	Ser	Ala	Gly 575	Ala	
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Lys	Ala	Glu 595	Gly	Leu	Leu	qaA	Val 600	Phe	Gln	Ala	Val	L y s 605	Ser	Leu	Arg	
Leu	Gln 610	Arg	Pro	His	Met	Val 615	Gln	Thr	Leu	Glu	Gln 620	Tyr	Glu	Phe	Cys	
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ggti	tate	gtg d	ctctt	tcca	gg tt	gtaa	attt	. aaa	agato	gtta	gaag	gaaat	gt o	ccaaa	aaagat	240
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aago	caata	aca a	attat	tctt	ca to	gagtt	teg	g gad	caaat	tag	ctgo	cacat	ct a	atcat	caaga	660

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720

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Сув	Leu 130	Lys	Asn	Tyr	Arg	Lys 135	Thr	Leu	Ile	His	Cys 140	Tyr	Gly	Gly	Leu	
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<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

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Cys Glu Val Thr Tyr Asp Lys Thr Pro Leu Glu Lys Asp Gly Ile Thr 50 60

Val Val Asp Trp Pro Phe Asp Asp Gly Ala Pro Pro Pro Gly Lys Val 65 70 75 80

Val Glu Asp Trp Leu Ser Leu Val Lys Ala Lys Phe Cys Glu Ala Pro 85 90 95

Gly Ser Cys Val Ala Val His Cys Val Ala Gly Leu Gly Arg Ala Pro $100 \hspace{1.5cm} 105 \hspace{1.5cm} 105$

Val Leu Val Ala Leu Ala Leu Ile Glu Ser Gly Met Lys Tyr Glu Asp 115 120 125

Ala Ile Gln Phe Ile Arg Gln Lys Arg Arg Gly Ala Ile Asn Ser Lys 130 140

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<210> SEQ ID NO 801
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<211> LENGTH: 167

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 801

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<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 802

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cacaccacca	tcctccgacc	ctcctacacc	gggctttctt	cttcttcagc	gagattcctg	1740
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<210> SEQ]						

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 803

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Asn Thr His Tyr Phe Ser Ile Asp Glu Glu Leu Val Tyr Glu Asn Phe $35 \hspace{1cm} 40 \hspace{1cm} 45$

Tyr Ala Asp Phe Gly Pro Leu Asn Leu Ala Met Val Tyr Arg Tyr Cys 50 60

Cys Lys Leu Asn Lys Lys Leu Lys Ser Tyr Ser Leu Ser Arg Lys Lys 65 70 75 80

Ile Val His Tyr Thr Cys Phe Asp Gln Arg Lys Arg Ala Asn Ala Ala 85 90 95

Phe Leu Ile Gly Ala Tyr Ala Val Ile Tyr Leu Lys Lys Thr Pro Glu 100 $$105\$

Glu Ala Tyr Arg Ala Leu Leu Ser Gly Ser Asn Pro Pro Tyr Leu Pro

Phe Arg Asp Ala Ser Phe Gly Asn Cys Thr Tyr Asn Leu Thr Ile Leu 130 135 140

Asp Cys Leu Gln Gly Ile Arg Lys Gly Leu Gln His Gly Phe Phe Asp 145 150 155 160

Phe Glu Thr Phe Asp Val Asp Glu Tyr Glu His Tyr Glu Arg Val Glu 165 170 175

Asn Gly Asp Phe Asn Trp Ile Val Pro Gly Lys Phe Leu Ala Phe Ser 180 \$180\$

Gly Pro His Pro Lys Ser Lys Ile Glu Asn Gly Tyr Pro Leu His Ala 195 200205

Pro Glu Ala Tyr Phe Pro Tyr Phe Lys Lys His Asn Val Thr Ala Val 210 215 220

Val Arg Leu Asn Lys Lys Ile Tyr Glu Ala Lys Arg Phe Thr Asp Ala 225 230 235 240

Gly	Phe	Glu	His	Tyr 245	Asp	Leu	Phe	Phe	Ile 250	Asp	Gly	Ser	Thr	Pro 255	Ser
Asp	Asn	Ile	Val 260	Arg	Arg	Phe	Leu	Asn 265	Ile	Сув	Glu	Asn	Thr 270	Glu	Gly
Ala	Ile	Ala 275	Val	His	Сув	Lys	Ala 280	Gly	Leu	Gly	Arg	Thr 285	Gly	Thr	Leu
Ile	Ala 290	Сув	Tyr	Val	Met	L y s 295	His	Tyr	Arg	Phe	Thr 300	His	Ala	Glu	Ile
Ile 305	Ala	Trp	Ile	Arg	Ile 310	Cys	Arg	Pro	Gly	Ser 315	Ile	Ile	Gly	Pro	Gln 320
Gln	His	Phe	Leu	Glu 325	Glu	Lys	Gln	Ala	Ser 330	Leu	Trp	Val	Gln	Gly 335	Asp
Ile	Phe	Arg	Ser 340	Lys	Leu	Lys	Asn	Arg 345	Pro	Ser	Ser	Glu	Gly 350	Ser	Ile
Asn	Lys	Ile 355	Leu	Ser	Gly	Leu	Asp 360	Asp	Met	Ser	Ile	Gly 365	Gly	Asn	Leu
Ser	Lys 370	Thr	Gln	Asn	Met	Glu 375	Arg	Phe	Gly	Glu	Asp 380	Asn	Leu	Glu	Asp
Asp 385	Asp	Val	Glu	Met	Lys 390	Asn	Gly	Ile	Thr	Gln 395	Gly	Asp	Lys	Leu	Arg 400
Ala	Leu	Lys	Ser	Gln 405	Arg	Gln	Pro	Arg	Thr 410	Ser	Pro	Ser	Сув	Ala 415	Phe
Arg	Ser	Asp	Asp 420	Thr	Lys	Gly	His	Pro 425	Arg	Ala	Val	Ser	Gln 430	Pro	Phe
Arg	Leu	Ser 435	Ser	Ser	Leu	Gln	Gly 440	Ser	Ala	Val	Thr	Leu 445	Lys	Thr	Ser
Lys	Met 450	Ala	Leu	Ser	Pro	Ser 455	Ala	Thr	Ala	Lys	Arg 460	Ile	Asn	Arg	Thr
Ser 465	Leu	Ser	Ser	Gly	Ala 470	Thr	Val	Arg	Ser	Phe 475	Ser	Ile	Asn	Ser	Arg 480
Leu	Ala	Ser	Ser	Leu 485	Gly	Asn	Leu	Asn	Ala 490	Ala	Thr	Asp	Asp	Pro 495	Glu
Asn	Lys	Lys	Thr 500	Ser	Ser	Ser	Ser	L y s 505	Ala	Gly	Phe	Thr	Ala 510	Ser	Pro
Phe	Thr	Asn 515	Leu	Leu	Asn	Gly	Ser 520	Ser	Gln	Pro	Thr	Thr 525	Arg	Asn	Tyr
	Glu 530					535					540				
Gly 545	Asn	Leu	Asn	Ser	Pro 550	Pro	Gly	Pro	His	Ser 555	Ala	Lys	Thr	Glu	Glu 560
His	Thr	Thr	Ile	Leu 565	Arg	Pro	Ser	Tyr	Thr 570	Gly	Leu	Ser	Ser	Ser 575	Ser
Ala	Arg	Phe	Leu 580	Ser	Arg	Ser	Ile	Pro 585	Ser	Leu	Gln	Ser	Glu 590	Tyr	Val
His	Tyr														

<210> SEQ ID NO 804 <211> LENGTH: 2646 <212> TYPE: DNA <213> ORGANISM: Homo sapiens <220> FEATURE: <221> NAME/KEY: misc_feature <222> LOCATION: 2300

<pre><223> OTHER INFORMATION: n = A,T,C or G</pre>	
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gaatatgaga acttctacgc agattttgga ccactcaatc tggcaatggt ttacagatat	300
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tttactggct ctgatcagag aaaacaagca aatgctgcct tccttgttgg atgctacatg	420
gttatatatt tggggagaac cccagaagaa gcatatagaa tattaatctt tggagagaca	480
tcctatattc ctttcagaga tgctgcctat ggaagttgca atttctacat tacacttctt	540
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genageegee coefficient focuseagys actionaged tryingly coefficient	2100

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<210> SEQ ID NO 805

<211> LENGTH: 459

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 805

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Ser Ser Thr Gln Gln Asp Pro Arg Arg Arg Asp Pro Gln Asp Asp Val 35 40 45

Tyr Leu Asp Ile Thr Asp Arg Leu Cys Phe Ala Ile Leu Tyr Ser Arg 50 60

Pro Lys Ser Ala Ser Asn Val His Tyr Phe Ser Ile Asp Asn Glu Leu 65 70 75 80

Glu Tyr Glu Asn Phe Tyr Ala Asp Phe Gly Pro Leu Asn Leu Ala Met 85 90 95

Val Tyr Arg Tyr Cys Cys Lys Ile Asn Lys Lys Leu Lys Ser Ile Thr 100 105 110

Met Leu Arg Lys Lys Ile Val His Phe Thr Gly Ser Asp Gln Arg Lys 115 120 125

Gln Ala Asn Ala Ala Phe Leu Val Gly Cys Tyr Met Val Ile Tyr Leu 130 135 140

Gly Arg Thr Pro Glu Glu Ala Tyr Arg Ile Leu Ile Phe Gly Glu Thr 145 150155160

Ser Tyr Ile Pro Phe Arg Asp Ala Ala Tyr Gly Ser Cys Asn Phe Tyr 165 170 175

Ile Thr Leu Leu Asp Cys Phe His Ala Val Lys Lys Ala Met Gln Tyr 180 \$180\$

Gly Phe Leu Asn Phe Asn Ser Phe Asn Leu Asp Glu Tyr Glu His Tyr 195 200200205

Glu Lys Ala Glu Asn Gly Asp Leu Asn Trp Ile Ile Pro Asp Arg Phe 210 215 220

Ile Ala Phe Cys Gly Pro His Ser Arg Ala Arg Leu Glu Ser Gly Tyr 225 230 235 240

His Gln His Ser Pro Glu Thr Tyr Ile Gln Tyr Phe Lys Asn His Asn

Val Thr Thr Ile Ile Arg Leu Asn Lys Arg Met Tyr Asp Ala Lys Arg 260 265 270

Phe Thr Asp Ala Gly Phe Asp His His Asp Leu Phe Phe Ala Asp Gly 285 Ser Thr Pro Thr Asp Ala Ile Val Lys Glu Phe Leu Asp Ile Cys Glu 295 Asn Ala Glu Gly Ala Ile Ala Val His Cys Lys Ala Gly Leu Gly Arg 305 310 320 Thr Gly Thr Leu Ile Ala Cys Tyr Ile Met Lys His Tyr Arg Met Thr 325 335 Ala Ala Glu Thr Ile Ala Trp Val Arg Ile Cys Arg Pro Gly Ser Val 340 340 Ile Gly Pro Gln Gln Gln Phe Leu Val Met Lys Gln Thr Asn Leu Trp 355 360 Leu Glu Gly Asp Tyr Phe Arg Gln Lys Leu Lys Gly Gln Glu Asn Gly 370 Gln His Arg Ala Ala Phe Ser Lys Leu Leu Ser Gly Val Asp Asp Ile 395 400 Ser Ile Asn Gly Val Glu Asn Gln Asp Gln Gln Glu Pro Glu Pro Tyr 405 405 Ala Leu Lys Ser Arg Arg Gln Ser Lys Thr Ala Ile Pro Leu Thr 435 Ala Leu Lys Ser Arg Thr Lys Thr Val Leu Arg 445 Leu Ser Ile Ser Arg Thr Lys Thr Val Leu Arg 450 -210 SEQ ID NO 806 -211																		
Asn Ala Glu Gly Ala Ile Ala Val His Cys Lys Ala Gly Leu Gly Arg 305 310 310 320 Thr Gly Thr Leu Ile Ala Cys Tyr Ile Met Lys His Tyr Arg Met Thr 325 335 335 335 335 335 335 336 336 336 33	Phe	Thr		Ala	Gly	Phe	Asp		His	Asp	Leu	Phe	Phe 285	Ala	Asp	Gly		
Thr Gly Thr Leu Ile Ala Cys Tyr Ile Met Lys His Tyr Arg Met Thr 325 Ala Ala Glu Thr Ile Ala Trp Val Arg Ile Cys Arg Pro Gly Ser Val 340 Ile Gly Pro Gln Gln Gln Phe Leu Val Met Lys Gln Thr Asn Leu Trp 355 Leu Glu Gly Asp Tyr Phe Arg Gln Lys Leu Lys Gly Gln Glu Asn Gly 370 Gln His Arg Ala Ala Phe Ser Lys Leu Leu Ser Gly Val Asp Asp Ile 385 Ser Ile Asn Gly Val Glu Asn Gln Asp Gln Gln Glu Pro Glu Pro Tyr 405 Ser Asp Asp Asp Glu Ile Asn Gly Val Thr Gln Gly Asp Arg Leu Arg 420 Ala Leu Lys Ser Arg Arg Gln Ser Lys Thr Asn Ala Ile Pro Leu Thr 435 Leu Ser Ile Ser Arg Thr Lys Thr Val Leu Arg 455 **C210> SEQ ID NO 806 **211> LENGTH: 3415 **212> TYPFE: DNA 213> Asp Glogadgag aagcacggt gctgtgtgt ctgaaccgc ggggccdgg	Ser		Pro	Thr	Asp	Ala		Val	Lys	Glu	Phe		Asp	Ile	Cys	Glu		
Ala Ala Glu Thr Ile Ala Trp Val Arg Ile Cys Arg Pro Gly Ser Val 340 340 345 345 345 350 Ile Gly Pro Gln Gln Gln Phe Leu Val Met Lys Gln Thr Asn Leu Trp 355 365 360 360 360 365 365 365 365 365 360 365 365 365 360 365 365 365 360 365 365 360 365 365 360 365 360 365 360 365 360 365 360 365 360 365 360 365 360 365 360 365 360 365 360 365 360 360 365 360 360 360 360 360 360 360 360 360 360		Ala	Glu	Gly	Ala		Ala	Val	His	Cys		Ala	Gly	Leu	Gly			
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Leu Glu Gly Asp Tyr Phe Arg Gln Lys Leu Lys Gly Gln Glu Asn Gly 370 Gln His Arg Ala Ala Phe Ser Lys Leu Leu Ser Gly Val Asp Asp Ile 385 Ser Ile Asn Gly Val Glu Asn Gln Asp Gln Glu Pro Glu Pro Tyr 405 Ser Asp Asp Asp Glu Ile Asn Gly Val Thr Gln Gly Asp Arg Leu Arg 425 Ala Leu Lys Ser Arg Arg Gln Ser Lys Thr Asn Ala Ile Pro Leu Thr 435 Leu Ser Ile Ser Arg Thr Lys Thr Val Leu Arg 455 <a href="#</td><td>Ala</td><td>Ala</td><td>Glu</td><td></td><td>Ile</td><td>Ala</td><td>Trp</td><td>Val</td><td></td><td>Ile</td><td>Cys</td><td>Arg</td><td>Pro</td><td></td><td>Ser</td><td>Val</td><td></td><td></td></tr><tr><td>Gln His Arg Ala Ala Phe Ser Lys Leu Leu Ser Gly Val Asp Asp Ile 385 390 395 400 Ser Ile Asn Gly Val Glu Asn Gln Asp Gln Gln Glu Pro Glu Pro Tyr 405 410 415 Ser Asp Asp Asp Asp Glu Ile Asn Gly Val Thr Gln Gly Asp Arg Leu Arg 425 430 Ala Leu Lys Ser Arg Arg Gln Ser Lys Thr Asn Ala Ile Pro Leu Thr 435 440 455 Leu Ser Ile Ser Arg Thr Lys Thr Val Leu Arg 450 455 <td>Ile</td><td>Gly</td><td></td><td>Gln</td><td>Gln</td><td>Gln</td><td>Phe</td><td></td><td>Val</td><td>Met</td><td>Lys</td><td>Gln</td><td></td><td>Asn</td><td>Leu</td><td>Trp</td><td></td><td></td>	Ile	Gly		Gln	Gln	Gln	Phe		Val	Met	Lys	Gln		Asn	Leu	Trp		
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Met Asp Gln Leu Gln Gly Leu Gly Ser Asp Tyr Glu Gln Pro Leu Glu 50 55 60

Val Lys Asn Asn Ser Asn Leu Gln Arg Met Gly Ser Ser Glu Ser Thr 65 70 75 80

Asp Ser Gly Phe Cys Leu Asp Ser Pro Gly Pro Leu Asp Ser Lys Glu 85 90 95

Asn Leu Glu Asn Pro Met Arg Arg Ile His Ser Leu Pro Gln Lys Leu 100 105 110

Leu Gly Cys Ser Pro Ala Leu Lys Arg Ser His Ser Asp Ser Leu Asp

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Leu Leu Gly Ser His Gly Leu Leu Gly Ser Pro Val Arg Ala Ala Ala 35 4045

Ser Ser Pro Val Thr Thr Leu Thr Gln Thr Met His Asp Leu Ala Gly 50 60

Leu Gly Ser Arg Ser Arg Leu Thr His Leu Ser Leu Ser Arg Arg Ala 65 70 75 80

Ser Glu Ser Ser Leu Ser Ser Glu Ser Ser Glu Ser Ser Asp Ala Gly 85 90 95

Leu Cys Met Asp Ser Pro Ser Pro Met Asp Pro His Met Ala Glu Gln $100 \hspace{1.5cm} 105 \hspace{1.5cm} 105 \hspace{1.5cm} 110 \hspace{1.5cm}$

Thr Phe Glu Gln Ala Ile Gln Ala Ala Ser Arg Ile Ile Arg As
n Glu 115 120 125

Gln Phe Ala Ile Arg Arg Phe Gln Ser Met Pro Val Arg Leu Leu Gly 130 135 140

His Ser Pro Val Leu Arg Asn Ile Thr Asn Ser Gln Ala Pro Asp Gly 145 150 155 160

Arg Arg Lys Ser Glu Ala Gly Ser Gly Ala Ala Ser Ser Gly Glu 165 170 175

Asp Lys Glu Asn Asp Gly Phe Val Phe Lys Met Pro Trp Lys Pro Thr 180 \$180\$

His Pro Ser Ser Thr His Ala Leu Ala Glu Trp Ala Ser Arg Arg Glu

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Ala	Phe 210	Ala	Gln	Arg	Pro	Ser 215	Ser	Ala	Pro	Asp	Leu 220	Met	Cys	Leu	Ser
Pro 225	Asp	Arg	Lys	Met	Glu 230	Val	Glu	Glu	Leu	Ser 235	Pro	Leu	Ala	Leu	Gly 240
Arg	Phe	Ser	Leu	Thr 245	Pro	Ala	Glu	Gly	Asp 250	Thr	Glu	Glu	Asp	Asp 255	Gly
Phe	Val	Asp	Ile 260	Leu	Glu	Ser	Asp	Leu 265	Lys	Asp	Asp	Asp	Ala 270	Val	Pro
Pro	Gly	Met 275	Glu	Ser	Leu	Ile	Ser 280	Ala	Pro	Leu	Val	Lys 285	Thr	Leu	Glu
Lys	Glu 290	Glu	Glu	Lys	Asp	Leu 295	Val	Met	Tyr	Ser	Lys 300	Cys	Gln	Arg	Leu
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Arg	Leu	Glu	Arg	Pro 325	Gln	Asp	Arg	Asp	Thr 330	Pro	Val	Gln	Asn	Lys 335	Arg
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Ala 385	Phe	Leu	Leu	Gln	Thr 390	Val	Asp	Gly	Lys	His 395	Gln	Asp	Leu	Lys	Tyr 400
Ile	Ser	Pro	Glu	Thr 405	Met	Val	Ala	Leu	Leu 410	Thr	Gly	Lys	Phe	Ser 415	Asn
Ile	Val	Asp	Lys 420	Phe	Val	Ile	Val	Asp 425	Суѕ	Arg	Tyr	Pro	Tyr 430	Glu	Tyr
Glu	Gly	Gly 435	His	Ile	Lys	Thr	Ala 440	Val	Asn	Leu	Pro	Leu 445	Glu	Arg	Asp
Ala	Glu 450	Ser	Phe	Leu	Leu	Lys 455	Ser	Pro	Ile	Ala	Pro 460	Cys	Ser	Leu	Asp
Lys 465	Arg	Val	Ile	Leu	Ile 470	Phe	His	Cys	Glu	Phe 475	Ser	Ser	Glu	Arg	Gly 480
Pro	Arg	Met	Сув	Arg 485	Phe	Ile	Arg	Glu	Arg 490	Asp	Arg	Ala	Val	Asn 495	Asp
Tyr	Pro	Ser		Tyr	Tyr	Pro	Glu		Tyr			Lys	Gly 510		Tyr
Lys	Glu	Phe 515	Phe	Pro	Gln	His	Pro 520	Asn	Phe	Сув	Glu	Pro 525	Gln	Asp	Tyr
Arg	Pro 530	Met	Asn	His	Glu	Ala 535	Phe	Lys	Asp	Glu	Leu 540	Lys	Thr	Phe	Arg
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Gly Ser Pro Gly Phe Phe Arg Thr Ser Gly Ser Ala Phe Ser Trp Asp 75 80	
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<400> SEQUENCE: 817

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1860

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<210> SEQ ID NO 821

<211> LENGTH: 432

<212> TYPE: PRT

<213> ORGANISM: Rattus norvegicus

<400> SEQUENCE: 821

Met Glu Met Glu Lys Glu Phe Glu Gln Ile Asp Lys Ala Gly Asn Trp 1 $$ 10 $$ 15

Ala Ala Ile Tyr Gln Asp Ile Arg His Glu Ala Ser Asp Phe Pro Cys $20 \hspace{1cm} 25 \hspace{1cm} 30$

Arg Ile Ala Lys Leu Pro Lys Asn Lys Asn Arg Asn Arg Tyr Arg Asp $35 \hspace{1cm} 40 \hspace{1cm} 45$

Val Ser Pro Phe Asp His Ser Arg Ile Lys Leu His Gln Glu Asp Asn 50 60

Asp Tyr Ile Asn Ala Ser Leu Ile Lys Met Glu Glu Ala Gln Arg Ser 65 70 75 80

Tyr Ile Leu Thr Gln Gly Pro Leu Pro Asn Thr Cys Gly His Phe Trp 85 90 95

Glu Met Val Trp Glu Gln Lys Ser Arg Gly Val Val Met Leu Asn Arg 100 105 110

Ile Met Glu Lys Gly Ser Leu Lys Cys Ala Gln Tyr Trp Pro Gln Lys $115 \hspace{1.5cm} 120 \hspace{1.5cm} 125 \hspace{1.5cm}$

Glu Glu Lys Glu Met Val Phe Asp Asp Thr Asn Leu Lys Leu Thr Leu 130 140

Ile Ser Glu Asp Val Lys Ser Tyr Tyr Thr Val Arg Gln Leu Glu Leu 145 $$ 150 $$ 155 $$ 160

Glu Asn Leu Ala Thr Glu Glu Ala Arg Glu Ile Leu His Phe His Tyr $165 \hspace{1cm} 170 \hspace{1cm} 175$

Thr Trp Pro Asp Phe Gly Val Pro Glu Ser Pro Ala Ser Phe Leu
180 185 190

Asn Phe Leu Phe Lys Val Arg Glu Ser Gly Ser Leu Ser Pro Glu His

Phe 225	Cys	Leu	Ala	Asp	Thr 230	Сув	Leu	Leu	Leu	Met 235	Asp	Lys	Arg	Lys	Asp 240
Pro	Ser	Ser	Val	Asp 245	Ile	Lys	Lys	Val	Leu 250	Leu	Glu	Met	Arg	A rg 255	Phe
Arg	Met	Gly	Leu 260	Ile	Gln	Thr	Ala	Asp 265	Gln	Leu	Arg	Phe	Ser 270	Tyr	Leu
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Asp	Gln 290	Trp	Lys	Glu	Leu	Ser 295	His	Glu	Asp	Leu	Glu 300	Pro	Pro	Pro	Glu
His 305	Val	Pro	Pro	Pro	Pro 310	Arg	Pro	Pro	Lys	Arg 315	Thr	Leu	Glu	Pro	His 320
Asn	Gly	Lys	Сув	Lys 325	Glu	Leu	Phe	Ser	Asn 330	His	Gln	Trp	Val	Ser 335	Glu
Glu	Ser	Сув	Glu 340	Asp	Glu	Asp	Ile	Leu 345	Ala	Arg	Glu	Glu	Ser 350	Arg	Ala
Pro	Ser	Ile 355	Ala	Val	His	Ser	Met 360	Ser	Ser	Met	Ser	Gln 365	Asp	Thr	Glu
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Val 385	Pro	Thr	Glu	Glu	Glu 390	Leu	Ser	Pro	Thr	Glu 395	Glu	Glu	Gln	Lys	Ala 400
His	Arg	Pro	Val	His 405	Trp	Lys	Pro	Phe	Leu 410	Val	Asn	Val	Сув	Met 415	Ala
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<210> SEQ ID NO 822

<211> LENGTH: 2287

<212> TYPE: DNA <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 822

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<210> SEQ ID NO 823
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<211> LENGTH: 415

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 823

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Pro His Arg Val Ala Lys Phe Pro Glu Asn Arg Asn Arg Asn Arg Tyr 35 40 45

Arg Asp Val Ser Pro Tyr Asp His Ser Arg Val Lys Leu Gln Asn Ala 50 60

Glu Asn Asp Tyr Ile Asn Ala Ser Leu Val Asp Ile Glu Glu Ala Gln 65 70 75 80

Arg Ser Tyr Ile Leu Thr Gln Gly Pro Leu Pro Asn Thr Cys Cys His $85 \hspace{1cm} 90 \hspace{1cm} 95$

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Asn	Arg	Ile 115	Val	Glu	Lys	Glu	Ser 120	Val	Lys	Сув	Ala	Gln 125	Tyr	Trp	Pro
Thr	Asp 130	Asp	Gln	Glu	Met	Leu 135	Phe	Lys	Glu	Thr	Gly 140	Phe	Ser	Val	Lys
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Tyr	Thr	Thr	Trp 180	Pro	Asp	Phe	Gly	Val 185	Pro	Glu	Ser	Pro	Ala 190	Ser	Phe
Leu	Asn	Phe 195	Leu	Phe	Lys	Val	Arg 200	Glu	Ser	Gly	Ser	Leu 205	Asn	Pro	Asp
His	Gly 210	Pro	Ala	Val	Ile	His 215	Cys	Ser	Ala	Gly	Ile 220	Gly	Arg	Ser	Gly
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Gly	Leu	Ile	Gln 260	Thr	Pro	Asp	Gln	Leu 265	Arg	Phe	Ser	Tyr	Met 270	Ala	Ile
Ile	Glu	Gly 275	Ala	Lys	Сув	Ile	Lys 280	Gly	Asp	Ser	Ser	Ile 285	Gln	Lys	Arg
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Trp 385	Gln	Pro	Ile	Leu	Thr 390	Lys	Met	Gly	Phe	Met 395	Ser	Val	Ile	Leu	Val 400
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Thr	Asp 130	Asp	Gln	Glu	Met	Leu 135	Phe	Lys	Glu	Thr	Gly 140	Phe	Ser	Val	Lys
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Leu	Glu	Asn	Ile	Asn 165	Ser	Gly	Glu	Thr	Arg 170	Thr	Ile	Ser	His	Phe 175	His
Tyr	Thr	Thr	Trp 180	Pro	Asp	Phe	Gly	Val 185	Pro	Glu	Ser	Pro	Ala 190	Ser	Phe
Leu	Asn	Phe 195	Leu	Phe	Lys	Val	Arg 200	Glu	Ser	Gly	Ser	Leu 205	Asn	Pro	Asp
His	Gly 210	Pro	Ala	Val	Ile	His 215	Cys	Ser	Ala	Gly	Ile 220	Gly	Arg	Ser	Gly
Thr 225	Phe	Ser	Leu	Val	Asp 230	Thr	Cys	Leu	Val	Leu 235	Met	Glu	Lys	Gly	Asp 240
Asp	Ile	Asn	Ile	L y s 245	Gln	Val	Leu	Leu	Asn 250	Met	Arg	Lys	Tyr	Arg 255	Met
Gly	Leu	Ile	Gln 260	Thr	Pro	Asp	Gln	Leu 265	Arg	Phe	Ser	Tyr	Met 270	Ala	Ile
Ile	Glu	Gly 275	Ala	Lys	Cys	Ile	L y s 280	Gly	Asp	Ser	Ser	Ile 285	Gln	Lys	Arg
Trp	L y s 290	Glu	Leu	Ser	Lys	Glu 295	Asp	Leu	Ser	Pro	Ala 300	Phe	Asp	His	Ser
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Ile	Arg	Glu 355	Asp	Arg	Lys	Ala	Thr 360	Thr	Ala	Gln	Lys	Val 365	Gln	Gln	Met

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1680 1714

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Pro	His	Arg 35	Val	Ala	Lys	Phe	Pro 40	Glu	Asn	Arg	Asn	Arg 45	Asn	Arg	Tyr
Arg	Asp 50	Val	Ser	Pro	Tyr	Asp 55	His	Ser	Arg	Val	Lys 60	Leu	Gln	Asn	Ala
Glu 65	Asn	Asp	Tyr	Ile	Asn 70	Ala	Ser	Leu	Val	Asp 75	Ile	Glu	Glu	Ala	Gln 80
Arg	Ser	Tyr	Ile	Leu 85	Thr	Gln	Gly	Pro	Leu 90	Pro	Asn	Thr	Сув	С у в 95	His
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Thr	Asp 130	Asp	Gln	Glu	Met	Leu 135	Phe	Lys	Glu	Thr	Gly 140	Phe	Ser	Val	Lys
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Leu	Asn	Phe 195	Leu	Phe	Lys	Val	Arg 200	Glu	Ser	Gly	Ser	Leu 205	Asn	Pro	Asp
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Thr 225	Phe	Ser	Leu	Val	Asp 230	Thr	Сув	Leu	Val	Leu 235	Met	Glu	Lys	Gly	Asp 240
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Trp	Lys 290	Glu	Leu	Ser	Lys	Glu 295	Asp	Leu	Ser	Pro	Ala 300	Phe	Asp	His	Ser
Pro 305	Asn	Lys	Ile	Met	Thr 310	Glu	Lys	Tyr	Asn	Gly 315	Asn	Arg	Ile	Gly	Leu 320
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Met	Gln	Asp	Thr 340	Met	Glu	Glu	Asn	Ser 345	Glu	Ser	Ala	Leu	Arg 350	Lys	Arg
Ile	Arg	Glu 355	Asp	Arg	Lys	Ala	Thr 360	Thr	Ala	Gln	Lys	Val 365	Gln	Gln	Met

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<210> SEQ ID NO 828 <211> LENGTH: 1555

<211> LENGTH: 1555 <212> TYPE: DNA

<213> ORGANISM: Mus musculus

<400> SEQUENCE: 828

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<210> SEQ ID NO 829

<211> LENGTH: 382

<212> TYPE: PRT

<213> ORGANISM: Mus musculus

<400> SEOUENCE: 829

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Pro	His	Arg 35	Val	Ala	Lys	Phe	Pro 40	Glu	Asn	Arg	Asn	Arg 45	Asn	Arg	Tyr
Arg	Asp 50	Val	Ser	Pro	Tyr	Asp 55	His	Ser	Arg	Val	Lys 60	Leu	Gln	Ser	Thr
Glu 65	Asn	Asp	Tyr	Ile	Asn 70	Ala	Ser	Leu	Val	Asp 75	Ile	Glu	Glu	Ala	Gln 80
Arg	Ser	Tyr	Ile	Leu 85	Thr	Gln	Gly	Pro	Leu 90	Pro	Asn	Thr	Сув	С у в 95	His
Phe	Trp	Leu	Met 100	Val	Trp	Gln	Gln	Lys 105	Thr	Lys	Ala	Val	Val 110	Met	Leu
Asn	Arg	Thr 115	Val	Glu	Lys	Glu	Ser 120	Val	Lys	Cys	Ala	Gln 125	Tyr	Trp	Pro
Thr	Asp 130	Asp	Arg	Glu	Met	Val 135	Phe	Lys	Glu	Thr	Gly 140	Phe	Ser	Val	Lys
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Leu	Glu	Asn	Ile	Asn 165	Thr	Gly	Glu	Thr	Arg 170	Thr	Ile	Ser	His	Phe 175	His
Tyr	Thr	Thr	Trp 180	Pro	Asp	Phe	Gly	Val 185	Pro	Glu	Ser	Pro	Ala 190	Ser	Phe
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His	Gly 210	Pro	Ala	Val	Ile	His 215	Cys	Ser	Ala	Gly	Ile 220	Gly	Arg	Ser	Gly
Thr 225	Phe	Ser	Leu	Val	Asp 230	Thr	Cys	Leu	Val	Leu 235	Met	Glu	Lys	Gly	Glu 240
Asp	Val	Asn	Val	Lys 245	Gln	Leu	Leu	Leu	Asn 250	Met	Arg	Lys	Tyr	A rg 255	Met
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Ile	Glu	Gly 275	Ala	Lys	Tyr	Thr	Lys 280	Gly	Asp	Ser	Asn	Ile 285	Gln	Lys	Arg
Trp	Lys 290	Glu	Leu	Ser	Lys	Glu 295	Asp	Leu	Ser	Pro	Ile 300	Cys	Asp	His	Ser
Gln 305	Asn	Arg	Val	Met	Val 310	Glu	Lys	Tyr	Asn	Gly 315	Lys	Arg	Ile	Gly	Ser 320
Glu	Asp	Glu	Lys	Leu 325	Thr	Gly	Leu	Pro	Ser 330	Lys	Val	Gln	Asp	Thr 335	Val
Glu	Glu	Ser	Ser 340	Glu	Ser	Ile	Leu	Arg 345	Lys	Arg	Ile	Arg	Glu 350	Asp	Arg
Lys	Ala	Thr 355	Thr	Ala	Gln	Lys	Val 360	Gln	Gln	Met	Lys	Gln 365	Arg	Leu	Asn
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<210> SEQ ID NO 831
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<211> LENGTH: 181

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 831

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Thr Tyr Leu Arg Ile Pro Val Ala Asp Thr Pro Glu Val Pro Ile Lys Lys His Phe Lys Glu Cys Ile Asn Phe Ile His Cys Cys Arg Leu Asn 65 70 75 80 Gly Gly Asn Cys Leu Val His Cys Phe Ala Gly Ile Ser Arg Ser Thr $85 \hspace{1cm} 90 \hspace{1cm} 95$ Thr Ile Val Thr Ala Tyr Val Met Thr Val Thr Gly Leu Gly Trp Arg $100 \hspace{1.5cm} 105 \hspace{1.5cm} 105$ Asp Val Leu Glu Ala Ile Lys Ala Thr Arg Pro Ile Ala Asn Pro Asn Pro Gly Phe Arg Gln Gln Leu Glu Glu Phe Gly Trp Ala Ser Ser Gln Lys Gly Ala Arg His Arg Thr Ser Lys Thr Ser Gly Ala Gln Cys Pro Pro Met Thr Ser Ala Thr Trp Met Val Thr Gly Pro Lys Val Pro Asp Leu Ser Val Leu Arg <210> SEO ID NO 832 <211> LENGTH: 1807 <212> TYPE: DNA <213> ORGANISM: Homo sapiens <400> SEQUENCE: 832 ggccccccgt tccccgccag gctgcaggcg tcgggcctgg gccgtcaggg cagctgtgac 60 cggatcgctt cccgggcggc gagctggggg tgcacccgga ccgccgcccc cgggatcatg ggcaatggca tgaccaaggt acttcctgga ctctacctcg gaaacttcat tgatgccaaa gacctggatc agctgggccg aaataagatc acacacatca tctctatcca tgagtcaccc 240 cagectetge tgcaggatat cacetacett egcatecegg tegetgatae eeetgaggta 300 cccatcaaaa agcacttcaa agaatgtatc aacttcatcc actgctgccg ccttaatggg 360 gggaactgcc ttgtgcactg ctttgcaggc atctctcgca gcaccacgat tgtgacagcg 420 tatqtqatqa ctqtqacqqq qctaqqctqq cqqqacqtqc ttqaaqccat caaqqccacc aggcccatcg ccaaccccaa cccaggcttt aggcagcagc ttgaagagtt tggctgggcc 600 agttcccaga agggtgccag acataggacc tcaaaaacct ctggtgccca atgccctccg atgacttcag caacctgcct gctggctgca cgtgtggctc ttctctccgc agcgctggtg 660 cgcgaagcca ccgggcgcac agcccagcgc tgtcgtctga gtccgcgggc ggccgccgag 720 780 cgcctgctgg ggccgccacc tcacgttgca gcaggatggt caccggaccc aaagtaccag atetytetyt getteggtga ggaggaeeeg ggeeeeacae ageaeeecaa ggageagete 840 atcatggegg acgtgcaggt gcagcttegg cetgggaget egteetgeac tetaagtgee tcaaccgagc gcccagatgg gtcctcaacc cctggcaacc ccgatggcat cactcacctt caatgcagct gcctccatcc taagcgagcc gcttcctctt cttgtacccg ctgaaggcag 1020

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His Ile Ile Ser Ile His Glu Ser Pro Gln Pro Leu Leu Gln Asp Ile 35 4045

Thr Tyr Leu Arg Ile Pro Val Ala Asp Thr Pro Glu Val Pro Ile Lys 50 60

Lys His Phe Lys Glu Cys Ile Asn Phe Ile His Cys Cys Arg Leu Asn 65 70 75 80

Gly Gly Asn Cys Leu Val His Cys Phe Ala Gly Ile Ser Arg Ser Thr 85 90 95

Asp Val Leu Glu Ala Ile Lys Ala Thr Arg Pro Ile Ala Asn Pro Asn 115 120 125

Pro Gly Phe Arg Gln Gln Leu Glu Glu Phe Gly Trp Ala Ser Ser Gln 130 135 140

Lys Gly Ala Arg His Arg Thr Ser Lys Thr Ser Gly Ala Gln Cys Pro 145 150 155 160

Pro Met Thr Ser Ala Thr Cys Leu Leu Ala Ala Arg Val Ala Leu Leu 165 \$170\$

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Cys Phe Gly Glu Glu Asp Pro Gly Pro Thr Gln His Pro Lys Glu Gln 225 235 240

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What is claimed is:

- 1. An isolated small interfering RNA (siRNA) polynucleotide, comprising at least one nucleotide sequence selected from the group consisting of SEQ ID NOS:4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493.
- 2. The small interfering RNA polynucleotide of claim 1 that comprises at least one nucleotide sequence selected
- from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493 and the complementary polynucleotide thereto.
- 3. A small interfering RNA polynucleotide of either claim 1 or claim 2 that is capable of interfering with expression of a polypeptide, which polypeptide comprises an amino acid sequence as set forth in a sequence selected from the group

- consisting of SEQ ID NO: 779, SEQ ID NO:789, SEQ ID NO:791, SEQ ID NO:797, SEQ ID NO:809, SEQ ID NO:801, SEQ ID NO:803, SEQ ID NO:805, SEQ ID NO:807, SEQ ID NO:809, SEQ ID NO:811, and SEQ ID NO:813.
- 4. The siRNA polynucleotide of either claim 1 or claim 2 wherein the nucleotide sequence of the siRNA polynucleotide differs by one, two, three or four nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493.
- 5. The siRNA polynucleotide of either claim 1 or claim 2 wherein the nucleotide sequence of the siRNA polynucleotide differs by at least two, three or four nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493.
- 6. An isolated siRNA polynucleotide comprising a nucleotide sequence according to SEQ ID NO: 4, or the complement thereof.
- 7. An isolated siRNA polynucleotide comprising a nucleotide sequence selected from the group consisting of SEQ ID NOS: 100 and 105, or the complement thereof.
- **8**. An isolated siRNA polynucleotide comprising a nucleotide sequence selected from the group consisting of SEQ ID NOS: 120, 125, and 130, or the complement thereof.
- 9. An isolated siRNA polynucleotide comprising a nucleotide sequence selected from the group consisting of SEQ ID NOS: 140, 145, and 150, or the complement thereof.
- 10. An isolated siRNA polynucleotide comprising a nucleotide sequence selected from the group consisting of SEQ ID NOS: 440 and 445, or the complement thereof.
- 11. An isolated siRNA polynucleotide comprising a nucleotide sequence selected from the group consisting of SEQ ID NOS: 455 and 460, or the complement thereof.
- 12. An isolated siRNA polynucleotide comprising a nucleotide sequence according to SEQ ID NO: 465, or the complement thereof.
- 13. An isolated siRNA polynucleotide comprising a nucleotide sequence selected from the group consisting of SEQ ID NOS: 470 and 475, or the complement thereof.
- 14. An isolated siRNA polynucleotide comprising a nucleotide sequence selected from the group consisting of SEQ ID NOS: 480, 485, and 490, or the complement thereof.
- 15. The siRNA polynucleotide of claim 1 or claim 2 wherein the polynucleotide comprises at least one synthetic nucleotide analogue of a naturally occurring nucleotide.
- **16.** The siRNA polynucleotide of claim 1 or claim 2 wherein the polynucleotide is linked to a detectable label.
- 17. The siRNA polynucleotide of claim 16 wherein the detectable label is a reporter molecule.
- 18. The siRNA of claim 17 wherein the reporter molecule is selected from the group consisting of a dye, a radionuclide, a luminescent group, a fluorescent group, and biotin.
- 19. The siRNA polynucleotide of claim 18 wherein the fluorescent group is fluorescein isothiocyanate.
- 20. The siRNA polynucleotide of claim 16 wherein the detectable label is a magnetic particle.

- 21. A pharmaceutical composition comprising the siRNA polynucleotide of either claim 1 or claim 2 and a physiologically acceptable carrier.
- **22**. The pharmaceutical composition of claim 22 wherein the carrier comprises a liposome.
- 23. A recombinant nucleic acid construct comprising a polynucleotide that is capable of directing transcription of a small interfering RNA (siRNA), the polynucleotide comprising:
 - (i) a first promoter; (ii) a second promoter; and (iii) at least one DNA polynucleotide segment comprising at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493, or a complement thereto, wherein each DNA polynucleotide segment and its complement are operably linked to at least one of the first and second promoters, and wherein the promoters are oriented to direct transcription of the DNA polynucleotide segment and its reverse complement.
- 24. The recombinant nucleic acid construct of claim 23, comprising at least one enhancer that is selected from a first enhancer operably linked to the first promoter and a second enhancer operably linked to the second promoter.
- 25. The recombinant nucleic acid construct of claim 23, comprising at least one transcriptional terminator that is selected from (i) a first transcriptional terminator that is positioned in the construct to terminate transcription directed by the first promoter and (ii) a second transcriptional terminator that is positioned in the construct to terminate transcription directed by the second promoter.
- 26. The recombinant nucleic acid construct of claim 24 wherein the siRNA is capable of interfering with expression of a polypeptide, wherein the polypeptide comprises an amino acid sequence as set forth in a sequence selected from the group consisting of SEQ ID NO: 779, SEQ ID NO:789, SEQ ID NO:791, SEQ ID NO:797, SEQ ID NO:799, SEQ ID NO:801, SEQ ID NO:803, SEQ ID NO:805, SEQ ID NO:807, SEQ ID NO:809, SEQ ID NO:811, and SEQ ID NO:813.
- 27. A recombinant nucleic acid construct comprising a polynucleotide that is capable of directing transcription of a small interfering RNA (siRNA), the polynucleotide comprising at least one promoter and a DNA polynucleotide segment, wherein the DNA polynucleotide segment is operably linked to the promoter, and wherein the DNA polynucleotide segment comprises (i) at least one DNA polynucleotide that comprises at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493, or a complement thereto; (ii) a spacer sequence comprising at least 4 nucleotides operably linked to the DNA polynucleotide of (i); and (iii) the reverse complement of the DNA polynucleotide of (i) operably linked to the spacer sequence.
- 28. The recombinant nucleic acid construct of claim 27 wherein the siRNA comprises an overhang of at least one and no more than four nucleotides, the overhang being located immediately 3' to (iii).

- 29. The recombinant nucleic acid construct of claim 27 wherein the spacer sequence comprises at least 9 nucleotides.
- **30**. The recombinant nucleic acid construct of claim 27 wherein the spacer sequence comprises two uridine nucleotides that are contiguous with (iii).
- 31. The recombinant nucleic acid construct of claim 27 comprising at least one transcriptional terminator that is operably linked to the DNA polynucleotide segment.
- 32. A host cell transformed or transfected with the recombinant nucleic acid construct of any one of claims 23-31.
- 33. A pharmaceutical composition comprising an siRNA polynucleotide and a physiologically acceptable carrier, wherein the siRNA polynucleotide is selected from the group consisting of:
 - (i) an RNA polynucleotide which comprises at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493,
 - (ii) an RNA polynucleotide that comprises at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493 and the complementary polynucleotide thereto,
 - (iii) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by one, two or three nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493, and
 - (iv) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by two, three or four nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493.
- **34**. The pharmaceutical composition of claim 33 wherein the carrier comprises a liposome.
- 35. A method for interfering with expression of a polypeptide, or variant thereof, comprising contacting a subject that comprises at least one cell which is capable of expressing the polypeptide with a siRNA polynucleotide for a time and under conditions sufficient to interfere with expression of the polypeptide, wherein:
 - (a) the polypeptide comprises an amino acid sequence as set forth in a sequence selected from the group consisting of SEQ ID NO: 779, SEQ ID NO:789, SEQ ID NO:791, SEQ ID NO:797, SEQ ID NO:799, SEQ ID NO:801, SEQ ID NO:803, SEQ ID NO:805, SEQ ID NO:807, SEQ ID NO:809, SEQ ID NO:811, and SEQ ID NO:813,

- (b) the siRNA polynucleotide is selected from the group consisting of
 - (i) an RNA polynucleotide which comprises at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493,
 - (ii) an RNA polynucleotide that comprises at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493 and the complementary polynucleotide thereto,
 - (iii) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by one, two or three nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493, and
- (iv) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by two, three or four nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493.
- 36. A method for interfering with expression of a polypeptide that comprises an amino acid sequence as set forth in a sequence selected from the group consisting of SEQ ID NO: 779, SEQ ID NO:789, SEQ ID NO:791, SEQ ID NO:797, SEQ ID NO:799, SEQ ID NO:801, SEQ ID NO:803, SEQ ID NO:805, SEQ ID NO:807, SEQ ID NO:809, SEQ ID NO:811, and SEQ ID NO:813, or a variant of said polypeptide, said method comprising contacting, under conditions and for a time sufficient to interfere with expression of the polypeptide, (i) a subject that comprises at least one cell that is capable of expressing the polypeptide, and (ii) a recombinant nucleic acid construct according to either claim 23 or claim 27.
- **37.** A method for identifying a component of a signal transduction pathway comprising:
 - A. contacting a siRNA polynucleotide and a first biological sample comprising at least one cell that is capable of expressing a target polypeptide, or a variant of said polypeptide, under conditions and for a time sufficient for target polypeptide expression when the siRNA polynucleotide is not present, wherein
 - (1) the target polypeptide comprises an amino acid sequence as set forth in a sequence selected from the group consisting of SEQ ID NO: 779, SEQ ID NO: 789, SEQ ID NO: 791, SEQ ID NO: 797, SEQ ID NO: 799, SEQ ID NO: 801, SEQ ID NO: 803, SEQ ID NO: 805, SEQ ID NO: 807, SEQ ID NO: 809, SEQ ID NO: 811, and SEQ ID NO: 813,

- (2) the siRNA polynucleotide is selected from the group consisting of
 - (i) an RNA polynucleotide which comprises at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493,
 - (ii) an RNA polynucleotide that comprises at least one nucleotide sequence selected from the group consisting of SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493 and the complementary polynucleotide thereto,
 - (iii) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by one, two or three nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458,

- 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493, and
- (iv) an RNA polynucleotide according to (i) or (ii) wherein the nucleotide sequence of the siRNA polynucleotide differs by two, three or four nucleotides at any of positions 1-19 of a sequence selected from the group consisting of the sequences set forth in SEQ ID NOS: 4-7, 100-103, 105-108, 120-123, 125-128, 130-133, 140-143, 145-148, 150-153, 440-443, 445-448, 455-458, 460-463, 465-468, 470-473, 475-478, 480-483, 485-488, and 490-493; and
- B. comparing a level of phosphorylation of at least one protein that is capable of being phosphorylated in the cell with a level of phosphorylation of the protein in a control sample that has not been contacted with the siRNA polynucleotide,
- wherein an altered level of phosphorylation of the protein in the presence of the siRNA polynucleotide relative to the level of phosphorylation of the protein in an absence of the siRNA polynucleotide indicates that the protein is a component of a signal transduction pathway.

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